

A Review on Hepatoprotective and Immunomodulatory Herbal Plants

Uorakkottil Ilyas, Deepshikha P. Katare¹, Vidhu Aeri, Punnooth Poonguzi Naseef²

Department of Pharmacognosy and Phytochemistry, Hamdard University, New Delhi, ¹Department of Pharmaceutical Biotechnology, Amity University, Noida, Uttar Pradesh, ²Department of Pharmaceutics, Jamia Salafiya Pharmacy College, Kerala, India

ABSTRACT

The liver is the most important organ that plays an important role in maintaining various physiological processes in the body. Hepatitis is an inflammation of the liver and is characterized by the presence of inflammatory cells in the tissue of the organ. There are five main viruses, referred to as types A, B, C, D, and E. These five types are of the greatest concern because of the burden of illness and death. Liver injury or liver dysfunction is a major health problem that challenges not only health care professionals but also the drug regulatory agencies and the pharmaceutical industry. Herbal medicines have been used in the treatment of liver disease for a long time. The immune system is the part of body that diagnoses the pathogen by using a specific receptor to reveal immediate response by the activation of immune components cells, chemokines, and cytokines, and also the release of the inflammatory mediator. They potentiate and modulate the immune system. The plant-derived phytoconstituents (polysaccharides, proteins and flavanoids, lignans, rotenoids, etc.) stimulate the immune system and maintained hepatic diseases. There are a number of hepatoprotective and immunomodulatory herbs that have been reported. The present review is aimed at compiling data on promising phytochemicals from hepatoprotective and immunomodulatory herbs.

Key words: Hepatoprotective herb, immunomodulatory herb, nitric oxide

INTRODUCTION

The liver is the most important organ that plays an important role in maintaining various physiological processes in the body. It is involved in several vital functions, such as metabolism, secretion, and storage. It plays a central role in the detoxification and excretion of many exogenous and endogenous compounds. Hence, any injury to it or impairment of its function has grave implications for the health of the affected person. Every year, about 18,000 people are reported to die due to liver cirrhosis caused by hepatitis, although viral infection is one of the main causes for hepatic injury. It acts as a storage depot for proteins, glycogen, various vitamins, and metals. It also has a role in the regulation of blood volume by transferring the blood from the portal to the systemic circulation and its reticulo-endothelial system and participates in the immune mechanism. The human body identifies almost all drugs as foreign substances (i.e., xenobiotics) and subjects them to various chemical processes (such as metabolism) to make them suitable for elimination. This involves chemical transformations to (a) reduce fat solubility and (b) change biological activity. Although almost all tissues in the body have some ability to metabolize chemicals, smooth endoplasmic reticulum in liver is the principal "metabolic clearing house"

for both endogenous chemicals (e.g., cholesterol, steroid hormones, fatty acids, and proteins), and exogenous substances (e.g., drugs). The central role played by the liver in the clearance and transformation of chemicals also makes it susceptible to drug-induced injury.

Hepatitis is an inflammation of the liver and is characterized by the presence of inflammatory cells in the tissue of the organ. There are five main viruses, referred to as types A, B, C, D, and E. These five types are of the greatest concern because of the burden of illness and death. The condition can be self-limiting (healing on its own) or can progress to fibrosis (scarring) and cirrhosis. Hepatitis may occur with limited or no symptoms, but often leads to jaundice, anorexia (poor appetite), and malaise. Hepatitis is acute when it lasts less than 6 months and chronic when it persists for longer. Hepatic trouble, which includes parasites and viral infections; autoimmune diseases; and intoxication with various xenobiotics such as alcohol, herbal medicine, drugs, chlorinated solvents, peroxidized fatty acids, fungal toxins, industrial pollutants, and radioactive isotopes. In particular, types A and C lead to chronic disease in hundreds of millions of people and, together, are the most common cause of liver cirrhosis and cancer.

About 1 million deaths per year are attributed to viral hepatitis infection, that is, hepatitis B virus (HBV) and hepatitis C virus (HCV) taken together, which is the leading cause of liver cirrhosis and cancer, accounting for 78% of cases. Nearly 1 out of every 3 people in the world (approximately 2 million people) has been infected by HBV and HCV. On World Hepatitis Day, July 28, 2013, the World Health Organization (WHO) and its partners focused on the fact that although the burden of

Correspondence:

Dr. Vidhu Aeri,
Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy,
Hamdard University, Hamdard Nagar,
New Delhi - 110 062, India.
E-mail: vdhaeri@yahoo.com

Access this article online

Quick Response Code:



Website:

www.phcogrev.com

DOI:

10.4103/0973-7847.176544

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

Cite this article as: Ilyas U, Katare DP, Aeri V, Naseef PP. A review on hepatoprotective and immunomodulatory herbal plants. *Phcog Rev* 2016;10:66-70.

disease caused by viral hepatitis is growing, it remains largely ignored or unknown to many policymakers, health workers, and the public.

HERBAL HEPATOPROTECTIVE AGENTS

There are generally classified into three categories, as noted below.

Antihepatotoxic agents

These generally antagonize the effects of any hepatotoxins causing hepatitis or any liver disease.

Hepatoprotective agents

These generally prevent various types of liver infections prophylactically.

Hepatotoxic agents

These generally promote the healing process of the liver.

In India, The use of herbal products for the management of disease has a long history, starting with Ayurvedic management, and proceeding to the European and Chinese alternative systems of ancient medicines. Medicinal plants are significant sources of hepatoprotective drugs. According to one estimate, more than 700 mono- and polyherbal preparations in the form of decoction, tincture, and tablets have been used in various liver disorders. The 21st century has seen a paradigm shift toward therapeutic evaluation of herbal products in liver disease models by carefully synergizing the strengths of the traditional system of medicine with that of the modern concept of evidence-based therapeutical screening, authentication, and randomized placebo-controlled clinical trials to support clinical efficacy. A large number of plants and formulations have been claimed to show hepatoprotective activities. Around 160 active constituents from 101 plants are claimed to have post liver protecting activity. In India, quite eighty seven plants square measure used in 33 patented propitiatory multi-ingredient plant formulations. In spite of the tremendous advances made, no important and safe hepatoprotective agents are obtainable in modern medicine Therefore, due importance has been given globally to develop primarily plant-based hepatoprotective medications that are effective against a range of liver disorders. A drug having helpful results on the liver is understood as a hepatoprotective drug. On the other hand, drugs having toxic effects on the liver are called hepatotoxic drugs. Clinical analysis has conjointly shown that herbals have real utility in the treatment of diseases. In the last 30 years, several hepatotoxins have been used commonly in d-galactosamine, carbon tetrachloride, acetaminophen, and thioacetamide, and more recently Concanavalin A (ConA) and lipopolysaccharide (LPS) has been developed. ConA and LPS do not reflect the clinical pattern of human disease, which indicates a great advantage in the study of cellular mechanisms involved in autoimmune liver disease. The galactosamine model is a highly selective hepatotoxin that causes liver damage similar to human viral hepatitis via depletion of uridine nucleotides, which subsequently diminishes the synthesis of RNA and proteins.^[1] Galactosamine intoxication in rats disrupts the membrane permeability of the plasma membrane, causing leakage of the enzymes from the cell, which leads to the elevation of serum enzymes. Hence, a significant rise in the transaminase levels could be taken as an index of liver damage. Galactosamine has great liver specificity compared to other toxic groups, such as paracetamol, acetaminophen, and carbon tetrachloride because hepatocytes have high levels of galactokinase and galactose-1-uridylyltransferase, and galactosamine does not affect other organs. Galactosamine induces hepatotoxicity with spotty hepatocytes, necrosis, and marked portal and parenchymal infiltration.^[2] Galactosamine also induces the depletion of uridine diphosphate (UDP) by increasing the production of UDP-sugar derivatives, which causes inhibition of RNA and protein synthesis, leading to cell membrane deterioration.^[3] The current

study is aimed toward assembling information-supported reported works on promising phytochemical from herbal plants that are tested in hepatotoxicity models. The review deals with fact-finding work done on herbals helpful in the treatment of liver ailments. The failure of the synthetic drugs in the treatment of hepatic diseases and the search for potent immunomodulatory agents are leading us to the world of herbal medicine in search of a product in nature for use in the protection and cure of dreaded liver diseases. Till date, there is only one protective natural drug; that, too, is not curative and also has its limitations in protecting the liver from viral attacks. The list of herbal hepatoprotective agents has been summarized in Table 1.

IMMUNOMODULATORS

Immunomodulators are of three types:

- Immunoenhancers
- Immunosuppressants
- Immuno adjuvants.

Immunoenhancers

These are the agents that enhance or stimulate the immune system, and they are used in immunodeficiency diseases.

Immunosuppressants

These are the agents that suppress the immune system and are used in autoimmune diseases, or in organ transplantation.

Immuno adjuvants

These agents are used for enhancing the efficacy of vaccines and therefore could be considered as specific immune stimulants, e.g., Freund's adjuvant use in bacillus Calmette-Guérin (BCG) vaccination.

An immunomodulator is natural or artificial, which could stimulate, suppress, or modulate any of the elements of the immune system, including both innate adaptive arms of the immune response. Chronic inflammation is concerned within the pathological process of most common cancers. The etiology of the inflammation is varied and includes the organism, chemical, and physical agents. Many of the factors involved in chronic inflammation play a dual role in the process, promoting neoplastic progression, thereby facilitating cancer prevention.^[1] About 25% of all cancers are related to chronic infection and inflammation.^[2] Throughout, inflammation acts as associate accommodative multitude resistance against illness or injury and is primarily a self-limiting methodology; inadequate resolution of the inflammatory responses typically ends up in numerous chronic ailments besides cancer.^[3] A comprehensive understanding of the molecular and cellular inflammatory mechanisms involved is important for developing preventive and therapeutic ways in which against cancer.^[26] The nitric oxide (NO) acts through the stimulation of the soluble guanyl cyclase, which could be a heterodimeric enzyme with subsequent formation of cyclic-guanosine monophosphate (GMP). Cyclic-GMP activates protein kinase G, which causes the reuptake of Ca²⁺ and the opening of calcium, which activates K⁺ channels. The expression of inducible nitric oxide synthase (iNOS) and so the extent of NO is shown to be elevated in varied metastatic tumor lesions and carcinomas. The study demonstrated that the topical application of phorbol ester induced iNOS expression and subsequent NO production, which successively induced cyclooxygenase-2 expression via nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) activation in mouse skin.^[27] In response to inflammatory cytokines [e.g., tumor necrosis factor (TNF)-α and interleukin (IL)-1β] or other inflammatory stimuli [e.g., phorbol ester, ultraviolet B (UVB), LPS, and dextran sulfate sodium (DSS)], iNOS is transactivated by some transcription factors, including NF-κB.^[28,29] The NO acts through the

Table 1: The reported herbal hepatoprotective agents

Plant names	Category	Name of active constituent	Mechanism	References
<i>Allium sativum</i>	Organosulfur compounds	Organosulfur compounds	Prevention of GSH depletion, alteration of GSH-dependent enzymes	[4]
<i>Buddleja officinalis</i>	Phenyl ethanoid Glycoside	Acteoside	Decreased levels of AST, ALP	[5]
<i>Camellia sinensis</i>	Polyphenols	Catechin	Inhibited hepatocellular apoptosis and unregulated Bcl-2 protein expression	[6]
<i>Cistus laurifolius</i> L.	Flavonoid	Quercetin	MDA, AST, GSH levels decreased	[7]
<i>Corydalis saxicola</i>	Alkaloid	Dehydrocavidine	Decreased levels MDA, SOD, GPx	[8]
<i>Egletes viscosa</i> Less.	Flavonoid	Ternatin	Decreased lipid peroxidation	[9]
<i>Gardenia jasminoides</i>	Iridoid Glycoside	Geniposide	Antioxidant	[10]
<i>Ginkgo biloba</i> L.	Polyphenols	Polyprenols	ALT, AST, ALP, ALB, TP, HA, LN, TG, and CHO levels decreased	[11]
<i>Gossypium herbaceum</i>	Polyphenols	Gossypol	Antioxidant	[12]
<i>Hibiscus sabdariffa</i> L.	Polyphenols	Protocatechuic acid	LDH, AST, ALP, MDA levels decreased	[13]
<i>Larrea tridentata</i>	Resin	Nordihydroguaiaretic acid	Antioxidant	[14]
<i>Magnolia officinalis</i>	Polyphenols	Magnolol	Antioxidant	[15]
<i>Mangifera indica</i>	Triterpene	Lupeol	Decreased levels of SGOT, SGPT, ALP, bilirubin	[16]
<i>Nigella sativa</i>	Quinones	Thymoquinone (TQ)	Scavenger of superoxide, hydroxyl radical, and singlet molecular oxygen	[17]
<i>Ocimum basilicum</i>	Phenolic Acids	Rosmarinic acid	AST, ALP, SGOT levels decreased	[18]
<i>Peumus boldus</i>	Alkaloid	Boldine	Lipid peroxidation	[19]
<i>Phyllanthus amarus</i>	Polyphenols	Phyllanthin	SGOT, SGPT, ALKP, SBLN and total protein levels decreased	[20]
<i>Pinus maritima</i>	Polyphenols	Pycnogenol	SOD, GSH-Px, GSH-reductase, and TBARS levels decreased	[21]
<i>Rubia cordifolia</i>	Glycoside	Rubiadin	SGOT, SGPT, SALP, and gamma-GT levels decreased	[22]
<i>Schisandra chinensis</i>	Lignans	Wuweizisu	Antioxidant	[23]
<i>Sida cordifolia</i>	Organic compound	Fumaric acid	Antioxidant	[24]
<i>Silybum marianum</i>	Lignans	Silymarin	Antioxidant	[25]

GSH=Glutathione

Table 2: The reported herbal immunomodulatory agents

Plants	Parts	Active constituents	Mechanism of action	References
<i>Panax ginseng</i>	Root	Ginsenoside	Proliferation of lymphocytes	[31]
<i>Centella asiatica</i>	Root	Asaticoside A, asiaticoside B	Proliferation of lymphocytes and natural killer cells	[8]
<i>Glycyrrhiza glabra</i>	Root and rhizome	Glycyrrhizin	Increase in spleen weight	[32]
<i>Asparagus racemosus</i>	Root and leaves	Shatavarin 1-4	Increase in production of TNF	[33]
<i>Aralia mandshurica</i>	Dried root	Saponine	Increases phagocytosis	[34]
<i>Picrorhiza kurroa</i>	Dried rhizome	Picroside-I, II, kutkoside	Anticomplement activity	[35]
<i>Lawsonia alba</i>	Dried leaves	Lawson, apigenin, luteolin, and cosmosin	Stimulation of neutrophils and phagocytosis	[36]
<i>Brassica oleracea</i>	Root	Sulforaphane	Enhancement of antibody titer	[37]
<i>Viscum album</i>	Whole	Viscumin	Stimulates lymphocytes	[38]
<i>Canavalia ensiformis</i>	Whole	Lectins	Human neutrophil aggregation and H ₂ O ₂ release	[39]
<i>Linum usitatissimum</i>	Whole	Cyclopeptide A	Immunosuppressant	[40]
<i>Artemisia princeps</i>	Leaves	Protein	Induces interferon	[41]
<i>Echinacea purpurea</i>	Roots and rhizomes	Arabinogalactan	Stimulates phagocytosis	[42]
wheat bran	Seed	Hetroxylan	Stimulates phagocytosis	[43]
<i>Curcuma longa</i>	Rhizome	Curcumin	Inhibits human neutrophils	[44]
<i>Aloe Vera</i>	Dried juice of leaves	Acemannan	Anticomplement activity	[45]
<i>Rumex acetosella</i>	Leaves	Rhamnogalacturonans (pectins and related gums and mucilages, type A), acidic arabinogalactans (mainly plant mucilages, gums, and some hemicelluloses, type B), and neutral glucans and heteroglycans (reserve polysaccharides, type C)	Antiplogistic activity	[46]
<i>Dioscorea membranacea</i> Pierre	Rhizome	Dioscorealide B	Lymphocyte proliferation	[47]
<i>Timospora cordifolia</i>	Whole plant	Cardiofoliosides A and B	Activates macrophages	[48]
<i>Litchi chinensis</i> Sonn.)	Fruits	Epicatechins, proanthocyanidin B2, and proanthocyanidin B4	Proliferation of mouse splenocytes	[49]
<i>Plumbago zeylanica</i>	Root	Plumbagin	Stimulates phagocytosis	[50]
Rice bran	Seed	Ferulic acid ester of oligosaccharides	Increases phagocytosis	[51]
<i>Pimpinella anisum</i>	Fruit	Anethole	Increases leukocyte number	[52]

Contd...

Table 2: Contd...

Plants	Parts	Active constituents	Mechanism of action	References
<i>Catharanthus roseus</i>	Whole plant	Vincristine	Induces antibody production	[53]
<i>Claviceps purpurea</i>	Dried sclerotium of fungus	Ergot alkaloids	Immunomodulates TNF	[54]
<i>Withania somnifera</i>	Dried root	Withaferin A	NO production	[55]
<i>Uncaria tomentosa</i>	Bark	Two mixtures of tetracyclic and pentacyclic oxindole alkaloids	Peripheral blood mononuclear cells	[56]
<i>Phellodendron amurense</i>	Bark	Phellodendrine	Immunosuppressant	[57]
<i>Cissampelos pareira</i> Linn	Leaves	Berberine	Enhances phagocytosis	[58]

stimulation of the soluble guanyl cyclase; is expressed in the cytoplasm of almost all mammalian cells; and mediates a wide range of important physiological functions such as inhibition of platelet aggregation, vasodilatation, neuronal signal transduction, and immunomodulation. The NO is also generated by phagocytes (neutrophils, monocytes, and macrophages) as part of the human immune response.^[30] Phagocytes are formed with iNOS, which is activated by interferon-gamma (IFN- γ) as a single signal or by TNF along with a second signal. On the other hand, transforming growth factor- β (TGF- β) provides a strong inhibitory signal to iNOS, whereas IL-4 and IL-10 provide weak inhibitory signals. In this way, the immune system may regulate the armamentarium of phagocytes that play a role in inflammation and immune responses. The reported herbal immunomodulatory agents have been summarized in Table 2.

CONCLUSIONS

Herbal and traditional botanical products have been used since ancient times for the treatment of various disorders and diseases. Those herbal plants have been discussed that have been previously explored by various researchers for their hepatoprotective and immunomodulatory activities. Several medicinal plants exhibit not only hepatoprotective and immunomodulatory activities but also a wide range of anticancer, cardiatic, diuretic, antiarrhythmic, and other medicinal activities. New immunomodulatory and hepatoprotective plants are important for the discovery of drugs that are less costly, have fewer side effects, are more potent, and allow effective treatment developed for hepatoprotection and immune response. Herbal therapies are free from side effects and toxicity, unlike allopathic medicines. Studies on hepato- and immunomedicinal herbs will contribute to the benefit of the populations needing herbal treatment for both diseases without involving the use of synthetic drugs and reducing the side effects of synthetic drugs.

Acknowledgment

The authors acknowledge the financial support of the National Medicinal Plant Board (NMPB).

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

- Mueller MM, Fusenig NE. Friends or foes - bipolar effects of the tumour stroma in cancer. *Nat Rev Cancer* 2004;4:839-49.
- Hussain SP, Harris CC. Inflammation and cancer: An ancient link with novel potentials. *Int J Cancer* 2007;121:2373-80.
- Schottenfeld D, Beebe-Dimmer J. Chronic inflammation: A common and important factor in the pathogenesis of neoplasia. *CA Cancer J Clin* 2006;56:69-83.
- Sabayan B, Foroughinia F, Chohedry A. A postulated role of garlic organosulfur compounds in prevention of valproic acid hepatotoxicity. *Med Hypotheses* 2007;68:512-4.
- Lee KJ, Woo ER, Choi CY, Shin DW, Lee DG, You HJ, *et al.* Protective effect of acteoside on carbon tetrachloride-induced hepatotoxicity. *Life Sci* 2004;74:1051-64.
- Xu C, Shu WQ, Qiu ZQ, Chen JA, Zhao Q, Cao J. Protective effects of green tea polyphenols against subacute hepatotoxicity induced by microcystin-LR in mice. *Environ Toxicol Pharmacol* 2007;24:140-8.
- Kupeli E, Orhan DD, Yesilada E. Effect of *Cistus laurifolius* L. leaf extracts and flavonoids on acetaminophen-induced hepatotoxicity in mice. *J Ethnopharmacol* 2006;103:455-60.
- Wang T, Sun NL, Zhang WD, Li HL, Lu GC, Yuan BJ, *et al.* Protective effects of dehydrocavidine on carbon tetrachloride-induced acute hepatotoxicity in rats. *J Ethnopharmacol* 2008;117:300-8.
- Souza MF, Rao VS, Silveira ER. Prevention of acetaminophen-induced hepatotoxicity by ternatin, a bioflavonoid from *Egletes viscosa* less. *Phytother Res* 1998;12:557-61.
- Tseng TH, Chu CY, Huang JM, Shioh SJ, Wang CJ. Crocetin protects against oxidative damage in rat primary hepatocytes. *Cancer Lett* 1995;97:61-7.
- Yang L, Wang CZ, Ye JZ, Li HT. Hepatoprotective effects of polyphenols from *Ginkgo biloba* L. leaves on CCl₄-induced hepatotoxicity in rats. *Fitoterapia* 2011;82:834-40.
- Randel RD, Chase CC Jr, Wyse SJ. Effects of gossypol and cottonseed products on reproduction of mammals. *J Anim Sci* 1992;70:1628-38.
- Tseng TH, Hsu JD, Lo MH, Chu CY, Chou FP, Huang CL, *et al.* Inhibitory effect of Hibiscus protocatechuic acid on tumor promotion in mouse skin. *Cancer Lett* 1998;126:199-207.
- Arteaga S, Andrade-Cetto A, Cardenas R. Larrea tridentata (Creosote bush), an abundant plant of Mexican and US-American deserts and its metabolite nordihydroguaiaretic acid. *J Ethnopharmacol* 2005;98:231-9.
- Lin YR, Chen HH, Ko CH, Chan MH. Neuroprotective activity of honokiol and magnolol in cerebellar granule cell damage. *Eur J Pharmacol* 2006;537:6-9.
- Kumari A, Kakkar P. Lupeol prevents acetaminophen-induced *in vivo* hepatotoxicity by altering the Bax/Bcl-2 and oxidative stress-mediated mitochondrial signaling cascade. *Life Sci* 2012;90:561-70.
- Abdel-Wahab WM. Protective effect of thymoquinone on sodium fluoride-induced hepatotoxicity and oxidative stress in rats. *J Basic Appl Zoo* 2013;66:263-70.
- Liu Y, Flynn TJ, Ferguson MS, Hoagland EM. Use of the Combination Index to determine interactions between plant-derived phenolic acids on hepatotoxicity endpoints in human and rat hepatoma cells. *Phytomedicine* 2013;20:461-8.
- Lanthers MC, Joyeux M, Soulimani R, Fleurentin J, Sayag M, Mortier F, *et al.* Hepatoprotective and anti-inflammatory effects of a traditional medicinal plant of Chile, *Peumus boldus*. *Planta Med* 1991;57:110-5.
- Liu CL, Wang JM, Chu CY, Cheng MT, Tseng TH. *In vivo* protective effect of protocatechuic acid on tert-butyl hydroperoxide-induced rat hepatotoxicity. *Food Chem Toxicol* 2002;40:635-41.
- Yang YS, Ahn TH, Lee JC, Moon CJ, Kim SH, Jun WJ, *et al.* Protective effects of pycnogenol on carbon tetrachloride-induced hepatotoxicity in Sprague-Dawley rats. *Food Chem Toxicol* 2008;46:380-7.
- Rao GM, Rao CV, Pushpangadan P, Shirwaikar A. Hepatoprotective effects of rubiadin, a major constituent of *Rubia cordifolia* Linn. *J Ethnopharmacol* 2006;103:484-90.
- Cheng N, Ren N, Gao H, Lei X, Zheng J, Cao W. Antioxidant and hepatoprotective effects of *Schisandra chinensis* pollen extract on CCl₄-induced acute liver damage in mice. *Food Chem Toxicol* 2013;55:234-40.
- Silva RL, Melo GB, Melo VA, Antonioli AR, Michellone PR, Zucoloto S, *et al.* Effect of the aqueous extract of *Sida cordifolia* on liver regeneration after partial hepatectomy. *Acta Cir Bras* 2006;21(Suppl 1):37-9.
- Upadhyay G, Kumar A, Singh MP. Effect of silymarin on pyrogallol- and rifampicin-induced hepatotoxicity in mouse. *Eur J Pharmacol* 2007;565:190-201.
- Hold GL, El-Omar EM. Genetic aspects of inflammation and cancer. *Biochem J* 2008;410:225-35.
- Chun KS, Cha HH, Shin JW, Na HK, Park KK, Chung WY, *et al.* Nitric oxide induces expression of cyclooxygenase-2 in mouse skin through activation of NF-kappaB. *Carcinogenesis* 2004;25:445-54.
- Surh YJ, Chun KS, Cha HH, Han SS, Keum YS, Park KK, *et al.* Molecular mechanisms underlying chemopreventive activities of anti-inflammatory phytochemicals: Down-regulation of COX-2 and iNOS through suppression of NF-kappaB activation. *Mutat Res* 2001;480-481:243-68.
- Yang J, Liao X, Agarwal MK, Barnes L, Auron PE, Stark GR. Unphosphorylated STAT3 accumulates in response to IL6 and activates transcription by binding to NF-kappaB. *Genes Dev* 2007;21:1396-408.
- Green SJ, Mellouk S, Hoffman SL, Meltzer MS, Nacy CA. Cellular mechanisms of nonspecific immunity to intracellular infection: Cytokine-induced synthesis of toxic nitrogen oxides from L-arginine by macrophages and hepatocytes. *Immunol Lett* 1990;25:15-9.

31. Song X, Chen J, Sakwivatkul K, Li R, Hu S. Enhancement of immune responses to influenza vaccine (H3N2) by ginsenoside Re. *Int Immunopharmacol* 2010;10:351-6.
32. Bordbar N, Karimi MH, Amirghofran Z. The effect of glycyrrhizin on maturation and T cell stimulating activity of dendritic cells. *Cell Immunol* 2012;280:44-9.
33. Gautam M, Saha S, Bani S, Kaul A, Mishra S, Patil D, *et al.* Immunomodulatory activity of *Asparagus racemosus* on systemic Th1/Th2 immunity: Implications for immunoadjuvant potential. *J Ethnopharmacol* 2009;121:241-7.
34. Lupusoru CE, Zagnat M, Ghiciuc CM, Lupusoru R, Stănescu U, Grigorescu E. The antistress effect of the saponinic summ extracted from roots of *Aralia mandshurica* cultivated in Moldavia. *Rev Med Chir Soc Med Nat Iasi* 2008;112:1092-7.
35. Puri A, Saxena RP, Guru PY, Kulshreshtha DK, Saxena KC, Dhawan BN. Immunostimulatory activity of picroliv, the iridoid glycoside fraction of *picrorhiza kurroa*, and its protective action against leishmania donovani infection in hamsters 1. *Planta Med* 1992;58:528-32.
36. Mikhaeil BR, Badria FA, Maatooq GT, Amer MM. Antioxidant and immunomodulatory constituents of henna leaves. *Z Naturforsch C* 2004;59:468-76.
37. Thejass P, Kuttan G. Immunomodulatory activity of Sulforaphane, a naturally occurring isothiocyanate from broccoli (*Brassica oleracea*). *Phytomedicine* 2007;14:538-45.
38. Gardin NE. Immunological response to mistletoe (*Viscum album* L.) in cancer patients: A four-case series. *Phytother Res* 2009;23:407-11.
39. Timoshenko AV, Cherenkevich SN. H₂O₂ generation and human neutrophil aggregation as affected by lectins. *Gematol Transfuziol* 1995;40:32-5.
40. Picur B, Cebrat M, Zabrocki J, Siemion IZ. Cyclopeptides of *Linum usitatissimum*. *J Pept Sci* 2006;12:569-74.
41. Kim TH, Lee SJ, Rim HK, Shin JS, Jung JY, Heo JS, *et al.* *In vitro* and *in vivo* immunostimulatory effects of hot water extracts from the leaves of *Artemisia princeps* Pampanini cv. Sajabal. *J Ethnopharmacol* 2013;149:254-62.
42. Classen B, Thude S, Blaschek W, Wack M, Bodinet C. Immunomodulatory effects of arabinogalactan-proteins from *Baptisia* and *Echinacea*. *Phytomedicine* 2006;13:688-94.
43. Hromádková Z, Paulsen BS, Polovka M, Košťálová Z, Ebringerová A. Structural features of two heteroxylan polysaccharide fractions from wheat bran with anti-complementary and antioxidant activities. *Carbohydr Polym* 2013;93:22-30.
44. Antoine F, Simard JC, Girard D. Curcumin inhibits agent-induced human neutrophil functions *in vitro* and lipopolysaccharide-induced neutrophilic infiltration *in vivo*. *Int Immunopharmacol* 2013;17:1101-7.
45. Im SA, Lee YR, Lee YH, Lee MK, Park YI, Lee S, *et al.* *In vivo* evidence of the immunomodulatory activity of orally administered Aloe vera gel. *Arch Pharm Res* 2010;33:451-6.
46. Ovodov IuS. Polysaccharides of flower plants: Structure and physiological activity. *Bioorg Khim* 1998;24:483-501.
47. Panthong S, Ruangnoo S, Thongdeeying P, Sriwanthana B, Itharat A. Immunomodulatory activity of *Dioscorea membranacea* Pierre rhizomes and of its main active constituent Dioscorealide B. *BMC Complement Altern Med* 2014;14:403.
48. Sharma U, Bala M, Kumar N, Singh B, Munshi RK, Bhalerao S. Immunomodulatory active compounds from *Tinospora cordifolia*. *J Ethnopharmacol* 2012;141:918-26.
49. Zhao M, Yang B, Wang J, Liu Y, Yu L, Jiang Y. Immunomodulatory and anticancer activities of flavonoids extracted from litchi (*Litchi chinensis* Sonn) pericarp. *Int Immunopharmacol* 2007;7:162-6.
50. Zhang K, Ge Z, Da Y, Wang D, Liu Y, Xue Z, *et al.* Plumbagin suppresses dendritic cell functions and alleviates experimental autoimmune encephalomyelitis. *J Neuroimmunol* 2014;273:42-52.
51. Fang HY, Chen YK, Chen HH, Lin SY, Fang YT. Immunomodulatory effects of feruloylated oligosaccharides from rice bran. *Food Chem* 2012;134:836-40.
52. Domiciano TP, Dalalio MM, Silva EL, Ritter AM, Estevão-Silva CF, Ramos FS, *et al.* Inhibitory effect of anethole in nonimmune acute inflammation. *Naunyn Schmiedebergs Arch Pharmacol* 2013;386:331-8.
53. Whitley NT, Day MJ. Immunomodulatory drugs and their application to the management of canine immune-mediated disease. *J Small Anim Pract* 2011;52:70-85.
54. Fiserová A, Kovářů H, Hajduová Z, Mares V, Starec M, Kren V, *et al.* Neuroimmunomodulation of natural killer (NK) cells by ergot alkaloid derivatives. *Physiol Res* 1997;46:119-25.
55. Oh JH, Lee TJ, Park JW, Kwon TK. Withaferin A inhibits iNOS expression and nitric oxide production by Akt inactivation and down-regulating LPS-induced activity of NF- κ B in RAW 264.7 cells. *Eur J Pharmacol* 2008;599:11-7.
56. Winkler C, Wirleitner B, Schroecksnadel K, Schennach H, Mur E, Fuchs D. *In vitro* effects of two extracts and two pure alkaloid preparations of *Uncaria tomentosa* on peripheral blood mononuclear cells. *Planta Med* 2004;70:205-10.
57. Choi YY, Kim MH, Han JM, Hong J, Lee TH, Kim SH, *et al.* The anti-inflammatory potential of Cortex *Phellodendron* *in vivo* and *in vitro*: Down-regulation of NO and iNOS through suppression of NF- κ B and MAPK activation. *Int Immunopharmacol* 2014;19:214-20.
58. Bafna A, Mishra S. Antioxidant and immunomodulatory activity of the alkaloidal fraction of *Cissampelos pareira* linn. *Sci Pharm* 2010;78:21-31.



Uorakkottil Ilyas



Deepshikha P. Katare



Vidhu Aeri



Punnooth Poonguzi Naseef

ABOUT AUTHORS

Uorakkottil Ilyas, PhD Scholar, Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Hamdard University, New Delhi, India, 110062.

Prof. (Dr.) Deepshikha P. Katare, Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, Amity University, Noida, India, 201301.

Dr. Vidhu Aeri, Associate Professor Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Hamdard University, Hamdard Nagar, New Delhi - 110 062, India. E-mail: vdhaeri@yahoo.com

Punnooth Poonguzi Naseef, Associate Professor, Department of Pharmaceutics, Jamia Salafiya Pharmacy College, Kerala, India.