# A Comprehensive Review on Traditional Knowledge, Phytochemistry and Pharmacological Properties of Acalypha indica L.

Sahukari Ravi, Punabaka Jyothi, Bhasha Shanmugam, Ganjikunta Venkata Subbaiah, Singamala Hari Prasad, Kesireddy Sathyavelu Reddy

#### ABSTRACT

Acalypha indica is a significant medicinal plant. The purpose of this review is to bring traditional usage, phytochemistry, and scientific applications of *A. indica* up to date. Microbial infections, fertility, stomach ulcers, snake bites, pains, wounds, liver/kidney problems, and rheumatism have all been traditionally treated with *A. indica* paste, decoction, sap, and synergy with other plants/plant products, which have all been scientifically proven through *in vitro* and *in vivo* experiments. Regardless of traditional knowledge, this plant extracts have been scientifically proven to help against cancer, inflammation, cardiac damage, diabetes, TB, and malaria. Phytochemical investigation revealed that *A. indica* has phenols, flavonoids, tannins, coumarins, alkaloids and their glycosides, saponins, volatiles and fatty acids. In summary, the presence of phytoconstituents is responsible for the *A. indica* traditional and pharmacological qualities. Further, conformational clinical trails in humans are necessary to ascertain the extracts efficacy. Extensive future studies are to be conducted to reveal the mechanism of action, pharmacokinetic properties and active phythochemicals of *A. indica* extracts.

Key words: Acalypha indica, Phytochemicals, Pharmacological properties, Traditional uses, Medicinal plants.

# **INTRODUCTION**

Data collection

Among the 270,000 higher plants many are known to be of medicinally importance in India.<sup>[1]</sup> Due to plants diversity in India and their usage in traditional medicine to treat many diseases, India is hub for medicinal plants / natural medicine. The primary health care of Indian people greatly met through the consumption of traditional medicine (Ayurveda, Sidha and Unani) in different formulations due to lack of modern health facilities and very expensive of medical treatment. Despite the fact that there are 456 Acalypha species, <sup>[2]</sup> only a few were used traditionally and examined scientifically. A. indica is one among them, and it is frequently used to cure a variety of diseases. Besides of India, it also occupied around Asia and African countries etc, to treat diverse health issues. More importantly, available information on this plant has not been reviewed completely in previously published reviews. Hence, medicinal importance of this plant was made an attraction to review the available information for the asset to scientific community.

Information on traditional uses of A. indica and

scientific evidence as gathered from local books

published on A. indica and internet sources. In addition,

scientific literature databases such as Pubmed, Medline, Google scholar, Science direct, Springer, Wiley online library, The plant database,<sup>[2]</sup> Kew-Royal botanical garden<sup>[3]</sup> and other online resources have also been utilized. Doctoral theses available on A. indica have been obtained from Shodhganga,<sup>[4]</sup> a reservoir of Indian theses and also from local universities of Andharapradesh. The phytochemical structures have drawn in Marvin Sketch. To obtain information from above sources, initial search was started with A. indica. Further, some other scientific key words have used which include 'biological activity', 'phytochemicals', 'phytoconstituents', 'isolation of compounds', 'nanoparticles' and 'micro particles' where A. indica has been kept as a common prefix for all key words. By gathering all the available information, we comphressively reviewed about A. indica.

## Acalypha indica L. Synonyms

The plant *A. indica* belongs to *Euphorbiaceae*. It grows in India, Indian Ocean islands, South-East Asia, Oceania, East Africa to southern Africa including South Africa and introduced into warmer parts of the world (Figure 1).<sup>[5,6]</sup> The accepted binomial is *Acalypha indica* L. but there are other scientific

**Cite this article:** Ravi S, Jyothi P, Shanmugam B, Subbaiah GV, Prasad SH, Reddy KS. Comprehensive Review on Traditional Knowledge, Phytochemistry and Pharmacological Properties of *Acalypha indica* L. Pharmacog Rev. 2021;15(30):134-85.

Sahukari Ravi, Punabaka Jyothi, Bhasha Shanmugam, Ganjikunta Venkata Subbaiah, Singamala Hari Prasad, Kesireddy Sathyavelu Reddy

Division of Molecular Biology and Ethnopharmacology, Department of Zoology, Sri Venkateswara University, Tirupati, Andhra Pradesh, INDIA.

#### Correspondence

#### Dr. Kesireddy Sathyavelu Reddy,

Professor, Division of Molecular Biology and Ethnopharmacology, Department of Zoology, Sri Venkateswara University, Tirupati–517502, Andhra Pradesh, INDIA.

Phone no : +91 8772240265

E-mail: ksreddy2008@hotmail.com

#### History

- Submission Date: 28-05-2021;
- Review completed: 20-06-2021;
- Accepted Date: 06-08-2021

#### DOI: 10.5530/phrev.2021.15.16

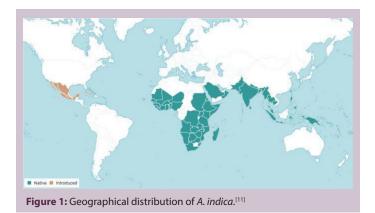
Article Available online

#### http://www.phcogrev.com/v15/i30

#### Copyright

© 2021 Phcog.Net. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.





synonyms including Acalypha bailloniana Müll.Arg.; Acalypha canescens Benth.; Acalypha caroliniana Blanco; Acalypha chinensis Benth.; Acalypha ciliata Benth.; Acalypha cupamenii Dragend.; Acalypha decidua Forssk.; Acalypha fimbriata Baill.; Acalypha indica K.Schum. and Hollr.; Acalypha indica var. australis F.M.Bailey; Acalypha indica var. bailloniana (Müll.Arg.) Hutch.; Acalypha indica var. minima (H.Keng) S.F.Huang and T.C.Huang.; Acalypha indica Vell.; Acalypha somalensis Pax.; Acalypha somalium Müll.Arg.; Acalypha spicata Forssk.; Cupamenis indica (L.) Raf.; Ricinocarpus baillonianus (Müll.Arg.) Kuntze; Acalypha indica var. mexicana (Müll.Arg.) Pax and K.Hoffm. Ricinocarpus deciduus (Forssk.) Kuntze. and Ricinocarpus indicus (L.) Kuntze; Acalypha tenuis Klotzsch ex Pax and K.Hoffm.<sup>[2,3]</sup>

#### Vernacular names

Though, this plant has scientific name and synonyms, the local people have given a specific name to it for their convenience as follows: Chikka Emas and Galak Kuching (Malay); Tie xian (Chinese); rumput bolongbolong, Rumput kokosongan, Kucing-kucingan and Lelatang (Indonesian); Ricinelle des Indes, Oreille de chatte, Horrisa and Herba chatte (French); Indian acalypha, Indian nettle and three-seeded mercury (English); Brennkraut (German); Ricinela (Spanish); Alcalifa (Brazil). Besides, Indians have given special prirority in the naming of this plant. Kuppichettu, Kuppinta, Gaayapaku and Muripinda (Telugu); Kuppaimeni and Kuppaveni (Tamil); Kuppigada (Kannada); Kuppameni (Malayalam); Sveta-basanta (Gujaratis); Kuppikhokli (Hindi); and Haritamanjari (Sanskrit).<sup>[7,8]</sup>

It is a common weed, annual herb; grows up to 30-70 cm height, sometimes little more. This plant has few ascending branches, which are angled and pubescent; the leaves are broadly ovate, longer than 3-5 cm with serrate edges and arranged pinnately; stipules are very minute; flowers are sessile, present on erect axillary spikes and grow longer than the leaf; male flowers are minute, crowded and distally arranged; the stamens of female flowers are scattered along with inflorescence axis, where each one is subtended by a conspicuous semicircular foliaceous toothed green bract; the capsules are hispid and about 1 mm wide with 3-locular.<sup>[9]</sup>

Effect of season (summer and monsoon) and altitudes (275, 350 and 550 m above mean sea level) on *A. indica* growth have been studied by Krishnan *et al.* (2000).<sup>[10]</sup> They considered six parameters including leaf (length, breadth and number), branches, and tap root (height and length). Finally, they confirmed that the monsoon and top hill 550 m were suitable growth conditions for *A.indica*.

#### Local and traditional uses of A.indica.

A common weed *A.indica* is used traditionally to alleviate spectrum of diseases of human. Though geographical occupancy of *A.indica* is large

but its traditional information is less abundant (Table 1). Traditional uses from India, Western Nepal, Bangladesh, Mozambique, North Djibouti, Seychelles, Réunion, East Africa, Namibia, Mauritius, Mozambique, Indonesia, Ethiopia, Northern Transvaal, Sri Lanka and Southern Thailand are available today. Among these, Indians used this plant very abundantly; next to Africans followed by other people. Indians are well aware of this plant because of its usage in their traditional medicine Ayurveda including in almost all states (Table 1).

People have used this plant leaves as a juice to treat cough,<sup>[12-14]</sup> ear ache, head ache, syphilitic ulcer, anti-parasiticide,<sup>[15,16]</sup> constipation,<sup>[14,17]</sup> rheumatoid arthritis, pneumonia, emetic,<sup>[18,20]</sup> and scabies.<sup>[21]</sup> The juice from the whole plant is used to treat bronchitis, <sup>[22]</sup> snake bites,<sup>[23]</sup>, pneumonia<sup>[24]</sup> and flatulence.<sup>[25]</sup> Besides, this plant leaves were made into paste alone and used to treat many ailments which include dermatological problems, wounds, chest pain, burns, snake bite and itching whereas the root paste applied to alleviate fungal infections while total plant paste used against diuretic, constipation, skin problems, severe cough.<sup>[26-36]</sup> Also, leaves were used to prepare a decoction for the treatment of asthma<sup>[37]</sup>, intestinal lavage,<sup>[38]</sup> cough, dysentery,<sup>[39]</sup> cold,<sup>[14]</sup> and joint pains<sup>[21]</sup> whereas decoction from the roots was used for asthma, liver and kidney cleaning, as laxative, against intestinal worms and for stomach-ache.<sup>[21,38]</sup> Other parts of this plant like the stem prepared as a decoction against hemorrhoids.<sup>[38]</sup> A decoction from the entire plant ingested to treat earache, toothache, burns and wheezing.<sup>[18,40]</sup> Along with juice, paste and decoction, the other sources also practiced which include infusion, sap and powder.<sup>[40,21]</sup>

This plant/ parts were combined with other plant/parts to treat many diseases in India including: leaves mixed with calcium hydroxide for cough relaxation;<sup>[12]</sup> external application of leaves grind in lime juice or common salt was used against parasites, dog bites, dermatitis and wounds of animals, whereas small balls prepared in lime juice taken orally to cure asthma.<sup>[15,17,21,27,37,41]</sup> Vomiting of animals was prevented by feeding the leaves combined with seeds of Acorus calamus L.; Black quarter disease using a combination of leaves with seeds of Piper nigrum L., leaves of Leucas aspera (Willd.) Link and the bulb of Allium cepa L.; leaf juice with Ferula assa-foetida L. plant used for constipation of animals <sup>[17]</sup> leaves and block pepper grains each nine crushed in cow ghee and taken twice a day with butter milk during jaundice;<sup>[35]</sup> leaves paste with lime juice applied on ring worms;<sup>[42]</sup> A paste of this plant's leaves with clove and black pepper was applied to cure the maggot wounds;<sup>[17]</sup> leaf powder or decoction combined with garlic is used against intestinal worms;<sup>[43]</sup> epilepsy was treated by orally using the leaves combined with pepper, garlic and Leucas aspera (Willd) Link leaves; on other way oral ingestion of leaves mixed with Cardiospermum halicacabum L. boiled in Azadirachta indica A.Juss. oil; leaves with onions were ground, then rubbed on the chest, neck and hips.<sup>[44]</sup> Paste of A.indica, Azadirachta indica, Mimosa pudica leaves with flowers of Albizzia lebbeck taken orally for itching.[45] Bronchitis was treated by combining A.indica leaves and Jumellea fragrans (Thouars) Schltr. tuber infusions with honey. Oral intake of leaves combined with roots of Tylophora indica (Burm.f.) Merr vomit the stomach poison.[21] A.indica roots with pepper, ginger and honey get rid of the hook and tape worms in children.<sup>[46]</sup>

The following benefcial effects of the plant without extraction method are : leaves and roots used against diarrhea;<sup>[47]</sup> seeds used against cholesterol and rheumatism;<sup>[48]</sup> total plant used to treat animal diseases such as anthrax and black quarter,<sup>[17]</sup> human diseases include severe cough,<sup>[43]</sup> antifertility;<sup>[49,50]</sup> eye infections,<sup>[51]</sup> piles, pistula,<sup>[52]</sup> expectorant,<sup>[29]</sup> arthritis, haemoptysi, mania, bed sores, syphilis,<sup>[50]</sup> wound healing,<sup>[53]</sup> flatulence,<sup>[25]</sup> rheumatism<sup>[54]</sup> and antiseptic.<sup>[31]</sup> This incomplete traditional information about *A.indica* extraction is an important one for further utilization in global community.

## Table 1: Local and traditional uses of A.indica.

S.No	Plant source	Practiced on	By whom/where	Refs.
		Leaves		
	Uniform mixture of leaf juice and garlic	Anthelmentic	North Maharashtra, India	[55]
	Decoction (50 mL/day) of leaves taken for one week orally/ small balls (50 mg each) prepared in lemon juice and taken twice a day orally; About 10-15 leaves boiled in one glass of water, then cooled extract about 3 glasses given to the asthma patients upto cure.	Asthma	Andhra Pradesh, India; Boro tribal people of Goalpara district, Assam, India	[37, 56,57 ]
	Dry leaves powder	Bedsores	North Maharashtra, India	[55]
	Leaves of this plant combined with seeds of <i>Piper nigrum</i> L., leaves of <i>Leucas aspera</i> (Willd.) Link and bulb of <i>Allium cepa</i> L. are fed to animals	Black quarter disease of animals	Tamil Nadu, India	[17]
	Combined leaf infusion with tuber infusion of <i>Jumellea fragrans</i> (Thouars) Schltr. were sweetened with honey; leaf juice.	Bronchitis	Seychelles, East Africa; Southern Bankura of West Bengal, India; Boro tribal people of Goalpara district, Assam, India;	[19, 21, 57]
	Leaf paste applied on burns	Burns	Terai forest, Western Nepal	[18]
	Oral ingestion of leaves paste	Chest pain	Malayali tribes in Jawadhu	[35]
			hills of Eastern ghats, Tamil Nadu, India; Silent valley of Kerala, India.	
1	Leaf decoction taken internally; leaf juice prescribed for children	Cold	Malayali tribes in Jawadhu hills of Eastern ghats, Tamil Nadu, India; People of Kalahandi District, Odisha, India.	[14]
	Leaf juice mixed with 5 grams of <i>Ferula assa-foetida</i> L. plant and then used against constipation; Leaf juice taken at night; A leaf petiole dipped in castor oil was inserted into anus of a child for 2 min to relief.	Constipation of animals and humans	Andhra Pradesh, India; Gulbarga district, Karnataka, India; People of Kalahandi District, Odisha, India	[14, 17, 58]
0	Juice of fresh leaves mixed with small amount of calcium hydroxide and applied externally on the throat twice a day for five days; Leaves decoction taken internally; A tea spoon of leaf juice taken orally three times a day for 5-6 days; leaf juice prescribed for children	Cough relaxation	Palliyars of Saduragiri Hills, Western Ghats, Tamil Nadu, India; Malayali tribes in Jawadhu hills of Eastern ghats, Tamil Nadu, India; Satpuda forest east, Maharashtra, India; People of Kalahandi District, Odisha, India.	[12, 13, 14]
1	Leaves	Diabetes	Vidarbha region, Maharashtra, India	[59]
2	Leaves	Diarrhea	Bangladesh	[47]
3	Leaf paste with lime applied on bitten area two times a day for 3-4 days	Dog bite	Village people of Shimoga District, Karnataka, India	[27]
4	Decoction of leaves	Dysentery	Philippines	[39]
5	Juice of leaves	Ear ache	Western Kachchh, Gujarat, India; Tharu tribe of Uttarakhand, India	[15, 60, 61]
6	Leaf juice	Emetic	Southern Bankura of West Bengal, India.	[19]
7	Leaves ground with pepper, garlic and leaves of <i>Leucas aspera</i> (Willd.)Link then given orally; Leaves mixed with <i>Cardiospermum halicacabum</i> L. then subjected to boiling in <i>Azadirachtaindica</i> A.Juss. oil and resulted extract given; Leaves ground with onion resulted mixture rubbed on chest, neck and hips or poured into ears and nose of children below 12 years old and the same	Epilepsy	Yanadis of Cuddapah district, Andhra Pradesh, India. Palliyans of Southern WesternGhats, Tamil Nadu, India.	[44]
8	Leaf sap used as eye drops	Eye infection	East Africa, Namibia	[21]
9	Topical application of leaf paste on head	Fungal infection	Palamalairegion of Eastern Ghats, India; Silent valley of Kerala, India.	[26, 35]
0	Leaf paste applied topically	Ganglion	Tadjourah District of Randa, in north Djibouti.	[62]

Table	1: Continued			
21	Crushed leaves applied topically and decoction taken orally.	Haemorrhoid	Canhane village, Massingir district, Mozambique	[38]
22	Juice of leaves used as nasal drop.	Head ache	Dhenkanal district, Odisha, India.	[15, 16]
23	Leaves and roots (2:1) crushed then supplied with food once a day for five days/ Leaf powder and decoction with small amount of garlic were given to children to expell worms; Ground leaves, decoction and macerated solution subjected to Enema procedure; the leaves ground in water then taken along with sap of Allium sativum	Intestinal worns of cattle / humans	Andhra Pradesh, India; Canhane village, Massingir district, Mozambique; Shaiji Community in Southwestern Bangladesh	[17, 43, 38, 34]
24	Leaf paste prepared and then taken orally	Itching	Kani tribals in Tirunelveli hills, India	[28]
25	Leaves and black pepper grains each nine crushed in cow ghee taken twice a day with butter milk; leaves paste ingested as orally	Jaundice	East Godavari district, Andhra Pradesh, India; Silent valley of Kerala, India.	[63, 35]
26	Leaf decoction used as massage cream/ Poultice of leaves and stem	Joint pains	Comoros; Tribals of Western Ghats, Kerala.	[21, 46]
27	Crushed leaves applied topically and infusion of leaves taken orally	Laxative	Canhane village, Massingir district, Mozambique; Southern Bankura of West Bengal, India; Western Kachchh, Gujarat, India.	[38, 19, 60]
28	Leaf of this plant mixed with lime juice or quicklime or common salt then applied externally	Parasiticide	India	[15]
29	Leaf juice	Pnemonia	Southern Bankura of West Bengal, India; Boro tribal people of Goalpara district, Assam, India	[19, 57]
30	Infusions of leaves and <i>Tylophora indica</i> (Burm.f.) Merr. roots drunk orally	Poisoning of stomach	Réunion	[21]
31	Leaf infusion; Oral administration of tender leaves aqueous extract	Purgative	Réunion and Madagascar; Nicobarese Tribals of Car Nicohar Island, India	[21, 64]
32	Freshly prepared leaf juice is applied on rheumatoid arthritis	Rheumatoid arthritis	Terai forest, Western Nepal; Southern Bankura of West Bengal, India.	[18, 19]
33	Leaf paste with lime juice applied on infected area; Leaf juice applied externally on ringworms.	Ringworm	Rewa district, Madhya Pradesh, India; Terai region of Uttar Pradesh, India	[42, 65]
34	Leaf paste mixed with lemon juice and applied on Scabies of animals; Juice of crushed leaves mixed with salt/ decoction of leaves	Scabies of animals and humans	Kalahandi district, Odisha, India; Mauritius; Terai region of Uttar Pradesh, India	[17, 21, 65]
35	Leaf paste alone/mixed with pepper and applied; paste prepared in water and externally applied on skin two times a day for period of one week; Crushed leaves; Leaf paste mixed with a pinch of lime and was applied;	Skin diseases of animals and humans	Andhra Pradesh, India; Valaiyans of Madurai District, Western Ghats, Tamil Nadu, India; Medagascar; Kodagu district, Karnataka, India; Kancheepuram District, Tamil Nadu, India; Bhadrak District of Odisha, India.	[17, 66, 21,41,32, 33]
36	Leaf paste applied on bitten area; leaves boiled with salt for half an hour and given to the affected animal usually cattle.	Snake bites	Village people of Shimoga District, Karnataka, India; Shaiji Community in Southwestern Bangladesh.	[27, 34]
37	Juice of leaves applied locally	Syphilitic ulcers	Southern Bankura of West Bengal, India.	[15, 19]
38	Combination of leaves of this plant with seeds of <i>Acorus calamus</i> L. were ground then fed to animals; Leaf juice prevents the vomiting in humans	Vomiting of animals/ humans	Tamil Nadu, India; Satpuda forest east, Maharashtra, India.	[17, 13]

Table	e 1: Continued			
39	Crushed leaves use to applied on wounds till cure; Leaf paste mixed with kitchen salt and applied on wounded area; This plant leaves, 3 cloves and 4 block peppers were made into paste then applied on maggot wounds;Leaves of <i>Acalypha indica</i> , <i>Mimosa pudica</i> , <i>Azadirachta indica</i> and flowers of <i>Albizzia lebbeck</i> made into paste and then taken orally once a day for 3 days; Leaf paste prepared and then taken orally; Leaf ground to make paste and applied on the wound	Wound healing of animals and humans	Nizamabad district, Telangana, India; Tamil Nadu, India; Tribal people of Western Ghats of Tamil Nadu, India; Kani tribals in Tirunelveli hills, India; Shaiji Community in Southwestern Bangladesh.	[17, 45, 28, 34]
		Roots		
1	Root decoction or infusion	Asthma	Seychelles and Réunion, Africa; Satpuda forest east, Maharashtra, India.	[13]
2	Roots boiled in water and then eat	Constipation	Shaiji Community in Southwestern Bangladesh.	[34]
3	Topical application of root paste	Fungal infection	Palamalairegion of Eastern Ghats, India	[26]
4	Decoction of root mixed with ginger, pepper and honey	Hook worms and tape worms in children	Tribals of Western Ghats, Kerala, India	[46]
5	Roots and leaves (1:2) crushed and then supplied with food once a day for five days; Root decoction	Intestinal worms of cattle and humans	Andhra Pradesh, India; Seychelles	[17, 21]
6	One table spoon per day of root decoction given orally for month	Lactation	Gondu tribes of Seethagondi Grampanchayath, Adilabad District, Andhra Pradesh, India;	[67]
7	Infusion and decoction of roots administrated orally	Laxative	Bangladesh; Canhane village, Massingir district, Mozambique.	[47, 38]
8	Root decoction or infusion	Liver and kidney cleaning	Seychelles and Réunion, Africa	[21]
9	Root	Rheumatism	Satpuda forest east, Maharashtra, India.	[13]
10	Root decoction	Stomach-ache. <b>Stem</b>	Seychelles	[21]
1	Decoction of stem administrated orally	Hemorrhoid	Canhane village, Massingir district, Mozambique	[38]
1	Seeds Seeds	Cholesterol	The Kabanjahe traditional market, North Sumatra, Indonesia	[48]
2	Seeds	Rheumatism	The Kabanjahe traditional market, North Sumatra, Indonesia	[48]
		Total plant		
1	Whole plant	Anthrax in cattle and camel	Ethiopia	[17]
2	Whole plant	Antifertility (Emmenagogue)	India	[49, 50]
3	Whole plant	Antiseptic	Siddha healers in Virudhunagar district of Tamil Nadu, India	[31]
4	Whole plant	Arthritis	India	[50]
5	15 to 20 mL of whole plant extract was used for one week ; Extract taken at morning expels the sputum and cures; Whole plant juice taken thrice a day	Asthma.	East Godavari and Krishna Districts, Andhra Pradesh, India; Gulbarga district, Karnataka. India; Jhalawar district, Rajasthan, India	[40,50, 68, 58, 24]
6	Whole plant	Bed sores and sores on lips	India	[50]
7	Whole plant	Black quarter disease in animals	Ethiopia	[17]
8	Whole palnt extract given orally	Brain weakness	Jhalawar district, Rajasthan, India	[24]

Table	1: Continued			
9	Crushed whole plant juice given to children/ Paste of whole plant used externally	Bronchitis	Irular, the tribal people of Marudhamalai hills, Coimbatore and Village people of Thoppampatti, Dindigul district, Tamil Nadu, India; East Godavari District, Andhra Pradesh, India;	[22, 40, 50, 30]
10	Whole plant decoction or powder	Burns	India	[40]
11	Whole plant paste used externally	Constipation	Village people of Thoppampatti, Dindigul district, Tamilnadu, India	[50, 30]
12	Whole plant paste used externally	Dermatological ailments	Siddha healers in Virudhunagar district and Village people of Thoppampatti, Dindigul district of Tamil Nadu, India; Araku Valley, Andhra Pradesh, India.	[31, 30, 69]
13	Whole plant	Digestive disorders	Ratanpur region of Bilaspur district , Chhattisgarh, India	[70]
14	Whole plant paste used externally	Diuretic	East Godavari District, Andhra Pradesh, India; Village people of Thoppampatti, Dindigul district, Tamilnadu, India; Malaysia	[40, 30, 52]
15	Plant decoction is given orally	Earache	Terai forest, Western Nepal	[18]
16	Whole plant in the form of fresh or dry eaten as raw material.	Emetic	East Godavari District, Andhra Pradesh, India;	[40, 50, 25, 71]
			Krabi and Songkhla provinces of southern Thailand; South America; Buxar district, Bihar, India.	
17	Whole plant	Expectorant	East Godavari District, Andhra Pradesh, India; Malaysia	[40, 50, 52]
18	Whole plant	Eye diseases	Northern Transvaal	[51]
19	Boiled juice of whole plant given orally	Flatulence	Krabi and Songkhla provinces of southern Thailand	[25]
20	Poultice of whole plant	Headache	India; Mauritius.	[50, 21]
21	Mixing this plant with equal amount of castor oil	Laxative	Malaysia	[52]
22	Whole plant	Mania	India	[50]
23	One table spoon of plant extract given twice a day for three days	Mouth ulcers of babies	Theoraon tribe of Jashpur District, Chhattisgarh, India	[72]
24	Whole plant	Piles	Malaysia	[52]
25	Whole plant	Pistula	Malaysia	[52]
26	Whole plant juice given orally thrice a day	Pneumonia	Jhalawar district, Rajasthan, India;	[24]
27	Tea made from boiled plant taken	Purgative	Malaysia	[52]
28	Whole plant	Rheumatism	Odisha, India.	[54]
29	Plant paste with little salt applied externally	Scabies	Gulbarga district, Karnataka. India	[58]
30	Whole plant paste applied on throat once a day for two days	Severe cough	Rural people of Sivagangai District, Tamil Nadu, Southern India	[36]
31	Whole plant	Severe cough associated with bleeding.	Siddha healers in Virudhunagar district of Tamil Nadu, India.	[43, 50, 31]
32	Juice of whole plant given orally	Snake bites	Western and Sabaragamuwa provinces of Sri Lanka	[23]
33	Whole plant	Syphilis	India	[50]
34	Plant decoction is given orally	Toothache	Terai forest, Western Nepal; India	[18, 40]
35	Plant extract with buttermilk taken	Urinary infections	Gulbarga district, Karnataka. India	[58]
36	Whole plant decoction or powder	Wheezing	India	[40]
37	Whole plant	Wound healing	Sugali tribes of Yerramalais of Kurnool district, Andhra, Pradesh, India; Washim District,Maharashtra, India.	[53,73]

# Phytochemicals Qualitatively estimated phytochemicals Total plant

Methanolic extract has phenolic, flavonoid, alkaloid, tannin, steroid, terpenoids and saponin compounds; whereas alkaloid, flavonoid, phenolic and saponins found in diethyl ether, ethyl acetate and ethanolic extracts.<sup>[74,75]</sup> In an experiment, Pragada *et al.* (2011)<sup>[76]</sup> have found steroids, amino acids and oils in hexane fraction of aqueous alcoholic extract; tannins, amino acids, steroids and oils in ethyl acetate fraction; saponins, flavonoids, aminoacids and oils in methanolic fraction.

## Leaves

leaves have alkaloid, tannin, steroid, saponin, flavonoid, glycoside and phenolic compounds (ethanolic extract); sterols (petroleum ether and chloroform extracts); reducing sugar, coumarin, antho cyanin, anthra quinone, saponin, cardiac glycoside, terpenoid, tannin, alkaloid, flavonoid and phenolic compounds (methanolic extract).<sup>[43,77,78]</sup> Balakrishnan *et al.* (2009)<sup>[79]</sup> were screened to identify the saponin, alkaloid, terpernoid, phenolic and flavonoids in methanol and water extracts of roots.

## Quantified phytochemicals

Subsequent to qualitative analysis of plant extracts, quantification gives an idea for further fractionation and isolation of compounds. Quantified phytochemicals of *A. indica* is shown in Table 2. Flavanones were quantified in methanolic extract, hydroalcoholic extract and its fractions of chloroform and butanol of leaves. Among these, a rich amount (2.56 mg/g) found in butanol insoluble fraction. Flavonoid (29.896 mg/g) and total phenolic (111.321 mg/g) content in lyophilized methanolic extract of leaves were higher than other extracts and fractions. About 16.1 mg/g of saponins were found in methanolic extract of leaves.<sup>[76,78,80-82]</sup>

## Isolated and identified phytochemicals

Traditional attempts on *A.indica* stated the possibilities for pharmacological investigation/research and to established potentiality by isolation of phyto chemicals. To explore phytochemistry of *A.indica*, researchers performed chromatography and spectroscopy techniques. Table 3 presents the phyto constituents of *A.indica* and their biological properties.

*A.indica* has major phytochemical classes such as alkaloids, polyphenols (flavonoids, tannins, coumarins, hydroxy benzoic acids and hydroxy cinnamic acids), volatile compounds, fatty acid derivatives and others.

# Alkaloids and their glycosides

Alkaloids are very essential secondary metabolites of plants and being exhibited potential activity on deadly diseases. In this concern, Hungeling *et al.* (2009)<sup>[87]</sup> and Nahrstedt *et al.* (1982)<sup>[93]</sup> have been isolated alkaloids from leaves-inflorescences powder, whereas Ravi *et al.* (2017)<sup>[78]</sup> identified in leaves of *A.indica* (Tables 3, 4). Among, 8 compounds have been noticed as toxic due to the presence of cyanide hence they called as cynogenic glycosides. These cynogenic compounds are important to the plants to overcome environmental stress and protection from predators but its consumption is very fatal to the humans, which will be discussed in the discussion section of this review. *A.indica* alkaloid rescinnamine used as antihypertensive drug, Pergolide sulfone acts as dopamine receptor inhibitor, Lupinine as anti-coagulant and ambelline has antiproliferative property<sup>[88, 89, 92,220]</sup> but other alkaloids are not tested in biological systems yet. The chemical structure of alkaloids is presented in Table 4.

# Polyphenols

Polyphenols are most significant and definitely abundant among the groups of phytochemicals of plant kingdom, classified into sub classes

and sub divisions based on their origin, structural features and biological function. The flavonoids occupied large portion in polyphenols than coumarins, tannins, hydroxy benzoates, hydroxy cinnamic acid and others. The followings are polyphenolic classes and their respective compounds of *A.indica*.

## Flavonoids

Ten flavonoid structures of this plant are shown in Table 5. Catechin, dehydrovariabilin, rutin and naringenin were identified in the leaves;<sup>[78, 84]</sup> quercetin 3-0- $\beta$ -D-glucoside, rutin and kaempfeorl have been isolated from total plant; whereas, nicotiflorin, biorobin, clitorin and mauritianin were isolated from extracts of leaves and flowers.<sup>[147]</sup> Production of these compounds in this plant is to overcome the oxidative stress caused by biotic and abiotic factors. Apart from this, these compounds also have other beneficial biological properties as mentioned in Table 3.

## Tannins

Eleven tannins were isolated from total plant of *A.indica* using chromatography techniques (Table 6).<sup>[137,210]</sup> These are poly hydroxyl compounds being played crucial role in the scavenging of free radicals and its associated diseases. Except few (Potassium brevifolincarboxylate, acaindinin, acetonylgeraniin A, euphormism M2 and repandusinic acid), rest of tannins have been tested experimentally for biological activities as presented in Table 3, hence further isolation and experimental validation of tannins of *A.indica* needs to be reported.

## Coumarins

Coumarins are one of sub classes of the polyphenols, having potential antioxidant, anti-inflammatory and anti-cancer activities.<sup>[106,108-110]</sup> Three coumarins have ben found in *A.indica* by the application of RP-HPLC and HR-LC-MS as mentioned in Table 3.<sup>[78, 84]</sup> The 2D structures have drawn and illustrated in Table 7.

# Other polyphenols

Apart from the earlier mentioned polyphenols, hydroxy benzoic acid (Gallic acid and Syringic acid) and hydroxy cinnamic acid (Caffeic acid) are present in *A.indica* (Table 8).

# Volatile compounds and fatty acids

The compounds come under volatile category are aldehydes, alcohols, alkanes, alkenes, aromatics, esters, ethers, ketones, steroids and terpenoids. To some extent, fatty acids also come under volatile category based on their chain length. If chain length increases eventually volatilability decreases. Table 9 shows volatile compounds of *A.indica* and their biological properties are mentioned in Table 3.

# Other Phytochemicals

Along with major phytochemical classes of *A.indica* as mentioned above, it has other classes including acetyl, acyclic alkane, alkylated phenol, aluminium glycinate, amine, amino acid, aminoglycoside, anthraquinone, benzene derivative, benzimidazole, benzoate glycoside, benzofurans, benzoxazole, carbazole, carboxy aldehyde, cardenolide, carotenoid, ceramide, cyclic imide, cyclitol, cyclohexanes, dicarboxylic acids, enkephaline peptide, ethanolamine. glucosides, glyceride, glycol, hetero-cycle, heterocyclic amine, imidazoles, imidazole derivative, imide, imino acid, indene, indoles, indole glycoside, lactones, leukotriene, limnoid, malate, modified dipeptide, n-substituted glycine, nucleosides, organosiloxine, peptides, phenols, phenyl ether, phenyl propionate, phthalic acids, piperidine, porphyrin, prostaglandins, purine, pyrazine derivative, pyridine, quinolines, salicylate, siloxane, sugar alcohol, sulfanilamide, tocopherol,

## Table 2: Quantitative estimation of different phytochemicals from various solvent extracts.

S.No	Plant part-Extract	Phytochemicals quantified	Equivalent standard	Quantity/dry weight	Reference
l	Leaves- Methanol	Flavanones	Naringenin	6.55% w/w	[81]
	Hydroalcoholic extract	Flavanones	Hesperidin	0.80 mg/g	[80]
	Leaves -Chloroform soluble fraction from hydroalcoholic extract	Flavanones	Hesperidin	0.097 mg/g	[80]
	Leaves -Butanol soluble fraction of hydroalcoholic extract	Flavanones	Hesperidin	0.61 mg/g	[80]
	Leaves -Butanol insoluble fraction of hydroalcoholic extract	Flavanones	Hesperidin	2.56 mg/g	[80]
	Leaves- Petroleum ether	Flavonoids	Catechin	8.38 mg/g	[83]
	Leaves- Chloroform	Flavonoids	Catechin	11.04 mg/g	[83]
	Leaves- Ethyl acetate	Flavonoids	Catechin	14.48 mg/g	[83]
	Leaves- Acetone	Flavonoids	Catechin	19.57 mg/g	[83]
)	Stem- Petroleum ether	Flavonoids	Catechin	6.83 mg/g	[83]
1	Stem- Chloroform	Flavonoids	Catechin	8.50 mg/g	[83]
2	Stem- Ethyl acetate	Flavonoids	Catechin	11.27 mg/g	[83]
3	Stem- Acetone	Flavonoids	Catechin	15.05 mg/g	[83]
ł	Stem- Methanol	Flavonoids	Catechin	13.78 mg/g	[83]
5	Root - Petroleum ether	Flavonoids	Catechin	7.68 mg/g	[83]
5	Root - Chloroform	Flavonoids	Catechin	9.08 mg/g	[83]
7	Root - Ethyl acetate	Flavonoids	Catechin	13.52 mg/g	[83]
8	Root - Acetone	Flavonoids	Catechin	16.55 mg/g	[83]
)	Root - Methanol	Flavonoids	Catechin	14.08 mg/g	[83]
)	Hydroalcoholic extract	Flavonoids	Quercetin	2.02 mg/g	[80]
l	Leaves -Chloroform soluble fraction from hydroalcoholic extract	Flavonoids	Quercetin	0.0156 mg/g	[80]
2	Leaves -Butanol soluble fraction of hydroalcoholic extract	Flavonoids	Quercetin	3.21 mg/g	[80]
3	Leaves -Butanol insoluble fraction of hydroalcoholic extract	Flavonoids	Quercetin	0.96 mg/g	[80]
1	Total plant -Ethanolic extract	Flavonoids	Rutin	8.75 mg/g	[82]
5	Leaves-Methanol	Flavonoids	Epicatechin Quercetin	29.896 mg/g; 1.76% w/w;	[78, 81, 83, 84
			catechin	67.87 mg/g 17.85 mg/g	
5	Leaves-Petroleum ether (Sequential extraction)	Flavonoids	Quercetin	81.00 mg/g	[83]
7	Leaves- Chloroform (Sequential extraction)	Flavonoids	Quercetin	30.00 mg/g	[83]
3	Leaves- Methanol (Sequential extraction)	Flavonoids	Quercetin	70.67 mg/g	[83]
)	Aqueous Methanol-70% ((Sequential extraction))	Flavonoids	Quercetin	6.67 mg/g	[83]
0	Leaves-Methanol	Phenols	Gallic acid	111.321 mg/g 10.89% w/w 373.54 mg/g	[78, 81, 84]
L	Leaves -Hydroalcoholic extract	Phenols	Gallic acid	7.9 mg/g	[80]
2	Leaves -Chloroform soluble fraction from hydroalcoholic extract	Phenols	Gallic acid	1.14 mg/g	[80]
3	Leaves -Butanol soluble fraction of hydroalcoholic extract	Phenols	Gallic acid	6.7 mg/g	[80]
1	Leaves -Butanol insoluble fraction of hydroalcoholic extract	Phenols	Gallic acid	0.58 mg/g	[80]
5	Total plant -Hydro alcoholic	Phenols	Gallic acid	1.63 mg/g	[76]
5	Total plant -Ethyl acetate fraction	Phenols	Gallic acid	7.21 mg/g	[76]
7	Total plant -Methanolic fraction	Phenols	Gallic acid	2.11 mg/g	[76]
3	Total plant -Hexane fraction	Phenols	Gallic acid	1.45 mg/g	[76]
)	Total plant -Ethanolic extract	Phenols	Tannic acid	9.27 mg/g	[76]
)	Leaves-Petroleum ether (Sequential extraction)	Phenols	Gallic acid	20.00 mg/g	[76]

Tabl 2	: Continued				
41	Leaves-Chloroform (Sequential extraction)	Phenols	Gallic acid	10.00 mg/g	[84]
42	Leaves-Methanol (Sequential extraction)	Phenols	Gallic acid	306.67 mg/g	[84]
43	Leaves-Aqueous Methanol (70%) (Sequential extraction)	Phenols	Gallic acid	60.00 mg/g	[84]
44	Leaves- Petroleum ether	Phenols	Gallic acid	10.56 mg/g	[83]
45	Leaves- Chloroform	Phenols	Gallic acid	13.85 mg/g	[83]
46	Leaves- Ethyl acetate	Phenols	Gallic acid	16.15 mg/g	[83]
47	Leaves- Acetone	Phenols	Gallic acid	20.03 mg/g	[83]
48	Leaves- Methanol	Phenols	Gallic acid	18.68 mg/g	[83]
49	Stem- Petroleum ether	Phenols	Gallic acid	7.98 mg/g	[83]
50	Stem- Chloroform	Phenols	Gallic acid	11.36 mg/g	[83]
51	Stem- Ethyl acetate	Phenols	Gallic acid	13.57 mg/g	[83]
52	Stem- Acetone	Phenols	Gallic acid	15.66 mg/g	[83]
53	Stem- Methanol	Phenols	Gallic acid	14.58 mg/g	[83]
54	Root - Petroleum ether	Phenols	Gallic acid	8.03 mg/g	[83]
55	Root - Chloroform	Phenols	Gallic acid	12.05 mg/g	[83]
56	Root Ethyl acetate	Phenols	Gallic acid	14.71 mg/g	[83]
57	Root - Acetone	Phenols	Gallic acid	17.34 mg/g	[83]
58	Root - Methanol	Phenols	Gallic acid	15.03 mg/g	[83]
59	Leaves- Methanol	Saponins	-	16.1 mg/g	[78]

toluidines and tyronine. The respective compound names of each class are available in Table 3. Miscellaneous compounds are mentioned at the end of Table 3 (Serial number starting from 218 to 250).

## Pharmacological Properties Safety

Safety is a measurement of toxic levels of plant extracts during the toxicity studies. In case of reports available on safety aspect of this plant extracts, aqueous ethanolic extract of 2000 mg/kg and 3000 mg/kg didn't show any abnormalities in mice and rats respectively.<sup>[221]</sup> The chloroform and n-butanol soluble/insoluble fractions of hydroalcoholic extract of leaves didn't show any behavioral changes and mortality in rats up to 4000 mg/kg.<sup>[80]</sup> Dosage studies ranging from 100-2000 mg/kg for 14 days with methanolic extract/fraction of leaves not exhibited any changes in biochemical parameters (SGOT, SGPT, ACP, ALP, LDH, creatinine, glucose, protein and uric acid,), haematological issues (hemoglobin, packed cell volume, white blood cell and erythrocyte sedimentation rate), organs weight (heart, brain, spleen, liver and kidney) and mortality.<sup>[81,222]</sup> Godipurge et al. (2015)<sup>[82]</sup> have reported that polyphenolic extract (5000 mg/kg) didn't show any adverse effcts such as mild diarrhea, depression and weight loss. Similarly, toxicity study in rats with different doses of methanolic extract (Phase I: 10-1000 mg/kg and phase II: 1000-5000 mg/kg) for 12 hrs have not shown any mortality and toxic symptoms. [84] Shirwaikar et al. (2004)[223] have reported that intra peritoneal administration of ethanolic extract (5000 mg/kg) didn't produce any toxic effects in rats even after 72 hrs.

#### Antioxidant activity

Several *in vitro* studies indicated that *A.indica* has antioxidant/radical scavenging potentiality. In final concentration of 0.640 mg/mL of hydro ethanolic extract (70%), fractions of methanol, ethyl acetate and hexane of total plant on DPPH radical showed scavenging of 51.09 % (IC<sub>50</sub>: 0.61632 mg/mL), 66.5 % (IC<sub>50</sub>: 0.5542 mg/mL), 52.09% (IC<sub>50</sub>: 0.19125 mg/mL) and 63.19 % (IC<sub>50</sub>: 0.24914 mg/mL) respectively, whereas the same concentration of ascorbic acid shown 94.74%  $(IC_{co}: 0.022 \text{ mg/mL}).^{[76]}$  The same DPPH mitigation property of methanolic extract of leaves has been tested by Badami and Channabasavaraj, (2007),<sup>[224]</sup> Shanmugapriya et al. (2011),<sup>[225]</sup> Sanseera et al. (2012),<sup>[99]</sup> Selvamani (2015)<sup>[83]</sup> and Ravi et al. (2017).<sup>[78]</sup> Among their results, the good scavenging property was noticed by Ravi et al. (2017) [78] (62.948 % at 0.050 mg/mL, IC<sub>50</sub>: 0.028 mg/mL) whereas, the standard ascorbic acid had IC<sub>50</sub>: 0.004 mg/mL. All the effective doses/IC<sub>50</sub> values of leaves and other plant parts on DPPH is summarized in Table 12. An insight into ABTS radical assay for methanol, chloroform and hexane extracts of leaves showed the IC<sub>50</sub> values of 6.370, 6.310 and 6.130 mg/mL respectively, whereas the standard trolax has 1.320 mg/mL.<sup>[99]</sup> The other study also revealed potential ABTS scavenging property of methanolic extract of leaves, root and stem with the IC<sub>50</sub>: 0.005, 0.024 and 0.014 mg/mL respectively, whereas the standard ascorbic acid and rutin showed 0.011 mg/mL and 0.52  $\mu g/mL$  respectively.  $^{\scriptscriptstyle [224]}$  In a subsequent study, Selvamani, (2015)<sup>[83]</sup> found IC<sub>50</sub> concentrations of acetone extract of leaves (0.252 mg/mL), root (0.288 mg/mL), stem (0.323 mg/mL) and compared with standard BHT (0.225 mg/mL). Similarly, A.indica leaf, root and stem extracts/fractions scavenge the hydrogen peroxide, superoxide radicals, nitric oxide, and metal ions (iron and molybdenum); protect the hydroxyl radicals induced sugar damage and lipid peroxidation. [78, 79, 83, 99, 126, 224, 226]

#### Anticancer activity

Anticancer property of *A.indica* for methanolic extract of leaves on NCI-H187-small cell lung cancer cell lines using Resazurin cell viability fluorescent assay revealed good activity with  $IC_{50}$  concentration of 25.00 µg/mL, but it has no potent activity on KB-oral cavity and MCF7-

## Ravi, et al.: A Comprehensive Review on Acalypha indica

S.No	ist of phytochemical compounds isc. Compound Name	Туре	Method	Plant source	Pubchem ID	<b>Biological Property</b>	References
1	3,8-Nanodiene-2-one,(E)-	Acetyl	GC-MS	TP	-	-	[75, 43]
2	2-methyl tricosane	Acyclic Alkane	GC-MS	TP	-	-	[85]
3	Hexanal	Aldehyde	GC-MS	TP	CID: 6184	Incresed the anxiety	[85, 86]
4	Acalyphin amide	Alkaloid	MLCCC	L-Inf	CID: 102286669	-	[87]
5	Epiacalyphin amide cycloside	Alkaloid	MLCCC	L-Inf	-	-	[87]
6	Ar-Acalyphidone	Alkaloid	MLCCC	L-Inf	CID: 102286671	-	[87]
7	Rescinnamine	Alkaloid	HR-LC-MS	L	CID: 5280954	Antihypertensive action	[78, 88]
8	Pergolide sulfone	Alkaloid	HR-LC-MS	L	CID: 155750	Acted as dopamine receptor inhibitor	[78, 89, 90]
9	Lupinine	Alkaloid	HR-LC-MS	L	CID:91461	Anti coagulant in the form of artificial polymer. Binds to the nicotinic and muscarine acetylcholine receptors	[78, 91, 92]
10	Ambelline	Alkaloid	HR-LC-MS	L	CID:25092366	Antiproliferative	[78]
11	2-acetyl-4-methoxy-1,2-dimethyl-6- oxo-3-{[3,4,5-triacetyl-6-(2- oxopropyl)oxan-2-yl]oxy}-1,2,3,6- tetrahydropyridine-3-carbonitrile	Alkaloid	C.Chromat	L-Flw	-		[93]
12	2-acetyl-5-methoxy-1-methyl-6- oxo-3-{[3,4,5-triacetyl-6-(2- oxopropyl)oxan-2-yl]oxy}-1,2,3,6- tetrahydropyridine-3-carbonitrile	Alkaloid	C.Chromat	L-Flw			[93]
13	2-hydroxy-5-methoxy-1-methyl-6- oxo-3-{[3,4,5-trihydroxy-6- (hydroxymethyl)oxan-2-yl] oxy}-1,2,3,6-tetrahydropyridine-3- carbonitrile	Alkaloid	C.Chromat	L-Flw	-	-	[93]
14	Acalyphin	Alkaloid	MLCCC/ C.Chromat	L-Flw	CID: 49787014	-	[87, 93]
15	Epiacalyphin	Alkaloid	MLCCC	L-Inf	CID: 102286666	-	[87]
16	Noracalyphin	Alkaloid	MLCCC	L-Inf	CID: 102286667	-	[87]
17	Epinoracalyphin	Alkaloid	MLCCC	L-Inf	CID: 102286668	-	[87]
18	Seco-Acalyphin	Alkaloid	MLCCC	L-Inf	CID: 102286670	-	[87]
19	Ethyl tetradecane	Alkane	GC-MS	TP	CID: 41311	-	[85]
20	2-methyl pentadecane	Alkane	GC-MS	TP	CID:15267	-	[85]
21	7-butyl docosane	Alkane	GC-MS	TP	-	-	[85]
22	7-hexyl eicosane	Alkane	GC-MS	TP	-	-	[85]
23	2,2-dimethyl dodecane	Alkane	GC-MS	TP	-	-	[85]
24	Decane	Alkane	GC-MS	TP	CID: 15600		[85]
24	Tricosane	Alkane	GC-MS	TP	CID: 12534	_	[83]
25 26	Hexatriacontane	Alkane	GC-MS	TP	CID: 12334 CID: 12412		[94]
26 27	Octacosane	Alkane	GC-MS GC-MS	L	CID: 12412 CID: 12408		[85]
						-	
28	Tridecane	Alkane	GC-MS	TP	CID: 12388	-	[85]
29	Heptacosane	Alkane	GC-MS	L	CID: 11636	-	[49]
30	Octadecane	Alkane	GC-MS	TP	CID:11635	-	[85]

Table 3: List of phytochemical compounds isolated and identified by different chromatographic tenchniques in A.indica

Table	3: Continued						
31	2,4-bis(1,1-dimethylethyl) phenol	Alkylated phenol	GC-MS	L	CID:528937	-	[95]
32	Dihydroxyaluminumaminoacetate	Aluminium glycinate	HR-LC-MS	L	CID: 18502861	Used in the treatment of stomach ulcers and gastritis	[78, 96]
33	3-Methylglutarylcarnitine	Amine	HR-LC-MS	L	CID: 128145	-	[78]
34	Cysteine	Amino acid	GC-MS	TP	CID: 5862	-	[75]
35	Amikacin	Aminoglycoside	HR-LC-MS	L	CID: 37768	Antibacterial agent used to treat urinary tract infections, Intra abdominal infections, Meningitis, Pneumonia, Sepsis, Multi drug resistant TB. Inactivates aminoglycoside- inactivating enzyme.	[78, 97]
36	2-Methylanthraquinone	Anthraquinone	NM	NM	CID: 6773	-	[98-100]
37	Benzene, 1,2,3-trimethyl	Benzene derivative	GC-MS	L	CID: 10686	-	[49]
38	O-Toluenesulfamide	Benzene derivative	HR-LC-MS	L	CID: 10334	-	[78]
39	Emedastine	Benzimidazole	HR-LC-MS	L	CID: 3219	Anti-histamine and H1 receptor inhibitor.	[78, 101]
40	Mebeverine metabolite (Veratric acid glucuronide)	Benzoate glycoside	HR-LC-MS	L	-	-	[78]
41	2-methyl-3-phenyl-1H-Indole	Benzofuran	GC-MS	ТР	CID: 282402	-	[102]
42	Loliolide	Benzofuran	GC-MS	TP	CID: 100332	-	[94]
43	Coumaran	Benzofuran	GC-MS	ТР	CID: 10329	Acetylcholine esterase inhibitor	[85, 103]
44	Dihydroactinidiolide	Benzoxazole	GC-MS	ТР	CID: 6432173	-	[94]
45	Desmethylondansetron	Carbazole	HR-LC-MS	L	CID: 10891224	-	-
46	Isocyclocitral	Carboxy aldehyde	GC-MS	L		-	[95]
47	Peruvoside	Cardenolide	HR-LC-MS	L	CID: 12314120	Cardioprotective	[78, 104]
48	β-ionone	Carotenoid	GC-MS	ТР	CID: 638014	Anti-carcinogenic agent	[85, 105]
49	C16 Sphinganine	Ceramide	HR-LC-MS	L	CID: 5283572	-	[78]
50	Ferulic acid	Coumaric acid	RP-HPLC	L	CID: 445858	Antioxidant; Anticancer; Anticardiovascular disease; Skin diseases treatment and Antidiabetic.	[84,106]
51	4-Methyldaphnetin	Coumarin	HR-LC-MS	L	CID: 5355836	Free radical scavenger, 5-lipoxygenase inhibitor, Anticancer, ERK/MAPK signalling inhibitor. Apoptosis inducer.	[78, 107-109]
52	3,3' Methylene bis (4-hydroxyl coumarin)	Coumarin	RP-HPLC	L	CID:54676038	Anticancer	[84, 110]
53	Succinimide	cyclic imide	NM	NM	CID: 11439	-	[111]
54	Quebrachitol	cyclitol	C.Chromat	L	CID: 151108	-	[98, 100]
55	Picrotin	Cyclohexane	HR-LC-MS	L	CID: 442291	Activates human bitter receptors (hTAS2Rs); Blocks the Homomeric Glycine Receptors	[78, 112]

Table	3: Continued						
56	Dicyclomine	Cyclohexane	HR-LC-MS	L	CID: 3042	M1 muscarinic antagonist. Used to treat irritable bowel syndrome, Ulcerative colitis; Antispasmodic and Anticholinergic	[78]
57	Traumatic acid	Dicarboxylic acid	HR-LC-MS	L	CID: 5283028	Antioxidant; Enhances collagen biosynthesis; Growth factor for algae	[78, 113-114]
58	L-2-Aminoadipic acid	Dicarboxylic acid	HR-LC-MS	L	CID: 92136	Increases the protein synthesis and inhibits the autophagy in myotubes	[115]
59	Enkephaline, (D-Ala)2-Leu	Enkephaline peptide	HR-LC-MS	L	-	-	[78]
60	2-propenoic acid, 3-[5-acetyl-2,2- dimethylcyclopentyl], methyl ester,[1a(E),5a]-	Ester	GC-MS	L	-	-	[78]
61	Pentadecanoic acid, 14-methyl-, methyl ester	Ester	GC-MS	L	CID:21205	-	[78]
62	10-octadecenoic acid, methyl ester	Ester	GC-MS	L	CID:5364425	-	[78]
63	Heptadecanoic acid, 16-methyl- methyl ester	Ester	GC-MS	L	-	-	[78]
64	Hexadecanoic acid, 14-methyl, methyl ester	Ester	GC-MS	L	-	-	[78]
65	Nonadecanoic acid, 18-oxo, methyl ester	Ester	GC-MS	L	-	-	[78]
66	14-methylhexadecanoic acid, picolinyl ester	Ester	GC-MS	L	-	-	[78]
67	9-octadecenoic acid [Z], 2-hydroxy- 1-(hydroxymethyl) ethyl ester	Ester	GC-MS	L	CID:5319879	-	[78]
68	Hexanedioic acid, bis(2-ethylhexyl) ester	Ester	GC-MS	TP	-	-	[94]
69	Eicosatrienoic acid methyl ester	Ester	GC-MS	ТР	CID:6421258	-	[85]
70	Methyl arachidate	Ester	GC-MS	TP	CID:14259	-	[85]
71	Trifluoro acetic acid, n-heptadecyl ester	Ester	GC-MS	TP	-	-	[85]
72	Trifluoro acetic acid, n-octadecyl ester	Ester	GC-MS	TP	CID:522719	-	[85]
73	Propionylglycine methyl ester	Ester	HR-LC-MS	L	-	-	[78]
74	GPEtn(18:0/22:6(4Z,7Z,10Z,1 3Z,16Z,19Z))	Ester	HR-LC-MS	L	-	-	[78]
75	GPCho(16:0/3:1(2E))	Ester	HR-LC-MS	L	-	-	[78]
76	Ethyl ester of hexadecanoic acid	Ester	GC-MS	L	CID:12366	-	[95]
77	Hexadecanoic acid methyl ester	Ester	GC-MS	TP	CID: 8181	-	[85]
78	1,2-Benzenedicarboxylic acid, diisooctyl ester	Ester	GC-MS	L	CID: 33934	-	[116]
79	Choline	Ethanolamine	HR-LC-MS	L	CID:6209	A precursor for acetylcholine; maternal immune stimulator at lactation period; Used in PET imaging for cancer tracing.	[117, 118]
80	1,1-Diethoxy butane	Ether	GC-MS	L	CID:77225	-	[119]
81	1,1-Diethoxy pentane	Ether	GC-MS	L	CID:77223	-	[119]

Table 3	3: Continued						
82	1,1-Diethoxy hexane	Ether	GC-MS	L	CID: 77224	-	[119]
83	1,1,3-Triethoxy propane	Ether	GC-MS	L	CID: 24624	-	[119]
84	1-Monolinoleoylglycerol trimethylsilyl ether	Ether	GC-MS	L	CID: 5366692	-	[119]
85	Ethyl pentanate	Fatty acid	GC-MS	L	CID:10882	-	[119]
86	Ethyl decanoate	Fatty acid	GC-MS	L	CID:8048	-	[119]
87	9,12-Octadecadienoic acid(Z,Z)-	Fatty acid	GC-MS	L/ TP	CID: 5280450		[49, 94]
88	Oleic acid	Fatty acid	GC-MS	L	CID: 445639	-	[116]
89	Tetradecanoic acid	Fatty acid	GC-MS	L	CID: 11005	-	[20, 116,120, 121]
90	Octadecanoic acid	Fatty acid	GC-MS	L	CID: 5281	-	[49]
91	n-Hexadecanoic acid	Fatty acid	GC-MS	L/ TP	CID: 985	-	[49, 78, 102, 116, 119]
92	13-Oxo-ODE	Fatty acid	HR-LC-MS	L	CID: 6446027	PPARa agonist, Ameliorates dyslipidemia and hepatic steatosis; Reduce mucosal damage; Down regulates the inflammation and inhibitor of LOX isozymes.	[78, 122, 123]
93	N-(2hydroxyethyl)palmitamide (Propylene glycol)	Fatty acid	HR-LC-MS	L	CID: 4671	Antiinflammatory; Antitoxic; Anti- traumatic shock; Antianaphylactic; AntiSerotonine; Antiviral; Induce DNA and RNA synthesis.	[78, 124, 125]
94	Lactone of PGF-MUM	Fatty acid	HR-LC-MS	L	-	-	[78]
95	9,12,15-Octadecatrienoic acid	Fatty acid	GC-MS	TP		-	[102]
					CID: 5282822		
96	9,12,15-Octadecatrienoic acid, (Z,Z,Z)-	Fatty acid	GC-MS	L	CID: 5280934	-	[49, 116, 126]
97	Isodecane	Fatty acyl	GC-MS	TP	CID: 13379	-	[85]
98	9-Nanodecene	Fatty acyl	GC-MS	TP	CID: 5364436	-	[102]
99	8,9-Dihydroxy-5,11,14eicosatrienoic acid	Fatty acyl	HR-LC-MS	L	CID:5312971	-	[78]
100	2R-hydroperoxy- 9Z,12Z,15Zoctadecatrienoic acid	Fatty acyl	HR-LC-MS	L	CID:16061057	-	[78]
101	2,10-Dihydroxy-4,6,8decatriynoic acid	Fatty acyl	HR-LC-MS	L	CID: 5312795	-	[78]
102	9-Tricosene	Fatty acyl	GC-MS	ТР	CID: 6385060	Stimulates female mating in Aphrodisiac of spider	[94, 127]
103	9,12,15 octadecatrienal	Fatty acyl	GC-MS	TP	CID: 5283384	-	[85]
104	9Z,12Z,15E-Octadecatrienoic acid	Fatty acyl	HR-LC-MS	L	CID:5312504	-	[78]
105	6,11-Octadecadiynoic acid	Fatty acyl	HR-LC-MS	L	CID:5312660	-	[78]
106	5,6-DiHETrE-EA	Fatty acyl	HR-LC-MS	L	CID:16061178	-	[78]
107	3,4-Tetradecadienoic acid	Fatty acyl	HR-LC-MS	L	CID: 5312405	-	[78]

Table 3	8: Continued						
108	13-Tetradecen-2,4-diyn-1-ol	Fatty acyl	HR-LC-MS	L	CID: 5283270	-	[78]
109	13-Keto-9Z,11E,15Z octadecatrienoic acid	Fatty acyl	HR-LC-MS	L	CID: 11426350	-	[78]
110	12-Hydroxy-10-octadecynoic acid	Fatty acyl	HR-LC-MS	L	CID: 5312859	-	[78]
111	N-octacosonal	Fatty alcohol	NM	NM	CID:68406	Antinociceptive; Anti-Inflammation; Antiparkinson; Improve reproductive performance.	[78]
112	Tetradecen-1-ol	Fatty alcohol	GC-MS	TP	CID: 120110	-	[85]
113	Hexenol	Fatty alcohol	GC-MS	TP	CID: 5281167	-	[85]
114	Docosanol	Fatty alcohol	GC-MS	TP	CID: 12620	-	[127]
115	1-Eicosanol	Fatty alcohol	GC-MS	TP	CID: 12404	-	[127]
116	1-Hexadecanol	Fatty alcohol	GC-MS	L	CID: 2682	-	[95]
117	1-Triacontanol	Fatty alcohol	GC-MS	TP	CID: 68972	Anti-cancer	[127,128]
118	Octacosanol	Fatty alcohol	GC-MS	TP	CID: 68406	-	[127]
119	Palmitaldehyde	Fattyaldehyde	GC-MS	TP	CID: 984	-	[85]
120	Catechin	Flavanoids	HR-LC-MS	L	CID: 73160	Antioxidant; Neuroprotective; Neurodegenerative disorders; Anticancer; Reduce mitochondrial dysfunction; Anti- diabetic; Inhibitor of polyphenoloxidase and melanosis; Anti- inflammation; Used to treat dry eye disease; Activate brown adipose tissue and Ameliorates graft-versus-host disease.	[78, 129-135]
121	Dehydrovariabilin	Flavonoid	HR-LC-MS	L	CID: 624785	Antiprion	[78,136]
122	Quercetin 3-0-β-D-glucoside	Flavonoid	C.Chromat	TP	CID:44259136	Antiproliferative; Anti- cancer; Wound healing; Antimicriobial; Acute myocardial ischemia protection; Antioxidant; Melanin inhibition; $\beta_2$ - adrenergic signaling; Anti-aging and Anti- inflammatory.	[137-145]

Table 3	3: Continued						
123	Rutin	Flavonoid	C.Chromat/ RP-HPLC	TP/L	CID: 5280805	Antioxidant; Anti- Neuroinflammation; Sedative; Anticonvulsant; Antialzheimer; Treatment for hyperkinetic movement disorder; Antidepressant; Antiischemic; Antigeressant; Antiischemic; Antiocceptive; Antifungal; Anti-arthritic; Antifungal; Anti-arthritic; Antidiabetic; Anti hypercholesterolemic; Antijalatelet aggregatory; Antiulcer; Antiosteoporotic; Antiosteopenic; Antiosteopenic; Antiosteopenic; Antiosteopenic; Antifungal; Larvicidal; Antifungal; Larvicidal; Antifungal; Larvicidal; Antimalarial; Antiretroviral; Antiviral;Atopic dermatitis; Immune effects; Anti fatigue; Cardioprotective; Nephroprotective; Hepatoprotective and Wound healing.	[137,146]
124	Nicotiflorin	flavonoid	MLCC	L-F	CID: 5318767	-	[147]
125	Biorobin	flavonoid	MLCC	L-F	CID: 15944778	-	[147]
126	Clitorin	flavonoid	MLCC	L-F	CID: 11592917	-	[147]
127	Mauritianin	flavonoid	MLCC	L-F	-	-	[147]
128	Kaempferol	Flavonoid	GC-MS	TP	CID:5280863	Antioxidant activity; Apoptosis inducer; Anticancer; Anti- angiogenesis; Antiinflammation; Anticarcinogenic; Antimicrobial; Neuroprotective	[75, 148-151]
129	Naringenin	Flavonoid	RP-HPLC	L	CID:932	Antioxidant; Antimicrobial; Antiinflammatory; Antidiabetic; Anticancer; Neuroprotective and Cardioprotective	[84, 152]
130	β-D-glucoside	Glucoside	NM	NM	-	Antitumor and Antinecrosis	[153]
131	3-O-methyl-D-glucose	Glucoside	GC-MS	L	CID: 8973	-	[116, 154-156]
132	1-Hexadecanoyl-sn-glycerol	Glyceride	HR-LC-MS	L	CID:3084463	Food additive	[78,157]

Table 3	8: Continued						
133	N-Tris[hydroxymethyl]methyl2- aminoethanesulfonic acid	Glycol	HR-LC-MS	L	CID: 81831	Antiinflammation; Effect on delayed hypersensitivity.	[78,124,158]
134	Triacetonamine	heterocycle	NM	NM	CID:13220	-	[98]
135	Piperidine-2,5-dione	Heterocyclic amine	GC-MS	L	CID: 533930	-	[49]
136	Gallic acid	Hydroxybenzoate	RP-HPLC	L	CID: 370	Antioxidant; Anticancer; Antimelanogenic; Antiinflamatory; Antimicrobial; Antiviral; Antiallergic; Neuroprotective; Anti diabetic; Anti diabetic; Cardioprotective; Anti Alzheimer; Nephroprotective and Hepatoprotective.	[84, 159- 162]
137	Syringic acid	Hydroxybenzoate	RP-HPLC	L	CID:10742	Antidiabetic; Antioxidant; Anti acute pancreatitis.	[84,163,164]
138	Caffeic acid	hydroxycinnamic acid	RP-HPLC	L	CID:689043	Anticancer; Antioxidant; Radical scavenger; AntiInflammatory; AntiHuman Immunodeficiency Virus (HIV) and Antimicrobial.	[84, 165, 166]
139	Imidazole, 4-fluoro-5- hydroxyazomethyl	Imidazole	GC-MS	L	CID: 574814	-	[78]
140	Ondansetron	Imidazole	HR-LC-MS	L	CID: 4595	Prevents vomiting and nausea during chemo therapy and radio therapy of cancer; Used in gastroenteritis.	[167, 168]
141	Clotrimazole	imidazole derivative	TLC	L	CID: 2812	Malaria chemotherapy; Antimicrobial and Antimycotic	[169- 171]
142	Ethosuximide M5	Imide	HR-LC-MS	L	-	-	[78]
143	Proline, 3,4-didehydro-	Imino acid	GC-MS	L/TP	CID:25202244	-	[43, 75]
144	Sulindac sulfide	Indene	HR-LC-MS	L	CID: 5352624	Non-steroidal anti- inflammatory drug targets the COX-2; Potential anticancer drug works on various upregulated (EGR- 1, ATF3, NF- $k$ B, E-cadherin, NAG-1, p21 and CHOP ) and down regulated ( $\beta$ -Catenin, NF- $k$ B, EGFR, PDE5, Nesprin-2, Cyclin D1) targets of lung, brest, colon, head/ neck, ovarian, gastric, prostate and pancreatic cancers	[172]

Table 3	3: Continued						
145	2-formyl-5,7dimethyl-1,2,3,4- tetrahydropyrimido(3,4-a) indole.	Indole	GC-MS	L		-	[78]
146	2-(4-methylphenyl) indolizine	Indole	GC-MS	L	CID:346948	-	[95]
147	Trandolapril glucuronide	Indole glycoside	HR-LC-MS	L	CID:92023960	-	[78]
148	13-hexyloxacyclotridec10-en-2-one	Lactone	GC-MS	L		-	[78]
149	2-Methyl-7-phenylindole	Lactone	GC-MS	L	CID: 610181	-	[95]
150	5-Methyl-2-phenylindolizine	Lactone	GC-MS	L	CID: 610180	-	[95]
151	Deoxykhivorin	Lactone	HR-LC-MS	L	CID: 6708722	Neuroprotective agent.	[173]
152	Leukotriene F4	Leukotriene	HR-LC-MS	L	CID: 5280938	Causes brochoconstriction.	[174]
153	Heudelottin C	Limnoid	HR-LC-MS	L	CID: 4270081	Filovirus inhibitor	[175]
154	Dimethyl citraconate	Malate	GC-MS	L	CID:5355715	-	[126]
155	Aurantiamide	Modified dipeptide	NM	NM	CID: 185904	-	[111]
156	Dimethylglycine	N-substituted glycine	HR-LC-MS	L	CID: 673	N-methyl-d- aspartate receptor (NMDAR) inhibitor; Antipsychotic activity. Effectively work on Autism , Pervasive developmental disorder, Allergies, Respiratory disorders, Inflammation, Cancer, Epilepsy, Alcoholism and Mitochondrial diseases.	[176-178]
157	Cytidine	Nucleoside	GC-MS	L	CID:6175	Antidepressant-like effects	[126, 179]
158	3-Deoxyguanosine	Nucleoside	HR-LC-MS	L	CID: 165138	Antitumor	[180]
159	Hexadecamethyl-heptasiloxane	organosiloxine	GC-MS	ТР	CID:10912	-	[85]
160	Val TrpThr	Peptide	HR-LC-MS	L	-	-	[78]
161	Tyr Phe Tyr	Peptide	HR-LC-MS	L	-	-	[78]
162	Lys His Cys	Peptide	HR-LC-MS	L	-	-	[78]
163	His Ala Ala	Peptides	HR-LC-MS	L	-	-	[78]
164	Phenol,24 BIS(1,1-Dimethylethyl)	Phenol	GC-MS	L	CID: 7311	-	[181]
165	3- Methylphenol	Phenol	GC-MS	L	CID:342	-	[95]
166	Benzenemethanol, 2-(2aminopropoxy)-3-methyl	Phenyl ether	HR-LC-MS	L	CID: 93285	-	[78]
167	Ibuprofen	Phenyl propionate	HR-LC-MS	L	CID: 3672	Relaxes the Menstrual cramps, Arthritis, Fever, Inflammation, Cold, Toothaches; Lowers the blood pressure; Anticancer; Antialzheimers and Antiparkinsons	[182]
168	Fenoprofen glucuronide	Phenyl propionate glycoside	HR-LC-MS	L	-	Urinary metabolite of Fenoprofen, a pain killer and Antiarthritic agent	[183]
169	Di-(2-ethylhexyl)phthalate	Phthalic acid	GC-MS	L	-	-	[126]
170	Didodecyl phthalate	Phthalic acid	GC-MS	L	CID: 17082	-	[119]

Table 3	8: Continued						
171	Terephthalic acid	Phthalic acid	HR-LC-MS	L	CID: 7489	Used in manufacture of polyester fibers, clothing and industrial filaments. Its co polymers used in packing of foods, water, edible oils, bevereges.	[184]
172	2,6-Piperidinedicarboxylic acid	Piperidine	HR-LC-MS	L	CID:557515	-	[78]
173	Harderoporphyrin	Porphyrin	HR-LC-MS	L	CID: 3081462	An intermediate of heme biosynthesis, elevated levels observed in harderoporphyria.	[185]
174	PGG2	prostaglandins	HR-LC-MS	L	CID: 5280883	Unstable prostaglandin G2, acts as vasodepressor agent, involved in platelet aggregation and release.	[186, 187]
175	4-Amino-3-methoxypyrazolo[3,4- d]pyrimidine, -	Purine	GC-MS	ΤР	CID:596791	-	[43, 75]
176	5,10-Diethoxy-2,3,7, 8-tetrahydro- 1H, 6H-dipyrrolo[1,2-a;1',2'-d] pyrazine	Pyrazine derivative	GC-MS	ТР	CID: 551125	-	[94]
177	Tropicamide	Pyridine	HR-LC-MS	L	CID:5593	Anticholinergic drug used as Pupillary dilation of eye.	[188]
178	2-Dimethylaminopyridine	Pyridine derivative	GC-MS	L	CID: 21885	-	[95]
179	4-(3-Pyridyl)-3-butenoic acid	Pyridine derivative	HR-LC-MS	L	CID:5478892	-	[78]
180	2,3-Dihydro-3,5-dihydroxy-6- methyl-4H-pyran4-one	Pyrone	GC-MS	L	CID:119838	-	[119]
181	Bilirubin	Pyrrole	HR-LC-MS	L	CID: 5280352	Antibacterial	[189]
182	1H-Pyrrole-2,5-dione,1- ethenyl-	Pyrroles	GC-MS	L/TP	CID: 24358	-	[43,75]
183	2,5-Pyrrolidinedione, 1-methyl-	Pyrrolidine	GC-MS	L	CID:21232168	-	[116]
184	Peucenin	Quinolines	HR-LC-MS	L	CID: 68477	-	[78]
185	Orthothymotinic Acid	Salicylate	HR-LC-MS	L	CID: 11052	-	[78]
186	Clotrisiloxane, Hexamathyl-	Siloxane	GC-MS	L	CID: 10914	-	[181]
187	27-Nor-5b- cholestane3a,7a,12a,24,25-pentol	Steroid	HR-LC-MS	L	CID:21252278	-	[78]
188	Beta-sitosterol	Steroid	GC-MS	TP	CID: 222284	Uterotrophic agent; Antifertility; Antiinflammatory and Anti-microbial	[102, 190,191]
189	Campesterol	Steroid	GC-MS	TP	CID: 173183	-	[102]
190	Stigmasterol	Steroid	GC-MS	TP	CID: 5280794	Antiosteoarthritic; Antihyper- cholestrolemic; Antitumor; Hypoglycemic; Antioxidant; Antiinflammatory; Antimutagenic	[102, 192]
191	3beta,6alpha,7alphaTrihydroxy- 5beta-cholan-24oic Acid	Steroid	HR-LC-MS	L	CID:5283822	-	[78]
192	4-C-methyl-myo-inositol	Sugar alcohol	GC-MS	L	CID: 244581		[119]

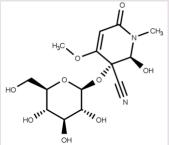
Table 3	3: Continued						
193	Sulfamerazine	Sulfanilamide	HR-LC-MS	L	CID: 5325	-	[78]
194	Potassium brevifolincarboxylate	Tannin	C.Chromat	TP	CID: 101933031	-	[137]
195	Acaindinin	Tannin	C.Chromat	TP	CID: 101933032	-	[137]
196	l-O-galloyl-β-D-glucose	Tannin	C.Chromat	TP		Anti-inflammatory; MyD88/NF-κ B and MyD88/MAPK signalling pathways inhibitor and Antidiabetic.	[137, 193,194]
197	1,2,3,6-tetra-O-galloyl·β-D-glucose	Tannin	C.Chromat	ТР	-	Antiviral and Anticancer	[137, 195- 197]
198	Corilagin	Tannin	C.Chromat	ТР	CID:73568	Anti-inflammatory; Antioxidant;. Anti- hyperalgesic; Antiulcer; Hepatoprotective; Anti-proliferation; Anti-tumour;	[137, 198- 203]
199	Geraniin	Tannin	C.Chromat	TP	CID:3001497	Antioxidant; Anti- tumour; Anti-infective; Antihyperglycemic and Anticancer activity	[137, 204- 208]
200	Acetonylgeraniin A	Tannin	C.Chromat	TP	-	-	[137]
201	Euphormism M2	Tannin	C.Chromat	TP	-	-	[137]
202	Repandusinic acid A	Tannin	C.Chromat	TP	CID: 14483070	-	[137]
203	Chebulagic acid	Tannin	C.Chromat	ТР	CID: 442674	Antibiabetic	[137, 209]
204	Tri-O-methylellagic acid	Tannin	NM	NM	-	Antimicrobial	[210]
205	Larixol Acetate	Terpene	HR-LC-MS	L	CID: 4615087	Transient receptor potential cation channel 6 (TRPC6) inhibitor	[78, 211]
206	Phytol	Terpene	GC-MS	L/ TP	CID: 5280435	Antischistosomal; Anxiolytic-like effects	[49,85,94, 116, 119, 212, 213]
207	Squalene	Terpene	GC-MS	L	CID: 638072	Protection of cyclophosphamide- induced toxicity	[49,119, 214]
208	Isophytol	Terpene	GC-MS	TP	CID: 10453	-	[102]
209	Pinane	Terpene	GC-MS	L	CID: 10129	-	[95]
210	Linalool	Terpene	GC-MS	ТР	CID: 6549	-	[85]
211	Tunaxanthin J/ Chiriquixanthin B	Terpene	HR-LC-MS	L	CID: 16061201	-	[78]
212	Methoprene (S)	Terpene	HR-LC-MS	L	CID: 1711973	Accelerates sexual maturation in male and female	[78, 215]
213	Elephantopin	Terpene	HR-LC-MS	L	CID: 442206	Anticancer	[78, 216]
214	Flavoxanthin	Terpenoid	HR-LC-MS	L	CID:5281238	Food additive and food colouring agent	[217]
215	Vitamin E	Tocopherol	GC-MS	L/TP	CID: 14985	Neuroprotection; Antioxidant;	[102,119, 218]
216	Pararosaniline	Toluidines	HR-LC-MS	L	CID: 11292	Cells staining	[78, 219]
217	Thyroacetic acid	Tyronine	HR-LC-MS	L	-	-	[78]
218	Propanenitrile,3-(5-diethylamino-1- methoxy-3-pentynyloxy)-	-	GC-MS	ТР	-	-	[75]
219	1-oxaspiro [2,5] octane, 5,5-dimethyl-4-(3-methyl-1,3- butadienyl)-	-	GC-MS	L	-	-	[78]

Ravi, et al.: A Com	prehensive Review on	Acalypha indica
---------------------	----------------------	-----------------

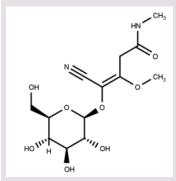
Table 3	3: Continued						
220	MOME inositol	-	GC-MS	L/TP	-	-	[94, 126]
221	Propanenitrile, 3-(5-diethylamino-l- methyl-3-pentynyloxy)-	-	GC-MS	L	-	-	[43]
222	Trifluoromethyl t-butyl disulfide	-	GC-MS	L	-	-	[119]
223	2,6,10 trimethyl undecatriene	-	GC-MS	ТР	-	-	[85]
224	α-tulenol	-	GC-MS	TP	-	-	[85]
225	α-toaldehyde	-	GC-MS	TP	-	-	[85]
226	Benzopyran	-	GC-MS	ТР	-	-	[85]
227	2-Methyl-3(3-Methyl-But-2 Enyl)- 2(4-Methul-Pent-3-Enyl)-Oxetane	-	GC-MS	L	CID: 550119	-	[181]
228	Trimethyl[4-(1,1,3,3, Tetramethylbutyl) phenox] Silane	-	GC-MS	L	CID: 528938	-	[181]
229	1,3-Dioxolane, 4- Ethyl-5-Octyl- 2,2Bis (Trifluoromethyl)-, Trans-	-	GC-MS	L		-	[181]
230	Bicyclo (3.1.1) heptane	-	GC-MS	TP	CID: 638057	-	[102]
231	hexamathyl and 1,3-dioxolane,4- ethyl-5-octyl-2,2- bis(Trifluoromethyl), Trans	-	GC-MS	L	-	-	[181]
232	Phenyl Methylhydrazino N-sulfamoylisosemicarbazide	-	GC-MS	L	-	-	[126]
233	3-Phenyl-1,2-pyrazole	-	GC-MS	L	-	-	[126]
234	(E)-1-(tert-butyldimethylsilyl)-4,4- dimethyl-2-penten-1-one	-	GC-MS	L	-	-	[126]
235	Thiophene, 2-propyl-	-	GC-MS	L	CID:73771	-	[126]
236	3-Oxo-20-methyl-11-à- hydroxyconanine-1,4-diene	-	GC-MS	L	-	-	[126]
237	Hexadecanoic acid, 2,3-dihydroxypropyl ester	-	GC-MS	L	-	-	[126]
238	4-cyanomethylquinoline	-	GC-MS	L	CID: 257387	-	[126]
239	Ramipril glucuronide	-	HR-LC-MS	L	CID: 71751964	-	[78]
240	Methyl N-(amethylbutyryl)glycine	-	HR-LC-MS	L	-	-	[78]
241	6-Hydroxydesmethylondansetron	-	HR-LC-MS	L	-	-	[78]
242	5-MethyltetrahydropteroyltriL- glutamate	-	HR-LC-MS	L	-	-	[78]
243	3-Deoxo- 3betaacetoxydeoxydihydroge Dunin	-	HR-LC-MS	L	-	-	[78]
244	24-Nor-5beta-chol-22- ene3alpha,7alpha,12alpha-triol	-	HR-LC-MS	L	-	-	[78]
245	1-Octadecanoyl- 2(5Z,8Z,11Z,14Zeicosatetraenoyl)- sn-glycero3-phosphate	-	HR-LC-MS	L	-	-	[78]
246	1-[[2-(2,3-dihydro-2-oxo-1Hindol- 4yl)ethyl]propylcarbamate] glucuronide	-	HR-LC-MS	L	-	-	[78]
247	1,2 Di- (9Z,12Z,15Zoctadecatrienoyl)-3-O- Beta-Dgalactosyl-sn-glycerol	-	HR-LC-MS	L	-	-	[78]
248	1-(9Z-hexadecenoyl)-2(5Z,8Z, 11Z,14Z,17Zeicosapentaenoyl)- snglycerol	-	HR-LC-MS	L	-	-	[78]
249	(3S,7R)-epi-jasmonic acid (Fatty acyl)	-	HR-LC-MS	L	CID: 7251177	-	[78]
250	(22R)-1alpha,22,25trihydroxy-26,27 -dimethyl23,24-tetradehydro- 24ahomo-20-epivitamin D3 / (22R)-1a	-	HR-LC-MS	L	-	-	[78]

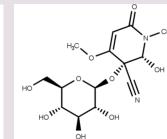
MLCC: Multilayer coutercurrent chroamatography; MLCCC: Multilayer coil coutercurrent chroamatography L-F:Combination of leaves and flowers; L:leaves; TP: Totalplant; HPLC:High performance liquid chromatography;GC-MS:Gas chromatograpy-mass spectroscopy; HR-LC-MS: High resolution-liquid chromatography;mass spectroscopy; L-Inf: combination of leaves-inflorescences; L-Flw: Leaves-flowers; TLC: Thin layer chromatography; NM: Not mentioned.

#### Table 4: Alkaloids of A.indica.

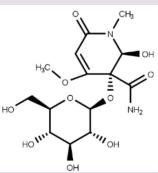


1.Acalyphin

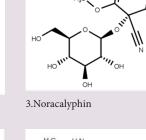


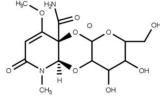


2.Epiacalyphin

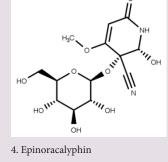


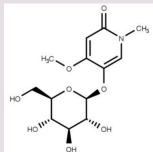
6.Acalyphin amide





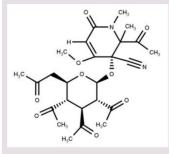
7.Epiacalyphin amide cycloside





8.Ar-Acalyphidone

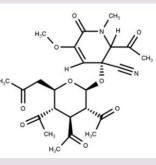
5.Seco-Acalyphin



9. 2-acetyl-4-methoxy-1,2-dimethyl-6oxo-

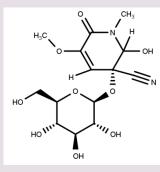
3-{[3,4,5-triacetyl-6-(2-oxopropyl) oxan-

2-yl]oxy}-1,2,3,6tetrahydropyridine-3-carbonitrile



10. 2-acetyl-5-methoxy-1-methyl-6oxo-

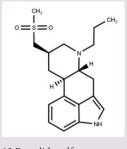
3-{[3,4,5-triacetyl-6-(2-oxopropyl)oxan-2-yl]oxy}-1,2,3,6-tetrahydropyridine-3carbonitrile

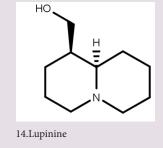


11. 2-hydroxy-5-methoxy-1-methyl-6oxo-3-

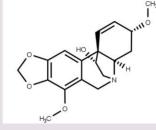
{[3,4,5-trihydroxy-6-(hydroxymethyl) oxan-

2-yl]oxy}-1,2,3,6tetrahydropyridine-3-carbonitrile





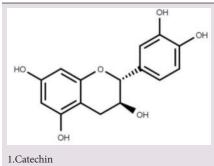
13.Pergolide sulfone

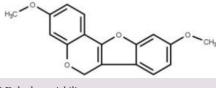


15.Ambelline



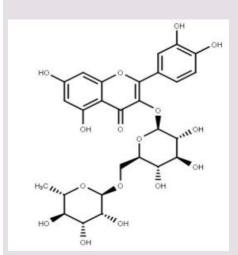
## Table 5: Flavonoids of A.indica.

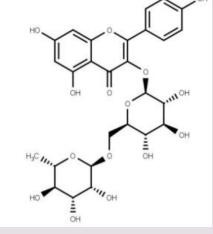


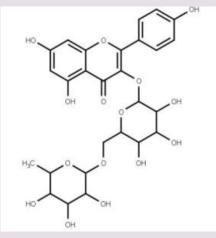


2.Dehydrovariabilin

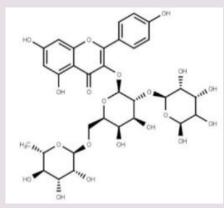
3.Quercetin 3-0-β-D-glucoside



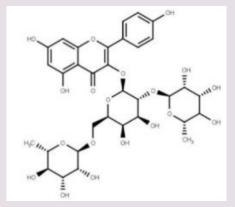




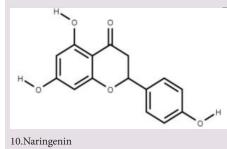
4.Rutin



5.Nicotiflorin

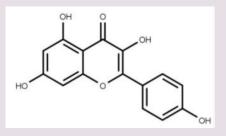


7.Clitorin



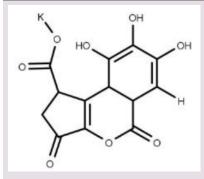
8.Mauritianin

6.Biorobin

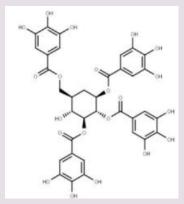


9.Kaempferol

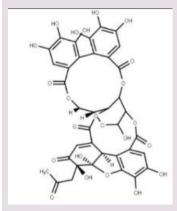
## Table 6: Tannins of A.indica.



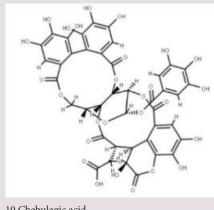
1.Potassium brevifolincarboxylate



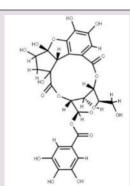
4. 1,2,3,6-tetra-O-galloyl $\cdot\beta$ -D-glucose



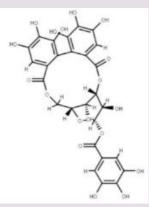
7. Acetonylgeraniin A



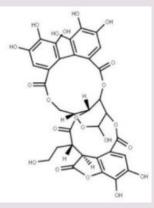
10.Chebulagic acid



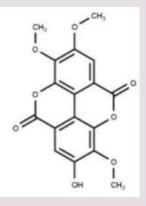
2.Acaindinin



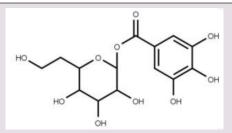
5.Corilagin



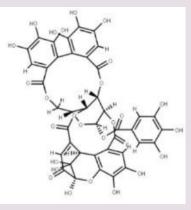
8.Euphormism M2



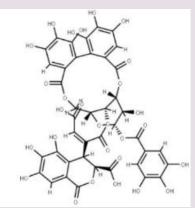
11.Tri-O-methylellagic acid



3. l-O-galloyl- $\beta$ -D-glucose

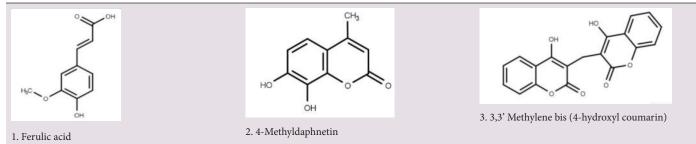


6.Geraniin

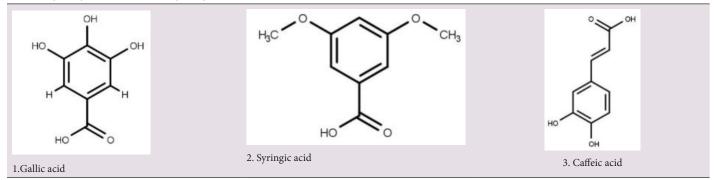


9.Repandusinic acid A

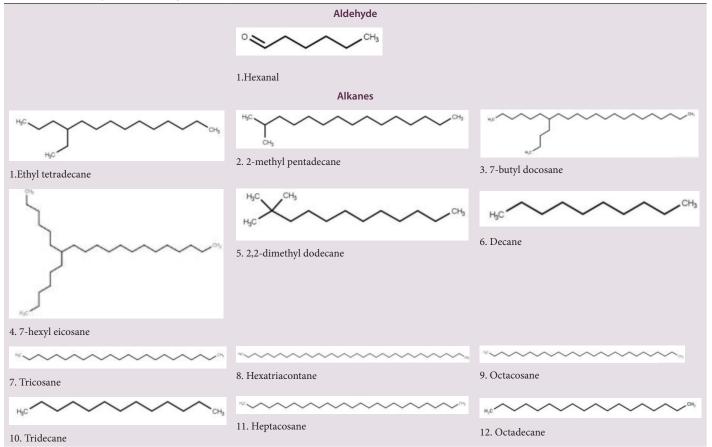
#### Table 7: Courmarins of A.indica.



#### Table 8: Hydroxy benzoic acid and hydroxy cinnamic acids of A.indica

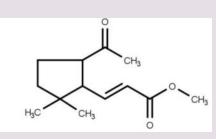


#### Table 9: Volatile compounds and fatty acids of A.indica.



Esters

#### Table 9: Continued...



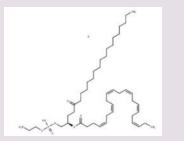
1. 2-propenoic acid, 3-[5-acetyl-2,2dimethylcyclopentyl], methyl ester,[1a(E),5a]-



4. Heptadecanoic acid, 16-methyl- methyl ester



7. 14-methylhexadecanoic acid, picolinyl ester



10. GPEtn(18:0/22:6(4Z,7Z,10Z,13Z,16Z,19Z))

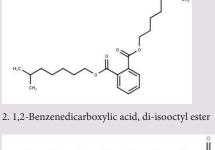


13. Trifluoro acetic acid, n-octadecyl ester

H

16. Pentadecanoic acid, 14-methyl-, methyl ester



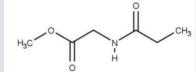


5. Hexadecanoic acid, 14-methyl, methyl ester

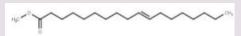
8. Ethyl ester of hexadecanoic acid



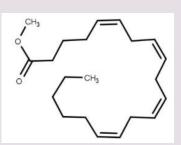
11. 9-octadecenoic acid [Z], 2-hydroxy-1-(hydroxymethyl) ethyl ester



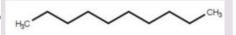
14. Propionylglycine methyl ester



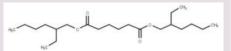
17. 10-octadecenoic acid, methyl ester



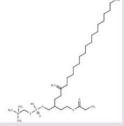
3. Eicosatrienoic acid methyl ester



6. Nonadecanoic acid, 18-oxo, methyl ester



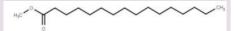
9. Hexanedioic acid, bis(2-ethylhexyl) ester



12. GPCho(16:0/3:1(2E))

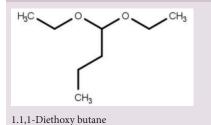


15. Methyl arachidate



18. Hexadecanoic acid methyl ester

19. Trifluoro acetic acid, n-heptadecyl ester



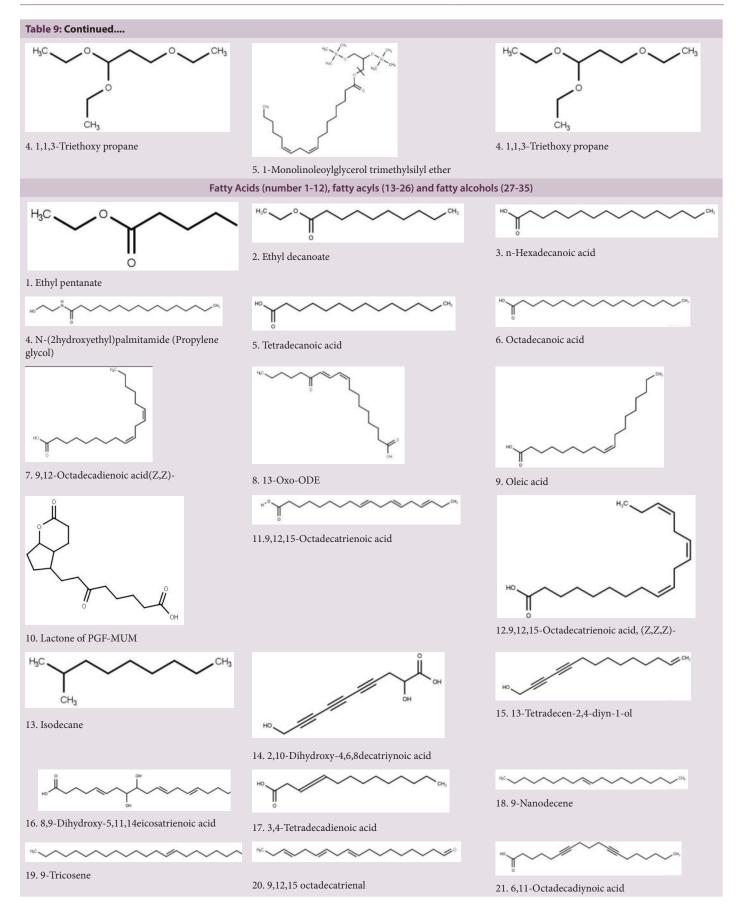
H<sub>3</sub>C O CH<sub>3</sub>

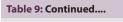
2. 1,1-Diethoxy pentane

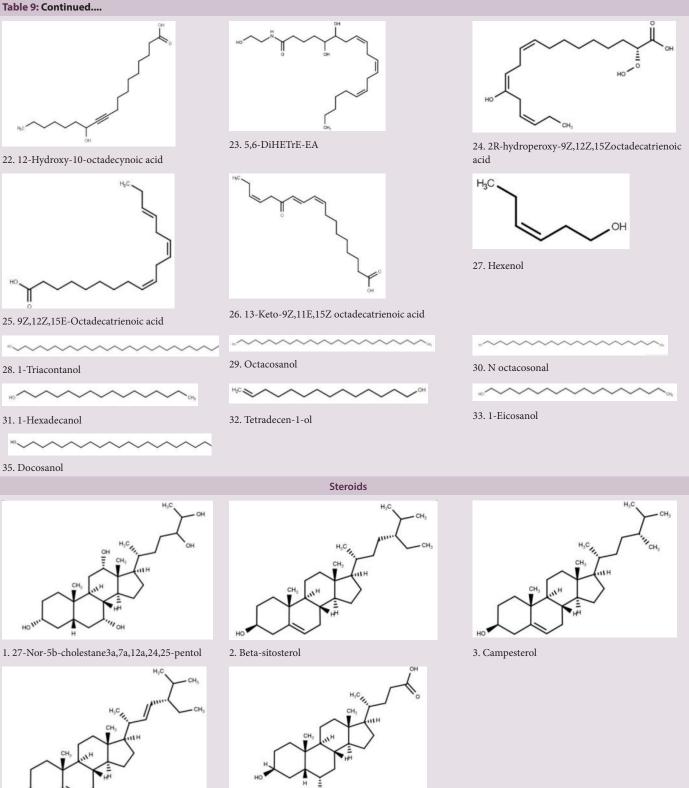
Ethers

H<sub>3</sub>C O CH<sub>3</sub> CH<sub>3</sub>

3. 1,1-Diethoxy hexane





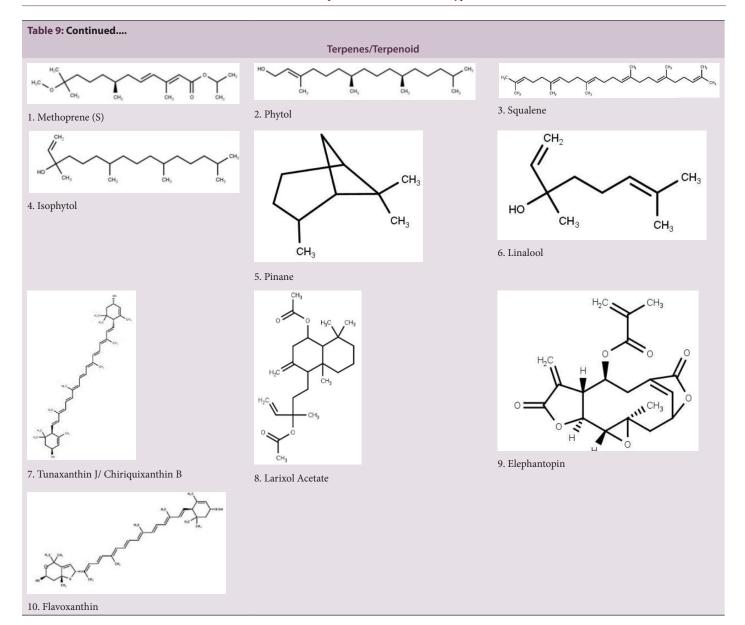


5. 3beta,6alpha,7alphaTrihydroxy-5beta-cholan-24oic

Acid

Continued....

4. Stigmasterol



breast cancer cell lines. Whereas, the positive controls ellipticine and doxorubicin have shown IC<sub>50</sub> of 0.88 and 0.05 µg/mL respectively. In another study by Reddy et al. (2012)[227] using MTT assay explored that ethyl acetate extract (ranging from 0.1- 1000 µg/mL) treatment for 24 hrs on skin cancer cell liens (A431) shown  $IC_{_{50}}$  value 220  $\mu g/mL$  whereas, the treatment extended upto 48 hrs exhibits the  $IC_{50}$  78 µg/mL . Further, the extract also inhibited the 12 R LOX enzyme by dose dependent manner (50-300 µg/mL). Similarly an experiment with MTT, ethanolic extract alone and its formulation as casein-chitosan micro particles at 20-70 µg/mL have been shown the effect on prostate cancer cell lines (PC3) by releasing LDH enzyme. Interestingly, both were shown similar cytotoxicity up to 24 hrs but micropaticles significantly more effective when incubated up to 72 hrs.<sup>[228]</sup> In addition to this, synthesized silver and gold nanoparticles with whole plant aqueous extract (100 µg/mL) have been showed anti-breast cancer (using MDA-MB-231 cell line) effect by showing 40% cytotoxicity, apoptotic changes with elevation of caspase-3 enzyme (more effect was shown by gold nano particles) and DNA damage.<sup>[229]</sup> Sivaraj et al. (2014)<sup>[230]</sup> reported that copper oxide nanoparticles from whole plant aqueous extract exhibited potential anti-breast cancer activity (97% cytotoxicity on MCF-7 cancer cell lines at 100  $\mu g/$  mL).

#### Wound healing property

Topical application of total plant ethanolic extract (10%) on wounds of male rats noticed that healing of excision wound per day was 18%, whereas breaking strength of incision wound was 279.75 g (breaking strength of control is 63.523 g).<sup>[231]</sup> In another study, 1 % hot water extract applied on excision wound for 19 days in either sex rats exhibited potential healing property (44%) than standard povidone-iodine ointment (28%). In incision wound, the extract treatment for 10 days showed the tensile strength 82.387 gnearer to the standard povidone-iodine of 86.113g.<sup>[232]</sup> Ganeshkumar et al. (2012)<sup>[233]</sup> have also beenbeen carried out experiments on similar wound models of male rats. Topical application of 50% ethanolic extract (40 mg/kg) of leaves healed the wounds by increasing percentage of collagen, DNA, protein, hexosamine (basic substance for collagen synthesis), uronic acid, TNF-α, TGF-β1, collagen 1a (I), collagen 3a (I) and decreasing lipid peroxidation. Further histopathological observation of treated tissues illustrated the fair deposition of fibroblast cells, cellular infiltration and collagen deposition,

is an indicative sign of wound healing. Experiment on angiogenesis, which is an important event being occurred in wound healing has been conducted with 1 % hot aqueous extract (0.1 mg/ $\mu$ L) of leaves in chick chorioallantoic membrane model. Extract treatment formed 10 blood capillaries whereas the standard vascular endothelial growth factor was resulted 13 capillaries in the duration of 72 hrs.<sup>[232]</sup> Hence, further purification of bioactive compounds from these extracts would help the scientists for the preparation of natural drugs in future.

### Antifertility activity

Petroleum ether and ethanol extracts of 600 mg/kg given to 7 days pregnant female rats for ten days treatment significantly inhibited the pregnancy progression by decreasing 75% and 62.5% of implantations respectively. Extracts also caused unfavourable conditions for fertility through increasing diameter of uterus, thickness of endometrium, height of the endometrial and epithelium weight. Moreover, extracts increased the uterine weight by effecting 37% and 32% of ethinyl estradiol levels respectively. Confirmation to this, female rats delivered healthy pups when discontinued the treatment with extracts.<sup>[234]</sup> Further, bioactive compounds isolation is required in the background of the established fact on petroleum ether and ethanol extracts to ascertain potential antifertility porperty.

#### Acute intravascular haemolysis prevention.

Clinically diagnosed patients characteristically showed low haemoglobin content, changes in peripheral blood, reticulocytosis, increased serum indirect bilirubin, haemoglobinuria and reduced levels of glucose-6-phosphate dehydrogenase enzyme confirmed that it is an acute intravascular haemolysis, when the same patients received *A.indica* leaves broth, they were recovered within 4 days by reestablishing of all hematological alterations. <sup>[5]</sup> This information gives a clarity that *A.indica* broth has capacity to reduce the acute intravascular haemolysis.

#### Antivenom property

An important aspect of A.indica is, its neutralizing ability on snake venom/poison as practiced by traditional healers of India and Srilanka.<sup>[23,27]</sup> The nullifying property of aqueous ethanol extract (95%) was studied on Viper russelli venom lethality (lethal dose of 61 µg/kg) in mice model; venom haemorrhagic (venom 340 µg/kg), mast cell degranulation (venom 61 µg/kg) and necrotic (venom 171 µg/kg) in rat models. Plant extract (750 mg/kg) treatment to venom affected rat models have recovered by significant inhibition of haemorrhage (from 10.1275 ± 8.53 to 3.1875  $\pm$  6.57), necrosis and mast cell degranulation; decreased plasma lipid peroxidation ( from 24.27  $\pm$  2.46 to 10.5200  $\pm$  0.17); and increased kidney GSH level and catalase enzyme activity. Interestingly, ethanolic extract (750 mg/kg) shown 100% survival rate of rats affected than venom antiserum (87.5%) (Shirwaikar et al. 2004). In another study with petroleum ether, benzene, chloroform and acetone extracts of leaves on Viper russelli venom, the acetone extract only at 500 mg/kg increases the survival rate upto 87 % which is equal to the positive control venom antiserum (87.5 %).<sup>[235]</sup> Beside these, in vitro experiments such as venom induced cardiac arrest on frog heart and neurotoxicity (1-4 µL of venom) in abdominus muscle of frog have significantly protected by ethanol extract with a dose of 1.6 mg/mL. Acetone extract (0.4 mg/mL) also protected the venom (0.1 mg/mL) induced haemolysis about 77.9%.[223,235] These results reveal that A.indica has potential antivenom phytochemical constituents.

#### Antitoxic effect

Along with snake venom neutralization, *A.indica* also nullified the Puffer fish muscle poison in mice. Hot water extract (5 g/L) of stem and leaves

given to mice nullified the 1% acetic acid extracted poison of puffer fish (*L. lunaris*) muscle. Plant extract reduced the toxicity by increasing body weights, ALT, AST, and ALP activities and HDL-C while decreasing CRE, UA, TC, TG, LDL-C, T Bil, D Bil, ALB, GLB, TP and GLU in serum; elevating antioxidant enzymes (SOD, CAT and GPx) in kidney, heart and liver; decreasing liver lipid peroxidation. Histopathology evidence of liver, kidney and heart tissues showed that extract protected these from fish muscle toxicants.<sup>[236]</sup>

#### Analgesic activity

Methanolic extract of *A. indica* total plant tested in mice (20-25g, 30 days age) to define its analgesic (pain perception) property. Writhing reflexes caused by 0.7% acetic acid was inhibited upto 51.1% and 57.2% by 200 and 400 mg/kg of extract respectively whereas the standard aminopyrine showed 89.9% inhibition at 50 mg/kg.<sup>[98]</sup> In another hot plate method, polyphenolic fraction (400 mg/kg) was increased the latency period upto 3 hrs in mice (20-25 g) whereas the standard diclofenac sodium (0.13 mg/kg) maintained upto 6 hrs. Tail-flick method for the same fraction revealed that it decreases the pain threshold of tail upto 6 hrs in rats (180-220 g) while diclofenac sodium showed significant results than fraction. The authors concluded that this property of fraction is due to the presence of polyphenol and flavonoid compounds.<sup>[82]</sup> The results obtained from these experiments shows that this plant is a good source for analgesic compounds for further isolation.

## Anti-inflammation property

The carrageenan (1%) induced paw edema in rats (140-160 g) was inhibited by 125 and 250 mg/kg of methanolic extract of A.indica to an extent of 21.51% and 30.64 % respectively but this is lower than (37.55%) that of phenylbutazone of 100 mg/kg.<sup>[98]</sup> Godipurge et al. (2015)<sup>[82]</sup> reported that 400 mg/kg of polyphenolic rich extract exhibited potential inhibition of paw volume (92.3%) than standard diclofenac sodium (61.5 %) of 0.9 mg/kg by the action on prostaglandin E2 production. The same authors conducted in vitro experiment with polyphenolic rich extract (250 µg/mL) which stabilized the membrane about 20 % in hypo saline induced inflammation on human red blood cells, is nearer to the diclofenac sodium (21%). In another in vitro experiment, 0.1 mg/mL dose of ethyl acetate and hexane extracts have potentially showed anti-inflammation activity by inhibiting COX-2 and 5-LOX enzymes whereas, hexane extract shown more effect on COX-1.[227] Fatima et al. (2017)<sup>[237]</sup> reported that ethyl acetate and water extracts at 0.3 mg/mL showed inhibitory activity on 12R-LOX in in vitro, are equal to the standard drug Zileuton (0.07 mg/mL) activity. Having obtained all results about inflammation, it is clear indication that A.indica is a potential source for anti-inflammation drug development.

#### Kidney stones digestion

In male rats, ethylene glycol (0.75%) induced kidney stones were digested with ethanolic extract of *A. indica* (200 mg/kg). It was observed a decrease in AST, ALT, ALP and ACP enzyme activities in urine and serum whereas the same were increased in kidney. At the same time, extract also brings back membrane bound enzymes (Na+ K+, Ca+ and Mg+ ATPases) in kidney, equal to standard drug thiazide (150  $\mu$ g/kg). The authors concluded that this kidney stones digestion of *A.indica* extract was due to the presence of antioxidant compounds.<sup>[238]</sup> The chemical constituents responsible and mechanism of action involved in the digestion of kidney stones is to be unraveled for further clinical trials in this area of research.

## Cardio protective property

The cardioprotective property of flavonoid rich methanolic extract of A. indica leaves (200 mg/kg) studied in isoproterenol (ISO) (85 mg/kg) induced myocardial ischemia rats. The extract protected the myocardium by decreasing total cholesterol, triglycerol, low-density lipoprotein cholesterol, very low density lipoprotein cholesterol, fatty acids and phospholipids, and by increasing high density lipoprotein cholesterol in plasma, hepatic and cardiac tissues.<sup>[239,240]</sup> In another study, pre and post treatment of 70% methanolic extract (500 mg/kg) of leaves given to ISO induced female rats protected the cardiac tissue by inhibiting over expression of serum marker enzymes (LDH, AST, ALT and CK-MB); decrease in serum CRP and Troponin T levels and MDA content; elevated antioxidant enzymes (SOD and CAT) and prevented caridiac tissue damage. In addition to this, it prevented the cardiac death caused by furosemide (10 mg/kg) and potassium chloride (10 mEq/kg) by reverting CRP, CK-MB and troponin levels; decreasing serum, urine electrolytes (K+ and Na+) and urine output ; protecting architecture of left ventrical tissue; gaining of body weight; and maintaining of glucose levels.<sup>[84]</sup>

#### Antidiabetic activity

Aqueous ethanolic (80%), hydro alcoholic (50 %), chloroform soluble fraction of hydro alcoholic, n-butanol soluble fraction of hydro alcoholic and butanol insoluble fraction of hydro alcoholic extracts with a dose of 400 mg/kg and methanol: acetone fraction (70:30) with a dose of 500 mg/kg of A.indica have been nullified the alloxan induced diabetes complications in rats with an evidence of recovered body weight and dropped blood glucose levels.Of these extracts, the aqueous ethanolic only maintained the glucose levels (296.5 to 153.83) which was nearer to the standard glibenclamide (10 mg/kg) treated glucose levels (296.5 to 142.7) of rats.<sup>[80,221,241]</sup> In streptozotocin induced diabetic neonate rats (5 days age), methanolic extract (100 mg/kg) showed antidiabetic potentiality by decreasing blood glucose levels upto 57%, whereas 67% was observed in standard glibenclamide (5 mg/kg) treatment.<sup>[242]</sup> Rani, (2014)<sup>[126]</sup> reported potentiality of 100% ethanolic extract (500 mg/kg) on STZ induced diabetic matured rats. The extract reverted the complications by reducing glucose; decreasing DNA, RNA, glucose-6-phosphatase, ALP, ACP, AST, ALT, LDH, lipid profiles (total cholesterol, free chlolesterol, triglycerides, phospholipids, free fatty acids, HDLcholesterol, LDL- cholesterol, VLDL- cholesterol); and elevating glycolytic enzymes (Hexokinase and Phosphoglucoisomerase), TCA cycle enzymes (succinate dehydrogenase, malate dehydrogenase), GSH, vitamin A, vitamin C and vitamin E. The extract also recovered the liver and kidney architecture to near normal state under diabetic condition. In another STZ induced experiment, the polyphenolic fraction (100 mg/kg) of leaves, methanolic extract of stem (600 mg/kg) and ethanolic extract of total plant (500 mg/kg) alleviate the oxidative stress in liver by elevating cellular antioxidant enzymes (SOD,CAT, GPx, GST, GR and G6PDH) equal to glibenclamide (20 mg/kg) and metformin (10 mg/kg) treatment in rats.[126,243,78]

The methanolic extract (600 mg/kg) of stem was tested for postprandial antihyperglycemic in maltose (2 g/kg), glucose (2 g/kg) and sucrose loaded rats which are also induced by STZ. Extract lowered the glucose levels upto 70.41% in maltose alone rats whereas extract and standard acarbose (5 mg/kg) lowered the glucose levels up to 69.10% and 44.87% respectively in maltose plus STZ rats. It was observerd that, 52.7% of glucose levels were dropped in sucrose loaded rats whereas 80.35% and 40.93% dropped by extract and acarbose respectively in sucrose with STZ rats. Further it was noticed that, the extract not suppressed the postprandial hyperglycemic condition in diabetic rats which are loaded with glucose. The extract (300 mg/kg) also protected the liver damage by decreasing liver markers SGOT, SGPT, ALP, and TB in serum.<sup>[243]</sup>

Nandhakumar *et al.* (2009)<sup>[8]</sup> have reported *in vitro* anti-amylase (a marker enzyme for diabetes) activity of ethanol, chloroform and hexane extracts. The chloroform and hexane extracts (0.1 mg/mL) inhibited the enzyme upto 75.32% and 84.51% respectively, whereas ethanol extract (0.001-0.1 mg/mL) didn't show any activity. From these reports, it is very clear that the extracts of *A.indica* have potential anti-diabetic property on alloxan and streptozotocin induced animals.

#### Anthelmintic activity

*Pheretima posthuma* is an Indian earthworm belongs to Annelida and having anatomically, physiologically resemblance with intestinal round worm parasite of humans. Treatment with 70% hydro alcoholic extract (10, 25 and 50 mg/mL) of roots induced the paralysis and death of annelidan in a dose dependent manner. The extract (50 mg/mL) caused the paralysis at 20 min of time and death within 30 min whereas the standard albendazole (10mg/ml) achieved the same condition at 20 min and 46 min respectively.<sup>[244]</sup>

#### Antiarthritic activity

Ethanolic and water extracts (250 mg/kg) of *A.indica* exhibited antiarthritic activity on heat killed *Mycobacterium tuberculosis* (0.5 mL of 5%) induced chronic arthritic rats (130- 150 g). Ethanolic extract inhibited swelling of paw upto 7.85%, water extract showed 41.96% whereas standard drug ibuprofen exhibited 49.10% by decreasing the levels of alkaline phosphatase.<sup>[245]</sup>

#### Hepatoprotective activity

A.indica extracts and synergy with other plants/bioactive compounds shown protective effect on thioacetamide, paracetamol, CCL4, rifampicin-isoniazid and hypoxia induced hepato toxicity in experimental animals. Methanolic extract (300 mg/kg) and methanolic fraction (250 mg/kg) protected the thioacetamide (100 mg/kg) induced hepatic tissue in wistar strain albino rats by decreasing SGOT, SGPT, ALKP, total cholesterol and total bilirubin; increasing albumin and total protein in serum; recovered the architecture of hepatic tissue.<sup>[81]</sup> In paracetamol (1 g/kg) induced male rats, 70% hydroalcoholic and ethanolic extracts (100 and 200 mg/kg) of leaves recovered the architecture of hepatic tissue by decreasing AST, ALT, ALP and lipid peroxidation; and elevating the levels of cellular antioxidants (SOD and GSH). [246,247] Vijayabhaskar et al. (2013)<sup>[248]</sup> have been stated hepatoprotective property of 70% alcoholic extract (300 mg/kg, post treatment for 10 days) of whole plant in CCL4 induced rats with an evident in drastic reduction of SGPT, SGOT and ALP in serum than silymerin (100 mg/kg), a standard drug.

Synergytic effect of methanolic extracts (70%) of *A.indica* (200 mg/kg) and *Centella asiatica* (150 mg/kg) have been protected the hepatic tissue of rats growing under hypoxia condition (10 %  $O_2$  and 90 %  $N_2$ ) by decreasing lipid peroxidation whereas standard vitamin C (100 mg/kg) didn't show any protection.<sup>[249]</sup> In another experiment, 70% ethanolic extract (150 mg kg) with piperine compound (20 mg) protected the rifampicin-isoniazid (50 mg/kg) induced hepatic tissue of rats equal to silymerin (100 mg/kg) with a mark of decreased SGPT, SGOT and ALP in serum.<sup>[248]</sup>

#### Antiulcer activity

Anti-ulcer property of 80% ethanolic extract of *A. indica* leaves and roots (100 and 200 mg/kg each) studied on pylorus ligated, acetyl salicylic acid (200 mg/kg), cold stress (4°C for 2 hrs) and 40% ethanol induced peptic ulcers. Root extract (200 mg/kg) shown 66.62 % defence against pylorus ligated ulcers by improving pH, reducing acidity/gastric volume whereas standard ranitidine exhibited 67.24%. Acetyl salicylic acid induced ulcer was reduced (55.61 %) by higher concentration of root extract whereas standard ranitidine showed 70.25%. Cold stress induced ulcers was

relaxed with the root extract (200mg/kg) by 58.14 % whereas 74.06 and 70.25 % were exhibited by diazepam (1mg/ kg) and ranitidine (50 mg/kg) respectively. Ethanol induced gastric lesions reduced 64.55 % by higher concentration of root extract while standard ranitidine reduces 68.19%. In all these studied experiments on ulcers, leaf extract didn't show any protentiality but root has comparatively equal with the standard drugs.<sup>[250]</sup>

## Metal accumulation property

The phytoextraction or hyper accumulation and translocation of metal ions (zinc, iron, copper, lead and cadmium) examined with A.indica. Olowu et al. (2015)<sup>[251]</sup> have identified heavy metals accumulation in leaves, stem and root tissues of A. indica growing at dump areas of Ibadan Metropolis, South West Nigeria. More accumulated metals found were: iron, zinc, and copper whereas moderately accumulated were: lead and cadmium followed by less accumulated nickel and chromium. It was also observed that the metals were more accumulated in leaves than other parts. In another experiment, accumulated metal ions resulted alterations of this plant have been reported. Different concentrations (0, 100, 200, 300, 400, 500 mg L<sup>-1</sup>) of Pb ion supplementation to A.indica for 12 days revealed that greater accumulation of Pb observed in root (121.6 mg  $g^{-1}$  DW) than shoot (17.5 mg  $g^{-1}$  DW). Accumulation decreases the length of root and shoot upto 49.9% and 50.9% respectively; the growth tolerance index of roots and shoots decreased upto 44.5% and 52.3% respectively; chlorophylls (chl a, chl b, chl a/b) and carotenoids content decreased in leaves. MDA levels (lipid peroxidation) increased in leaves by 117% and 151.4% at 100 mg  $L^{\text{-1}}$  and 500 mg  $L^{\text{-1}}$  concentrations respectively. Antioxidant enzymes (SOD, POX, CAT and APX) alteration also caused in leaves and roots. Protein banding analysis of leaves with higher concentration of Pb revealed that it causes disappearance of protein bands and decrease in protein intensity over the control plants. The RAPD-PCR profile of DNA of leaves exhibited that Pb induced the DNA damage, absence of bands and amplification of new bands.<sup>[252]</sup> It was observed that, A.indica plant has been struggled from metal toxicity however, its accumulation in this plant parts is helpful for the measurement of heavy metal toxicity, aswell as phyto remediation in water and soil polluted environments.

#### A.indica roots as cat attractant

For the protection of native endangered species of Christmas Island from cats, yellow crazy ants and black rats, a research group conducted an experiment to attract the cats using *A.indica* roots. Many cats attracted towards roots and chewed them (Figure 2). GC-MS analysis of dichloromethane and ethanol extracts of roots revealed two Iridoids such as (4R,4aR,7S,7aR)-isodihydronepetalactone and (4R,4aS,7S,7aR)-isoiridomyrmecin. Interestingly, these two compounds are known to have effects on behavioural activities of cats.<sup>[253]</sup>

#### Nanoparticles synthesis

Water and ethanol extracts of *A. indica* have been used by many research groups to synthesize metal based particles including silver, gold, palladium, copper oxide, yttrium oxide, zinc oxide and zirconium dioxide nanoparticles; and chitosan-casein micro particles. <sup>[228-230, 254-261]</sup> The biological properties of these particles are discussed in antimicrobial and cytotoxicity sections of this review.

### Antibacterial activity

Table 10 presents the antibacterial properties of solvent extracts of *A. indica.* Mainly three methods have been carried out to define the antibacterial property which includes dilution, well diffusion and disc diffusion. Various extracts have been exerted notable antibacterial activity on both gram positive (*Staphylococcus auereus, Bacillus subtilis, Bacillus*)

cereus, Bacillus megaterium, Staphylococcus epidermidis and Streptococcus faecalis) and gram negative (Escherichia coli, Proteus vulgaris, Salmonella typhi, Pseudomonas aeruginosa, Priteus mirabilis, Klebsiella pneumonia, Enterobacter aerogenes, Enterobacter cloacae and Vibrio cholerae.) strains of test organisms, human pathogens and multidrug resistance bacteria<sup>. [74,77,169,225,262,-266]</sup> The dilution method was done only for the leaves and total plant, the resulted MIC values were ranging from 0.004 mg (acetone insoluble fraction on Staphylococcus auereus) to > 5 mg and 0.02 mg (ethanol extract on E.coli) to 1 mg respectively.<sup>[77, 267]</sup> Here, we review few potential extracts with MIC values: Gopalakrishnan et al. (2000)<sup>[77]</sup> have reported that acetone insoluble and soluble fractions showed potential activity against Staphylococcus auereus (MIC: 0.004 mg) and Salmonella typhi (MIC: 0.05 mg) respectively. In another study, Govindarajan et al. (2008)<sup>[262]</sup> have been confirmed the MIC for chloroform, ethyl acetate, hexane and methanol extracts of leaves. The MIC of chloroform and hexane extracts on Streptococcus faecalis was 0.312 mg and 0.156 mg respectively whereas, the MIC of ethyl acetate on Staphylococcus auereus and Streptococcus faecalis was 0.312 mg. In all cases of dilution method, the activity was not compared with any standard drugs.

The disc diffusion method was performed by Poornima and Prabakaran, (2012),<sup>[74]</sup> Govindarajan *et al.* (2008),<sup>[262]</sup> Solomon *et al.* (2005),<sup>[169]</sup> and Shanmugapriya *et al.* (2011),<sup>[225]</sup> for antibacterial activity of leaves extracts. Among all, Govindarajan *et al.* (2008),<sup>[262]</sup> have been only reported the bacterial inhibition maximum at concentration of 5 mg by Ethyl acetate, chloroform, hexane and methanol extracts on *Staphylococcus aureus, Staphylococcus epidermidis, Bacillus cereus, Streptococcus faecalis* and *Pseudomonasaeruginosa*. Comparison with standard streptomycin disc (0.01 mg/mL), the 5 mg/mL of above extracts successfully showed similar activity. *A.indica* leaves water extract at 4% v/v inhibited the growth of *Mycobacterium tuberculosis* H37Rv, multi drug resistance strains such as DKU-156 and JAL-1236 about 68%, 95%, and 68% respectively. The same extract has not exhibited inhibition on fast growing *M. fortuitum* (TMC-1529) strain.<sup>[268]</sup>

Identification of antibacterial effective dose through well diffusion method is very difficult for extracts of leaves, stem, root and total plant on bacterial species. Almost all extracts studied at 100 mg/mL concentration except ethyl acetate extract of total plant (0.05 mg). In concern of zone of inhibition, the acetone, chloroform, ethanol, ethyl acetate, methanol and petroleum ether extracts shown maximum at 100 mg/mL against *Escherichia coli, Salmonella typhi, Pseudomonas aeruginosa, Klebsiella pneumonia* and *Vibrio cholera*. <sup>[263-266]</sup> Along with extracts, synthesized silver and copper oxide nanoparticles have been also shown antibacterial potentiality on *E. coli, V. cholerae, P.aeruginosa, B.subtilis, P. fluorescens, P.vulgaris* and *S.aureus* at the microgram level.<sup>[202,254,269]</sup>

## Antifungal activity

As shown in Table 11, the methanolic extract of *A.indica* leaves, stem and roots showed maximum activity against *Candida albicans* with the percent activity of 119%, 95% and 109% respectively whereas chloroform extract of leaves and roots have been shown 104% and 61% activity respectively and stem methanolic extract has 85% activity against *Aspergillus niger*. The authors have declared that the antifungal activity of extracts was due to having a bioactive compound clotrimazole.<sup>[169]</sup> In another experiment, minimum fugicidal concentrations (MFC) of hexane, methanol and ethanol extracts of *A.indica* whole plant against 12 fungals species (Table 11) revealed that hexane extract only shown maximum activity at 0.5 mg/mL against *Trichophyton rubrum* PSSF57/01, *Trichophyton tonsurans* PSSF46/01 and *Epidermophyton floccosum* PSSF73/01 whereas other extracts MFC were not better than hexane extract. In this study, the authors compared the results with standard drugs indirubicin, fluconazole

	References	[77]	[2		[262]	[262]	[262]	[262]	T Acaly		[74, 262]	[262]	[169]	[262]
		[7]	[77]	[77]	[20	[2(	[20	[3(	[74]	[74]	[]7,	[2(	[16	[20
	V	1	1	1	1	1	1	1	I	1	ری ا	1	I	5
	SF	1	1		MIC: 0.312 mg	MIC: 0.312 mg	MIC: 0.156 mg	MIC: 0.156 mg	ı	ı	5 mg-15 mm	5 mg- 15.0 mm	ı	5 mg-12
(uo	E	1	I.				1		I	ī	1		I	
	EA													
cone or		' Ba	0.25 -	0.4 -	' 50	< 5	1	< 5 '	י מל	י מל	A	- NA	I	- VN
ration-2	KP	MIC: 0.125 mg	MIC: 0.25 mg	MIC: 0.4 mg	MIC: > 5 mg	MIC: > . mg	MIC: >5 mg	MIC: > mg	100 mg- NA	100 mg- NA	100 mg- 12 mm; 5 mg-NA	5 mg- NA	I	5 mg: NA
oncent	PM		1			1	1	1	I	ī	1		I	
ective	PA	MIC: 0.15 mg	MIC: 0.15 mg	MIC: 0.4 mg	MIC: 2.5 mg	MIC: 5 mg	З в	MIC: 2.5 mg	100 mg- NA	100 mg- NA	100 mg- 15 mm; 5 mg-10 mm	5 mg- 10 mm		5 mg- 9.7 mm
iues/En		MIC: 0.15 n	MIC: 0.15 n	MIC			MIC: 5 mg						ı	5 mg mm
(IVIIC VAIUE Leaves	SE				MIC: : 1.25 mg	MIC: 1.25 mg	MIC: 1.250 mg	MIC: 0.625 mg	100 mg- NA	100 mg- NA	100 mg- 15 mm ; 5 mg- 13.8mm	5 mg- 13.8mm		5 mg- 11.0mm
ve dose	BM				1 ::	I		1		- 4			I	u, —
ещесці		ജ	- 0.0	0.4 -	1	1	1	1	I	I	1	1	I	1
ted and	ST	MIC: 0.05 mg	MIC:0.0 5 mg	MIC: 0.4 mg	I.	1	1	ı	ı	ı	1	I	ı	
lism tes	BC				MIC: 0.625 mg	MIC: 0.625 mg	MIC: 0.312 mg	MIC: 0.156 mg			5 mg-15 mm	5 mg- 15.0 mm		5 mg- 13.8mm
Micro-organism tested and effective dose (MIC values/Effective Concentration-Zone of inhibition) Leaves	BS	MIC: 0.15 mg	MIC: 0.15 mg	: 0.4			0 1		gu	gm (			I	u) —
MICI			MIC	MIC: 0.4 mg	i.	,		ı	100 mg- 14 mm	100 I NA	100 mg- 20 mm	i.	i.	
	SA	MIC:0.00 4 mg	MIC: 0.15 mg	MIC: 0.4 mg	MIC: 0.625 mg	MIC: 0.312 mg	MIC: 0.312 mg	MIC: 0.156 mg	100 mg- 27 mm	100 mg- NA	100 mg- NA; 5 mg-14.6 mm	5 mg- 14.6 mm	1	5 mg- 12.8 mm
	PV	1				MIC: > 5 mg	MIC: >5 mg	MIC: >5 mg	100 mg-	100 mg- NA	100 mg- 12mm; 5 mg- NA	5 mg- NA	1	5 mg: NA
	EC	MIC: 0.125 mg	MIC:0.25 mg	MIC: 0.4 mg	in an	ы Ш	ng B	MIC: >5 mg	100 mg- NA	100 mg- NA	100 mg- 10 mm; 5 mg- NA	5 mg- NA	0.001 mg	5 mg: NA
		MIC: 0.125	MIC	MIC	MIC: > 5mg	MIC: > 5 mg	MIC: >5 mg	MIC	100 I NA	100 NA	100 10 r. mg-	5 m	0.00	5 m
Method		Dilution	Dilution	Dilution		Dilution	Dilution	Dilution	Disc diffusion	Disc diffusion	Disc diffusion	Disc- diffusion	Disc- diffusion	Disc- diffusion
Extracts		Acetone insoluble fraction of methanol	Acetone soluble fraction of methanol	Chloroform		Ethyl acetate	Hexane	Methanol	Diethyl ether	Ethanol	Ethyl acetate	Chloroform	Clotrimazole	Hexane
S.No			5	6		4	J.	9	4	∞	6	10	11	12
		-				4				~				

				Ravi, et	al.: A Comprehensive F	Review on Ad	calypha indio	са			L
	[74, 225, 262]	[263]	[263]	[264, 265]	[264-266]	[263, 266]	[263, 266]	[263]		[225]	[169]
		100 mg- 29	100 mg- 27 mm	100 mg- mm	75 mg- 13 mm; 100 mg- mm 13 mm	100 mg- 15 mm	100 mg- mm	100 mg- 22 mg			
	5 mg- 16.5			100 mg-25 mm	100 mg-15 mm	1	1	1			
	1	1	1	1		ı	1	1		1	
										1	
	5 mg-NA; 100 mg- NA	100 mg- 28 mm	100 mg- 25 mm	100 mg- 20 mm	100 mg- 3.5 mm75 mg-20 mm	100 mg- 10 mm, 0.9 mm	100 mg- 1 mm, 33mm	100 mg- 20 mm			
		1			1						1
	5 mg- 12.7 mm; 100 mg- NA	100 mg- 35 mm	100 mg- 33mm	100 mg - 30 mm	100 mg- NA	100 mg- 20 mm	100 mg- 35 mm	100 mg- 25 mm			
	5 mg-14 mm; 100 mg- NA	I.	ı	ı			ı	ı		I	
		1	1	1			1		Roots	1	1
	40 mg-6 mm	100 mg- 30 mm	100 mg- 28 mm	100 mg- 20 mm, 25 mm	75 mg- 20 mm; 100 mg- 18 mm	100 mg- 18 mm	100 mg- 32 mm	100 mg- 22 mm	Ro	30 mg	
	15 mg- 14 mm; 40 mg-9 mm	1		100 mg- 15 mm	75 mg- 13 mm; 100 mg-11 mm			1		30 mg	
	40 mg- NA; 100 mg- NA	1		100 mg- 30 mm	100 mg- 1.9 mm, 13 mm ; 75 mg- 13mm	100mg- 10. mm	100 mg- 1.3 mm			NA	
	5 mg- 15.8 mm; 30 mg-8 mm; 100 mg- NA	100 mg- 23 mm	100 mg- 20 mm	100 mg- 12mm	100 mg- 2.7 mm, 15 mm	100 mg- 9mm, 0.8 mm	100 mg- 1.1 mm, 26mm	100 mg- 14 mm		30 mg	
	5 mg- NA; 100 mg- NA	1				1				1	
	5 mg-NA; 40 mg-6 mm; 100 mg- NA	100 mg- 35 mm	100 mg- 30 mm	100 mg- 30 mm	75 mg-25 mm; 100 mg-23 mm	100 mg- 20 mm	100 mg- 35mm	100 mg- 25 mm		30 mg	0.5 µg
:	Disc- diffusion	Well diffusion	Well diffusion	Well diffusion	Well diffusion	well diffusion	Well diffusion	Well diffusion		Agar diffusion method	Disc- diffusion
Table 10: Continued	Methanol	Acetone	Chloroform	Ethanol	Ethyl acetate	Hexane	Methanol	Petroleum ether		Methanol	Clotrimazole from Hexane, acetone, chloroform and methanol
Table	13	14	15	16	17	18	19	20		1	7

## Ravi, et al.: A Comprehensive Review on Acalvpha indica

				i di	, <i>ci ui</i>	.: A CC	mprer	lens	ive Review	v on Ac	calypha indi	ca				F
[266]	[266]	[266]		[169]	[266]	[266]	[266]		[267]	[267]	[76]	[76]	[76]	[76]	[280]	[75]
	ı	I.		1	1	I	ı		1	ı	1	1	ı	,	ı	ı
				1							1		1		,	
	ı	ı.		1		ī	ı		1 mg	ī	1.5 mg- 11 mm	1mg- 7 mm	0.5- 7mm mg	1 mg- 7mm	ı	
100 mg- 3.3 mm	100 mg- 0.8 mm	100 mg- 1.2 mm			ı	ı	1		1 mg	ı.	0.5 mg- 6mm	0.5 mg- 7mm	0.5 mg- 8mm	1 mg-9 mm	ı.	
100 mg- 3.3 mm	100 mg- 0.8 mm	100 mg- 1.2 mm			100 mg- 1.2 mm	100 mg- 1.1 mm	100 mg- 1.2 mm		MIC: 0.08 mg	MIC: 90 µg	0.5 mg-6 mm	0.5 mg-8 mm	0.5 mg-7 mm	1 mg- 9mm		
	ī	1		1		1	ī		MIC: 0.02 mg	MIC: 60 μg	1 mg- 9mm	0.5 mg-7 mm	0.5 mg-7 mm	1 mg- 7mm		
	1	I			I	I	1			ı	0.5mg- 6mm	0.5 mg- 7mm	0.5 mg-9 mm	1 mg-9 mm	300 μg-9 mm	50 μg-15 mm
1	1	1			1	1	1		1	I.	1 mg- 10mm	0.5 mg- 8mm	0.5 mg- 10mm	1 mg- 8mm	1	1
	1		۲	1							1 mg- 10 mm	0.5mg- 7 mm	0.5mg- 8 mm	1 mg- 9mm		
			Stem		· ·						0.5 mg -6 mm	0.5 mg- 9 mm	0.5 mg -10 mm	1 mg-8 mm		50 μg-16 mm
		ī			ī	ı			ı.		1	1	1			
100 mg- 1.7 mm	100 mg- 1.2 mm	100 mg- 1.5 mm			100 mg- 1.2 mm	100 mg- 1.1 mm	100 mg- 1.2 mm		1		1	1	1		300 μg-18 mm	
100 mg- 2.1 mm	100 mg- 1.1 mm	100 mg- 1.2 mm			100 mg-1 mm	100 mg- 0.8 mm	100 mg- 0.8 mm		MIC: 0.04 mg	MIC: 60 µg	,		1	ı	300 µg- 13 mm	50 μg- 8mm
	ī			1			,		1		1	1	ı		1	
1	1	1		0.01 mg	1	1	1		MIC: 0.04 mg	MIC:20 µg	1 mg- 10mm	0.5 mg-7 mm	0.5 mg-10 mm	0.5 mg-7 mm	300 μg-17 mm	50 µg- 18mm
Well diffusion	Well diffusion	Well diffusion		Disc- diffusion	Well diffusion	Well diffusion	Well diffusion		Dilution	Dilution	Cup plate method	Cup plate method	Cup plate method	Cup plate method	Disc diffusion	Well diffusion
Ethyl acetate	Hexane	Methanol		Clotrimazole ffrom Hexane, acetone, chloroform and methanol	Hexane	Ethyl acetate	Methanol	ant	Acetone	Ethanol	Hydro alcoholic	Hexane	Ethyl acetate	Methanol	Chloroform	Methanol
33	4	C)		-	2	9	4	Total plant	1	5	<i>භ</i>	4	5	9	4	∞



Figure 2: A.indica roots as cat attractant: chewing of roots by cats. [253]

and ketoconazole.<sup>[270]</sup> Experiments were conducted on six clinical isolates cum drug resistance Candida albicans strains with methanol, acetone, petroleum ether and water extracts of A. indica leaves. Among extracts, the methanol extract at 0.05 mg/mL showed good activity. The best activity concentrations are shown in Table 11.<sup>[271]</sup> Sakthi et al. (2011)<sup>[272]</sup> were also isolated 6 fungal species and their inhibition by ethanol and ethyl acetate extracts (100 mg/mL, 200 mg/mL and 300 mg/mL) of leaves revealed that the ethyl acetate extract at higher concentration (300 mg/mL) showed good antifungal property than ethanol. Somchit et al. (2010)<sup>[273]</sup> isolated four fungal strains which have sensitive to the chloroform extract at 30 mg/mL than ethanol and water extracts. The potential chloroform extract competed with fungicide ketoconazole but not with fluconazole and fraconazole. On other hand, A.indica leaves synthesized silver and copper oxide nanoparticles exhibited potential property on C. albicans, A. alternate, S. sclerotiorum, M. phaseolina, R. solani, B. cinerea, C. lunata and A. niger.<sup>[255, 269]</sup>

## Endophytic fungi in plant parts of A.indica

The endophytic fungal species present in A. indica plant parts such as leaves, petiole, stem and roots have been isolated and identified by Kurandawad and Lakshman, (2014)<sup>[274]</sup> using potato dextrose agar and malt extract agar. The leaves allowed the colonization of Aspergillus candidus Link ex. Fries, Aspergillus flavipes Bainer and Sartory, Bipolaris nodulosa (Bert and Curt. ex. Sacc.) Shoemaker, Fusarium oxysporum Schlechtendahl. Screening of stem revealed fungal species include Aspergillus candidus Link ex. Fries, Aspergillus niger Tiegh, Cunninghamella blacksleeana Lender, Fusarium oxysporum Schlechtendahl, Rhizopus nigricans Ehrenberg and one unidentified species. The petiole has Aureobasidium pullulans (de Bary) Arnaud. Les and Fusarium oxysporum Schlechtendahl, Penicillum purpurogenum stoll species. Nanda and Nayak, (2015)[275] conducted experiment on leaves and reported the presence of Alternaria alternate, Alternaria geophila, Alternaria tenuis, Botytis cinerea, Brown sterile mycelia, Cladosporium sp, Cladosporium herbarum, Colletotrichum falcatum, Curvularia lunata, Curvularia geniculata, Dreshlerea sp, Fusarium oxysporum, Green sterile mycelia, Geotrichum sp, Helminthosporium sp, Mortierella, Pencillium fumiculosum, Wallemia sebi, white sterile mycelia, Ulocladium langinosum species in young, mature, yellow, infected and dry leaves of A. indica. It was observed that fungal species colonization in A.indica would be responsible for production of huge number of secondary metabolites, bioactive compounds and antimicrobial agents.

#### Larvicidal potential

Larvicidal potentiality of *A.indica* extracts has been studied on *Anopheles* stephensi Liston, *Aedes aegypti* and *Culex. Quinquefasciatus* and *Anopheles* subpictus. In a study, methanol extract showed  $LC_{90}$  concentration 36.32 ppm on *Anopheles stephensi* Liston larvae than benzene, chloroform, ethyl acetate extracts of leaves.<sup>[262]</sup> In another experiment on the same larvae, petroleum ether extract ( $LC_{90}$  of 447.19) was more active than hexane, ethanol, acetone and chloroform extracts but same extracts on *Aedes aegypti* and *Culex. Quinquefasciatus* revealed that hexane extract

 $(LC_{90} \text{ of } 230.40 \text{ ppm})$  and acetone extract  $(LC_{90} \text{ that of } 411.48 \text{ ppm})$  have good activity on *Aedes aegypti* and Culex. *Quinquefasciatus* respectively.<sup>[49]</sup> In another observation, the hexane extract (1000 ppm) on early fourth instar larvae of *C. quinquefasciatus* showed highest mortality rate (about 66 %)<sup>[276]</sup> whereas Teklani and Perera, (2017)<sup>[95]</sup> have found highest mortality rate of acetone extract (100 % at 97 mg/mL) and water extract (100 % at 100 mg/mL). Santhoshkumar *et al.* (2012)<sup>[277]</sup> have reported that acetone extract (100 ppm) shown potential larvicidal activity on *Anopheles subpictus*.

#### Ovicidal activity

Ovicidal property of benzene, chloroform, ethyl acetate and methanol extracts of *A. indica* at different concentrations (25, 50, 75, 100, 125, 150, 175 and 200 ppm) on different ages (3, 6, 9, 12, 15 and 18 hrs) of *A. stephensi* eggs revealed that extracts at higher concentration affected the hatchability rate in younger eggs than older by 17.3%, 24.3%, 29.0% and 13.0 % respectively.<sup>[262]</sup>

## **Oviposition activity**

Various concentrations (0.01 to 0.1 %) of ethanol extract of *A. indica* leaves prevented the deposition of eggs by *A. Aegypti, A. Stephensi* and *C. quinquefasciatus* female mosquitoes. High concentration (0.1 %) of extract prevented the percent laying of eggs about 99.4%, 98.0% and 97.5 % by *A. aegypti, A. stephensi* and *C. quinquefasciatus* respectively. <sup>[49]</sup> Govindarajan *et al.* (2008)<sup>[262]</sup> have reported the contrast results on attraction of *A. stephensi* for deposition of eggs instead of prevention of laying eggs. Hundread ppm of benzene, chloroform, ethyl acetate and methanol extracts of leaves was attracted the *A. stephensi* for deposition of eggs about 90.09%, 94.20%, 85.43% and 95.75% respectively.

#### Mosquito repellent activity

Hexane, ethyl acetate, acetone, methanol, water extracts and essential oils of *A. indica* leaves at 5% concentration repel the *Aedes aegypti*. Among, hexane extract from maceration method and ethyl acetate from sonication have potentially been played 50 % static repellent role for initial 2 hrs, next 3 hrs they maintained 20-30% spatial repellency. Other extracts have shown static repellency but they are failed in spatial repellency. In another experiment, the hexane extract (0.02 ppm) protected the people from *Culex quinquefasciatus, Aedes aegypti and Anopheles stephensi* bite upto 122, 119 and 116 min respectively.<sup>[95]</sup> Hence, hexane extract is a potential mosquito repellent of *A. indica* leaves

#### Anti-plasmodial property

Ethanolic extract of leaves, stem and root have shown anti-plasmodial property on *P. falciparum* (200 µL) cultured in red blood cells for 48 hrs with IC<sub>50</sub> concentrations of 0.056, 0.043 and 0.069 mg/mL but this activity is lower than that of positive controls chloroquine (IC<sub>50</sub> of 0.018 mg/mL) and artemether (IC<sub>50</sub>: 0.005 µg/mL).<sup>[278]</sup>

#### Anti-tuberculosis activity

Anti-tuberculosis property of *A.indica* leaves was studied with water extract at 4% v/v revealed that the extract inhibited the growth of *Mycobacterium tuberculosis* H37Rv, multi drug resistance isolates such as DKU-156 and JAL-1236 about 68%, 95%, and 68% respectively. The same extract didn't exhibit inhibition on fastly growing *M. fortuitum* (TMC-1529) strain.<sup>[268]</sup>

## Insecticidal property

Cotton seeds and leaves (50 g of each) soaked overnight in various concentrations (0.385-6%) of water extract of *A.indica* revealed the insecticidal property. *Dysdercus cingulatus* (Red cotton bug) fed with

	References	[271]	[273]	[272]	[273] <sup>3</sup>	[272]	[271	[271 ]	د [273] ع	[271]	A _ 7 _ 1	[169] .	[270]	[270]	[270]
	CsP		_	_		_	_	_	_	_		_	MFC: [ NA	Ü	MFC: [ 4mg
	CaP	1					'	'		'			MFC: N 2 mg N		MFC: N 2mg 4
	MgD	1	1	I.	1	1	1	1	1	ı	ı	1	MFC: M 2mg 2		MFC: M 2mg 2r
	dqs	1	1	1	1	1	1	ı	1	1	1	1	ü	്	
		ı	1	1	1	1	1	ı	1	1	1	ı			
	CIb				i -			1				1	: MFC: 4mg		: MFC: NA
	Efp			i.						ı.	i.	1	MFC: NA		MFC: 4 mg
g/mL )	ЧţГ				1								MFC: 8mg	MFC 0.5 mg	MFC: 4 mg
and m	q <sub>2</sub> T												MFC: NA	MFC: NA	MFC: NA
µg/mL	٩mT		1	1	1	1	1	1	1	I	ı	1	MFC: 4 mg	MFC: 2mg	MFC: NA
ıg/mL,	ЧЛ												MFC: 8mg	<u>6</u> 0	MFC: 4 mg
Organism tested and effective dose (ng/mL, µg/mL and mg/mL )	MıT		1	1		1	1			1	1		MFC: N NA 8	്ക	MFC: N NA 4
ective	ЭW	1	gu	1	9.3	1	1	ı	8n	ı	1	1	ΣŻ	Μ 4	ΣŻ
and eff	514		30 mg- 1 13 mm		30 mg-9.3 mm				30 mg- NA			1	i.	,	,
tested	CT		30 mg- 10.3 mm		30 mg- NA				30 mg- NA						
anism	ЪС	1	6, <b>H</b>	gm	е) <u>д</u>	gm	'	ı	<del>с</del> ) д	ı	'	ı	ı	ı	ı
Org			- 2	300 mg- 14 mm		300 mg- 10 mm	i.	ı.	NT	ī		1	1	ī	ı
	ΑF	1	30 mg-8.7 mm	300 mg- 28 mm	30 mg- NA	300 mg- 18 mm	1	1	30 mg- NA	1	1	1			1
	nЯА														
		1	ı.		1		I.	i.	I.	I	i.	1	1	1	1
	כפ		I	300 mg- 11mm	1	300 mg- NA	I	I	I	I	I	1	1	1	
	AD	0.05 mg- 13 mm	30 mg- 12.7	300 mg- 18 mm	30 mg-8.7 mm	300 mg- 13 mm	0.05 μg- 20 mm	0.05 μg- 17 mm	30 mg- NA	0.05 μg- 12 mm	MIC: 0.4 mg	500 ng-20 mm			
		0.05 13 r	30 m 12.7		30 m mm		0.05 20 I	0.05 17 I	30 m NA	0.05 12 I	MIC		1	1	1
	NA	I.		300 mg- NA	1	300 mg- NA	I.	I.	I.	ı		500 ng- 22 mm	NA	NA	NA
I	bodtəM	Disc diffusion	Disc diffusion	Well Diffusion	Disc diffusion	Well Diffusion	Disc diffusion	Disc diffusion	Disc diffusion	Disc diffusion	Dilution	Disc diffusion	Dilution	Dilution	Dilution
	בצנומבנז										oform				
	s trad tnalq extracts	L-Acetone	L-Chloroform	L-Ethanol	L-Ethanol	L-Ethyl acetate	L-Methanol	L-Petrolium ether	L-water	L-Water	TP-Chloroform Dilution	TP- Clotrimazole	TP-Ethanol	TP-Hexane	TP-Methanol
	oN.2		2 1	3 ]	4	5	6 1		8 1	9 1	10	11	12	13	14
		1	. 4		N	~,	0		~					. –	. ,

S. No	Activity tested	Model used	Effective dose (Plant extract)	Effective dose (Standard drug)	Mechanism of action	Reference
				Total plant		
	Analgesic	Male Mice	ED:400 mg/kg (Polyphenolic extract)	0.13 mg/ 25 kg (Diclofenac sodium)	Increased latency period	[82]
		Either sex rats	ED:400 mg/kg (Polyphenolic extract)	0.9 mg/200 kg (Diclofenac sodium)	Decreased pain threshold	[82]
		Either sex mice	ED:400 mg/kg (Methanolic extract )	50 mg/kg (aminopyrine)	Inhibited writhing reflexes	[88]
	Antiinflammation	Either sex Rats	ED:250 mg/kg (Methanolic extract)	100 mg/kg (phenylbutazone)	Decreased paw volume	[86]
		Red blood cells of human	Red blood cells of ED:400 mg/kg (Polyphenolic extract) human	3.6 mg/kg (Diclofenac sodium)	Stabilized RBC membrane	[82]
	Anti-Fertility	Female rats	ED:600 mg/kg (Ethanolic extract ) and ED: 600 mg/kg (Petroleum ether extract)	Ethinyl estradiol (1 μg)	Increased ethinyl estradiol levels, diameter of uterus, thickness of endometrium, height of the endometrial and epithelium weight	[234]
	Hepatoprotective	Rats	ED: 300 mg/kg (methanol), ED: 250 mg/kg (methanol fraction)	100 mg/kg (Silymarin)	Decreased liver function markers, total bilirubin, and total cholesterol. Increasing total protein and albumin. Tissue recovery from damage.	[81]
		Rats	ED: 300 mg/kg (70% Ethanol)	100 mg/kg (Silymarin)	Reduced SGOT, SGPT and ALP	[248]
		Rats	ED: 150 mg/kg (70% Ethanol)+ 20 mg/kg piperine	100 mg/kg (Silymarin)	Reduced SGOT, SGPT and ALP	[248]
		Rats	ED: 200 mg/kg (70 % methanol) + 150 mg/ kg (70 % methanol- <i>Centella asiatica</i> )	100 mg/kg (vitamin C)	Decreased lipid peroxidation	[249]
	Anti-diabetic	Male rats	ED: 400 mg/kg (80% Aqueous ethanolic extract)	10 mg/kg (glibenclamide)	Decreased blood glucose levels and increased body weights.	[221]
		Neonate rats	ED: 100mg/kg (Petroleum ether, chloroform, acetone and methanol extracts)	5 mg/kg (glibenclamide)	Decreased blood glucose levels	[242]
		Male rats	ED: 500 mg (methanol: acetone fraction (70:30)		Decreased blood glucose levels	[241]
		in vitro	ED: 0.1 mg/mL (Hexane and chloroform extracts)		Inhibited $\alpha$ - amylase activity	[8]
	Anti-arthritic	Rats	ED: 250 mg/kg (Ethanolic extract) ED: 250 mg (water extract)	100 mg/kg (Ibuprofen )	Decreased paw volume. Alkaline phosphatases activity, SGOT and SGPT	[245]
	Kidney stones digestion	Male rats	ED: 200 mg/kg (Ethanolic extract)	150 μg/kg (Thiazide)	Accelerated Ca $^{2+}$ , Mg $^{2+}$ Na $^+$ and K $^+$ ATPases. Decreasing ACP, ALP, AST and ALT.	[238]
		Either sex rats	ED: 10% w/v (Ethanolic extract)	1	Heal of wound	[231]
	Wound healing	Either sex rats	ED: 1% (Hot water extract)	Povidone-iodine ointment	Heal of wound	[232]
		Chick chorioallantoic	ED: 100 μg/μL (Water extract)	vascular endothelial growth factor	Formed of new blood capillaries	[232]

Continued....

Table	Table 11: Continued					
6	Antioxidant	HddQ	$IC_{50}$ : 616.32 μg/mL (hydro alcoholic), $IC_{50}$ : 911.25 μg/mL (Ethyl acetate fraction), $IC_{50}$ : 554.2 μg/mL (methanol fraction) and $IC_{50}$ : 249.14µg/mL (hexane fraction)	IC <sub>so</sub> : 22.0 μg/mL (Ascorbic acid)	Scavenged the DPPH radical by donating either hydrogen or electron followed by proton	[76]
10	Insecticdal	Dysdercus Cingulatus	ED: 6% and 2% Water extract		Influenced the mortality, alterated Na <sup>+</sup> , K <sup>+</sup> , Ca <sup>+</sup> , changing glucose, protein levels of ovary and testes. Increased Aspertate levels in intestine and fat body	[279]
	Neutralization (toxicants from Lagocephalus lunaris fish)	Male mice	ED: 5g/L		Increased activities of ALT, AST, ALP, SOD, CAT, GPx and HDL-C levels. Decreased diarrhea, creatinine, uric acid, total cholesterol, triglyceride, LDL cholesterol, total bilirubin, direct bilirubin, albumin, globulin, total protein and Glucose. Liver, kidney and heart tissue recovered from damage	[236]
				Leaves		
1	Adulticidal	Haemaphysalis bispinosa and Hippobosca maculate	2000 ppm (Methanolic extract)			[277]
2	Anti ulcer	Either sex rats	200 mg/kg (80% ethanol extract)	50 mg/kg (Ranitidine) and 1 mg/kg (Diazepam)	Protected from acetyl salicylic acid, cold stress, ethanol induced and [250] pylorus ligated ulcers by improving pH, reducing gastric volume, acidity and mean ulcer score	[250]
<i>რ</i>	Antidiabetic	Either sex rats	<ul> <li>ED:400 mg/kg (hydro alcoholic extract),</li> <li>ED:400 mg/kg (Chloroform soluble fraction),</li> <li>ED:400 mg/kg (Butanol soluble fraction),</li> <li>ED:400 mg/kg (Butanol insoluble fraction)</li> </ul>	50 mg/kg (Metformin )	Decreased blood glucose levels	[80]
		Rats	ED:500 mg/kg (Ethanolic extract)	5mg/kg Glibenclamide)	Prevented Nucleic acids decrement. Decreased blood glucose levels, gluconeogenic enzymes, ALP, ACP, AST, ALT, LDH and lipid profile. Elevation of cellular antioxidant, glycolytic and TCA cycle enzymes. Recovery of liver and kidney tissues	[126]
		Male Rats	ED:100 mg/kg (Polyphenolic fraction)	20mg/kg (Glibenclamide)	Elevated cellular antioxidant enzymes	[78]
4	Hepato protective	Male rats	100 mg/kg ( 70% alcohol)		Decreased AST, ALP, LIP, Lipid peroxidation and elevating antioxidants. Protecting tissue damage	[246]
		Male rats	200 mg/kg (Ethanolic extract)		Decreased AST and ALT	[247]
						Continued

Table 11: Continued					
5 Antioxidant	Нда	IC <sub>30</sub> :28.330 μg/mL, 330 μg/mL, 230 μg/mL, 7079 μg/mL; ED: 600 μg/mL (Methanol	IC <sub>50</sub> :2.69 μg/ mL, 4.106 (Ascorbic acid)		[78, 99, 224, 225]
		extract)	$\label{eq:source} \begin{split} IC_{so}; & 3.91 \ \mu g/mL \ (Rutin); \ IC_{so}; \\ & 80 \ \mu g \ /mL \ (Trolax) \end{split}$		
		$\rm IC_{50}$ : 6019 µg/mL (Hexane extract)	IC <sub>50</sub> : 80 μg /mL (Trolax)		[66]
		IC <sub>50</sub> : 5.70 mg/mL (Chloroform extract)	IC <sub>50</sub> : 80 μg /mL (Trolax)	Scavenged the DPPH radical by donating either hydrogen or	[66]
		$IC_{\rm 50}$ : 300.58 $\mu g/mL$ (Poly phenolic fraction)	1	electron followed by proton	[78]
		IC <sub>50</sub> ; 476.19 μg/mL (Ethanolic extract); IC <sub>50</sub> :616.32 μg/mL (70%Ethanolic extract)	IC <sub>50</sub> : 13.1 μg/mL(Tannic acid); IC <sub>50</sub> : 22.0 μg/mL (Ascorbic acid)		[76, 126]
		IC <sub>50</sub> : 179.82 μg/mL (Acetone extract)	179.59 μg/mL (Ascorbic acid )		[83]
		$\rm IC_{s0}$ : 191.25 µg/ mL (Ethyl acetate fraction)	IC <sub>so</sub> : 22.0 μg/mL (Ascorbic acid)		[76]
		$\rm IC_{s0}$ : 554.2 µg/mL (Methanolic fraction)	IC <sub>50</sub> : 22.0 μg/mL (Ascorbic acid)		[76]
		$\rm IC_{50}$ : 249.14 µg/mL (Hexane fraction)	IC <sub>so</sub> : 22.0 μg/mL (Ascorbic acid)		[76]
	ABTS	$\rm IC_{50}$ : 6013 µg/mL (Hexane extract)	IC <sub>50</sub> : 1.32 mg/mL (Trolax),		[66]
		IC <sub>50</sub> : 6031 μg/mL (Chloroform extract)	IC <sub>50</sub> : 1.32 mg/mL (Trolax),		[66]
		IC <sub>so</sub> ; 6037 μg/mL, 5.93 μg/mL (Methanolic extract)	IC <sub>50</sub> : 1.32 mg/mL (Trolax), 0.52 μg/mL (Rutin), 11.25 μg/ mL (Ascorbic acid),		[99, 224]
		$\rm IC_{50}$ : 252.56 µg/mL (Acetone extract)	IC <sub>50</sub> : 225 μg/mL (BHT)	ABTS radical conversion into its stable form	[83]
	Sugar damage by hydroxyl radicals	IC <sub>50</sub> ; 35.933 μg/mL (Methanolic extract)	5.278 μg/mL (Ascorbic acid)	Mitigated hydroxyl radicals	[78]
	Lipid peroxidation by hydroxyl radicals	IC <sub>30</sub> : 84.775 μg/mL and >1000 μg/mL (Methanolic extract)	65.321 μg/mL (Ascorbic acid) and 95.00 μg/mL (BHA)	Mitigated hydroxyl radicals	[78, 224
	Hydrogen	IC <sub>50</sub> : 703.88 μg/mL (Polyphenolic fraction)	1	Converted hydrogen peroxide into water	[226]
	peroxide	IC <sub>so</sub> ; 84.415 μg/ mL, 193.33 μg/ mL (Methanolic extract)	IC <sub>50</sub> : 57.560, 187.33 μg/mL, (Ascorbic acid); IC <sub>50</sub> : 36.16 μg/mL (Rutin)		[78, 224]
		$\rm IC_{50}$ : 272.77 µg/mL (Acetone extract)	IC <sub>50</sub> : 195 μg/mL (Ascorbic acid)		[83]
	Hydroxyl radical	Hydroxyl radical IC <sub>s0</sub> : >1000 μg/mL (Methanolic extract)	IC <sub>50</sub> >1000 μg/mL (Ascorbic acid), IC <sub>50</sub> :205.83 μg/mL (Rutin), IC <sub>50</sub> :>1000 μg/mL (BHA)	Mitigated hydroxyl radicals	[224]
		$IC_{s_0}$ : 284.46 µg/mL (Acetone extract)	225.76 μg/mL (Ascorbic acid)		[83]
		IC <sub>50</sub> : 574.71 μg/mL (Ethanolic extract)	IC <sub>50</sub> : 38.3 μg/mL (Mannitol)		[126]
					Continued

Table 11: Continued	•				
	Superoxide radical	$\rm IC_{50}$ : 243.68 µg/mL (Acetone extract)	IC <sub>50</sub> : 148.53 μg/mL (Ascorbic acid)	Scavenged superoxide radical	[83]
		$\rm IC_{50}$ : 568.18 µg/mL (Ethanolic extract)	IC <sub>50</sub> : 52.1 μg/mL (Quercetin)		[126]
	Nitric oxide	$IC_{50}$ : 14.77 µg/mL (Methanolic extract),	IC <sub>50</sub> :68.44 μg/mL (Rutin)		[224]
		$\rm IC_{s_0}$ : 203.56 µg/mL (Acetone extract)	IC <sub>50</sub> : 231.24 μg/mL (Ascorbic acid)	Scavenged nitric oxide	[83]
		and $\rm IC_{50}$ : 427.35 µg/mL (Ethanolic extract)	IC <sub>50</sub> : 43.1µg/mL (Curcumin)		[126]
	Reducing power	ED:500 µg/mL (Polyphenolic fraction),	ED:500 μg/mL (Ascorbic acid)		[226]
		ED:100 µg/mL (Methanolic extract)	ED:100 μg/mL (Ascorbic acid)	Converted of Fe <sup>3+</sup> to Fe <sup>2+</sup>	[78]
		IC <sub>50</sub> :202.86 (Acetone extract)	IC <sub>so</sub> :244.52 μg/mL (Ascorbic acid)		[83]
	Total antioxidant	ED:1000 µg/mL (Polyphenolic fraction)	ED:1000 μg/mL (Ascorbic acid)	Reduced of Mo (VI) to Mo (V)	[226]
		ED:100 µg/mL (Methanolic extract)	ED:100 μg/mL (Ascorbic acid)		[78]
		$\rm IC_{s_0}$ : 2.86 µg/mL (Methanolic extract)			[224]
		IC <sub>50</sub> : 231.48 µg/mL (Acetone extract)	IC <sub>50</sub> : 196.80 μg/mL (Ascorbic acid)		[83]
6 Cytotoxicity/ Cancer	Small cell lung cancer	IC <sub>50</sub> : 25 μg/mL (Methanolic extract)	IC50: 0.88 $\mu g$ /mL (Ellipticine) Induced cell death and $IC_{30}^{\rm :}: 0.05  \mu g/mL \label{eq:result}$ (Doxorubicin)	Induced cell death	[66]
	Squamous skin	ED:100 μg/mL (Hexane extract)		Cytotoxicity	[227]
	cancer	$\mathrm{IC}_{\mathrm{50}}$ : 78 µg /ml (Ethyl acetate extract)	,	Cytotoxicity and inhibition of 12 R LOX	[227]
	prostate cancer	ED: 70 µg/mL (Ethanolic extract loaded casein-chitosan microparticles)		Cytotoxicity; LDH elevation	[228]
	Breast cancer	ED:100 μg/mL (Aqueous extract loaded silver and gold nanoparticles)		Cytotoxicity, apoptosis, DNA damage	[229]
		ED:100 µg/mL Aqueous extract loaded copper oxide nanoparticles	-	Cytotoxicity	[230]
7 Larvicidal	Anopheles subpictus	100 ppm (Acetone extract)		Lethality shown on laraval stage of mosquitoes	[277]
	Aedes aegypti	LC <sub>90</sub> : 462.17 ppm (Acetone extract), LC <sub>90</sub> : 1059.78 ppm (Chloroform extract), LC <sub>90</sub> : 230.40 ppm (Hexane extract), LC <sub>90</sub> : 336.75 ppm (Petroleum ether extract) and LC <sub>90</sub> : 1198.85ppm (Ethanol extract)		Lethality shown on laraval stage of mosquitoes	[49]
		ED:97 mg/mL (Acetone extract from soxhlet method), ED:100 mg/mL (Water extract from		Lethality shown on laraval stage of mosquitoes	[95]
		sonication method)			

Table 11	Table 11: Continued				
			ED: 1,000 ppm (Acetone, Chlorform, ethyl - acetate, hexane, Methanol extracts)	Lethality shown on laraval stage of mosquitoes	[276]
		Culex quinquefasciatus	LC <sub>90</sub> : 411.48 ppm (Acetone extract), - LC <sub>90</sub> : 760.38 ppm (Hexane extract), LC <sub>90</sub> : 950.80 ppm (Petroleum ether extract ), LC <sub>90</sub> : 1181.55 ppm (Ethanol extract) and LC <sub>90</sub> : 695.55 ppm (Chloroform extract).	Lethality shown on laraval stage of mosquitoes	[49]
		Anopheles stephensi	LC <sub>90</sub> : 41.29 ppm (benzene extract), - LC <sub>90</sub> : 58.27 ppm, (chloroform extract), LC <sub>90</sub> : 49.19 ppm (ethyl acetate extract), LC <sub>90</sub> : 36.32 ppm (methanolic extract)	Lethality shown on laraval stage of mosquitoes	[262]
			LC $_{90}$ : 1210.9ppm (chloroform extract), - LC $_{90}$ : 660.9 ppm (Hexane extract), LC $_{90}$ : 447.19 ppm (Petroleum ether extract), LC $_{90}$ : 1779.62 ppm (Ethanolic extract) and LC $_{90}$ : 660.9 ppm (Acetone extract)	Lethality shown on laraval stage of mosquitoes	[49]
8	Ovicidal	Anopheles stephensi	200 ppm ( Benzene, chloroform, ethyl acetate - and methanolic extracts)	Killed the mosquitoes eggs	[262]
6	Oviposition	Anopheles stephensi;	100 ppm ( Benzene, chloroform, ethyl acetate - and methanol extracts)	Attracted the mosquitoes to laying of eggs.	[262]
		C. quinquefasciatus, A. stephensi and A. aegypti.	0.1 % (Ethanolic extract) -	Greatly Reduced the laying of eggs	[49]
10	Mosquito repellent Aedes aegtpti, Anopheles stephensi and Culex. quinquefascia	Aedes aegtpti, Anopheles stephensi and Culex. quinquefasciatus	0.02 ppm (Hexane extract) -	Repelled mosquitoes from biting. Spatial repellencies of mosquitoes [49]	[49]
					Continued

Acades aggipti     5 % (Hexane extract-Maceration), 1.5 % (Hexane extract-Sonication), 1.5 % (Ethyl acetate extract-Sonication), 1.5 % (Ethyl acetate extract-Sonication), 2.6 % Acetone extract - Sonication), 1.5 % (Methanol extract-Sonication), 1.5 % (Methanol extract-Sonication), 1.5 % (Methanol extract-Sonication), 1.5 % (Mater extract), 1.5 % (Mater extract), 1.6 % (Mater extract), 1.7 % (Mater extrac					
Cardio protective Male rats Female rats Female rats Anti plasmodial Plasmodium falciparum Male Rats Male Rats Male mice Frog (snake venom) Mice RBC Wound healing Male rats	Aedes aegtp				[95]
Female rats         Anti plasmodial       Female rats         Anti plasmodial       Plasmodium         falciparum       Male Rats         Male Rats       Male mice         Frog       Male mice         Neutralization       Frog         (snake venom)       Mice         Wound healing       RBC         Wound healing       Male rats		ED:200 mg/kg (Flavonoid rich extract )	1	Restored plasma, serum markers, antioxidant enzymes, lipid profiles [239, 240] and cardiac tissue dammage	s [239, 240]
Female rats       Anti plasmodial     Plasmodium       Anti plasmodial     Plasmodium       falciparum     Male Rats       Male Rats     Male Rats       Neutralization     Frog       (snake venom)     Mice       KBC     Wound healing       Male rats     RBC	Female rats	ED:500 mg/kg (70%Methanolic extract)		Restored serum markers, antioxidant enzymes and cardiac tissue damage	[263]
Anti plasmodialPlasmodium56.89 81µg/mL (Ethanolic falciparumfalciparum56.89 81µg/mL (Ethanolic extrMale RatsED:750 mg (Ethanolic extrMale miceED:750 mg (Ethanolic extrFrogED:1.6 mg/mL (Ethanolic.NeutralizationMiceED:1.6 mg/mL (Ethanolic.NeutralizationMiceED:1.6 mg/mL (Acetone e(snake venom)MiceED:0.4 mg/mL (Acetone eWound healingMale ratsED:0.4 mg/mL (Acetone e	Female rats	ED:500 mg/kg (70%Methanolic extract)		Reverted the CRP, CK-MB and Troponin; maintaining architecture of left ventrical tissue	[263]
Male RatsED:750 mg (Ethanolic extMale miceED:750 mg (Ethanolic extFrogED:1.6 mg/mL (Ethanolic extNeutralizationFrog(snake venom)MiceBD: 500 mg/kg (Acetone eRBCED:0.4 mg/mL (Acetone eWound healingMale ratsED:40 mg/kg (aqueous et)		56.89 81μg/mL (Ethanolic		Showed antiplasmodial activity	[278]
Male mice     ED:750 mg (Ethanolic ext       Frog     ED:1.6 mg/mL (Ethanolic       Neutralization     BD:1.6 mg/mL (Ethanolic       (snake venom)     Mice     ED:500 mg/kg (Acetone e       RBC     ED:0.4 mg/mL (Acetone e       Wound healing     Male rats     ED:40 mg/kg (aqueous eti	Male Rats	ED:750 mg (Ethanolic extract )	Snake venom antiserum	Inhibited haemorrhage, necrosis and mast cell degranulation	[223]
Frog     ED:1.6 mg/mL (Ethanolic       Neutralization     ED:1.6 mg/mL (Ethanolic       (snake venom)     Mice     ED: 500 mg/kg (Acetone e       RBC     ED:0.4 mg/mL (Acetone e       Wound healing     Male rats     ED:40 mg/kg (aqueous et)	Male mice	ED:750 mg (Ethanolic extract )	Snake venom antiserum	Neutralized the venom	[223]
(snake venom) Mice RBC Wound healing Male rats			1	Inhibited venom on neuro and cardic cells	[223]
RBC Wound healing Male rats		ED: 500 mg/kg (Acetone extract).	Snake venom antiserum	Neutralized snake venom	[235]
Wound healing Male rats	RBC	ED:0.4 mg/mL (Acetone extract)		Inhibited heamolysis	[235]
		ED:40 mg/kg (aqueous ethanol)		Mitigated oxidative stress, lipid peroxidation, increased ascorbic acid, improving cell proliferation ; positive action on TNF- $\alpha$ and TGF- $\beta$ 1, collagen synthesis, collagen 1 $\alpha$ and collagen 3 $\alpha$ .	[233]
16 Haemolysis Humans Broth		Broth		Caused changes in peripheral blood, reticulocytosis, increase levels of serum indirect bilirubin and haemoglobinuria	[5]

Pharmacognosy Reviews, Vol 15, Issue 30, Jul-Dec, 2021

Continued....

	[227]	[237]		[278]	ld [250]	[244]	[224, 225]	[83]	[224]	[83]	[224]	[83]	[224])	[83]	[83]	[224]	[83]	[224]	[83]	
	Inhibited 5-LOX, 15 LOX, COX-1 and COX-2 enzymes	Inhibited 12 R-Lox			Protected from acetyl salicylic acid, cold stress, ethanol induced and [250] pylorus ligated ulcers by improving pH, reducing gastric volume, acidity and mean ulcer score	Caused paralysis and death		Scavenged DPPH radical		Scavenged AB TS radical	Converted hydrogen peroxide to water		Scavenged hydroxyl radical		Scavenged superoxide radical	Mitigation of hydroxyl radicals	Converted of $\mathrm{Fe}^{3+}$ to $\mathrm{Fe}^{2+}$		Reducted of Mo (VI) to Mo (V)	
		0.07 mg/mL (Zileuton)			50 mg/kg (Ranitidine); 1 mg/kg (Diazepam)	10mg/mL (Albendazole )	IC <sub>50</sub> : 2.69 μg/mL (Ascorbic acid); IC <sub>50</sub> : 3.91 μg/mL (Rutin)	179.59 μg/mL (Ascorbic acid)	IC $_{50}$ : 11.25 μg/mL (Ascorbic acid); IC $_{50}$ : 0.52 μg/mL (Rutin);	IC <sub>50</sub> :288.53 μg/mL (BHT)	IC <sub>50</sub> : 187.33 μg/mL (Ascorbic acid); IC <sub>50</sub> : 36.16 μg/mL (Rutin)	IC <sub>50</sub> : 195 μg/mL (Ascorbic acid)	IC50: >1000 μg/mL, (Ascrobic acid); IC <sub>50</sub> : 205.83 μg/mL (Rutin); IC <sub>50</sub> : > 1000 μg/mL (BHA)	IC50: 225.76 μg/mL	IC <sub>50</sub> : 148.53 μg/mL (Ascorbic acid)	IC <sub>50</sub> : 95.00 μg/ml (BHA)	IC <sub>50</sub> : 244.52 μg/mL (Ascorbic acid)		IC <sub>50</sub> : 196.80 μg/mL (Ascorbic acid)	
	ED:0.1 mg/mL (Ethylacetate and Hexane and Ethanol extract	ED: 0.3 mg (Ethyl acetate and water extracts) 0.07 mg/mL (Zileuton)		$\rm IC_{50}$ : 69 µg/mL (Ethanolic extract )	200 mg/kg (80% ethanol extract)	50 mg/mL (70% alcohol)	$\rm IC_{50}$ : 208.5 and ED: 600 µg/mL (Methanolic extract)	IC50: 227.02 μg/mL (Acetone extract)	$\rm IC_{50}$ : 24.00 µg/mL (Methanolic extract)	and $\mathrm{IC}_{\mathrm{50}};288.53~\mathrm{\mu g/mL}$ (Acetone extract)	$\mathrm{IC}_{\mathrm{50}}$ : 205.00 µg/mL(Methanolic extract)	IC5 <sub>0</sub> : 293.11 μg/mL (Acetone extract)	Hydroxyl radical IC <sub>30</sub> : >1000 µg/mL (Methanolic extract)	IC <sub>50</sub> : 288.61 μg/mL (Acetone extract)	$\rm IC_{50}$ : 277.78 µg/mL (Acetone extract)	$IC_{\rm 50}$ : 180.00 µg/mL (Methanolic extract)	$IC_{50}$ : 224.13 µg/mL (Acetone extract)	Total antioxidant $~~{\rm IC}_{\rm so}$ : 1.53 µg/mL (Methanolic extract)	$\mathrm{IC}_{\mathrm{so}}$ ; 253.12 µg/mL (Acetone extract)	
	In vitro			Plasmodium falciparum	Either sex Rats	Pheretima posthuma	HddC		ABTS		Hydrogen peroxide		Hydroxyl radical		Superoxide radical	Lipid peroxidation	Ferric reducing	Total antioxidant		
-	Table 11: Continued           18         Inflammation		Roots	Anti plasmodial	Anti Ulcer	Anthelmintic	Antioxidant													
	<b>Table 1</b> 18			1	ŝ	4	2													

Table 1	Table 11: Continued					
		Nitric oxide	IC50 : >700 μg/mL (Methanolic extract)	IC <sub>50</sub> : 68.44 μg/mL (Rutin)		[224]
			IC <sub>s0</sub> : 255.05 µg/mL (Acetone extract)	231.24 μg/mL (Ascorbic acid)	Scavenged nitric oxide	[83]
			IC <sub>50</sub> : 315.46 μg/mL (Ethanolic extract)	IC <sub>50</sub> : 103.59 μg/mL (Ascorbic acid)		[79]
			IC <sub>50</sub> : 667.82 µg/mL (Aquoes extract)	IC <sub>50</sub> : 103.59 μg/mL (Ascorbic acid)		[62]
	Stern					
1	Anti plasmodial	P.falciparum	$IC_{50}$ : 43.81µg/mL (Ethanolic extract )		Showed antiplasmodial activity	[278]
7	Diabetes	Rats	ED:600 mg/kg (Methanolic extract)	5 mg/kg (Acarbose)	Suppressed postprandial glucose and sucrose elevation.	[243]
		Rats	ED:300 mg/kg (Methanolic extract)	10 mg/kg (Metformin)	Decreased liver markers and elevating antioxidant enzymes	[243]
ŝ	Free radical scavenging	DPPH	$\mathrm{IC}_{\mathrm{sc}}$ ; 212.83 µg/mL (Methanolic extract)	IC <sub>50</sub> : 2.69 μg/mL (Ascorbic acid), IC <sub>50</sub> : 3.91 μg/mL (Rutin)		[224]
			and IC $_{\rm 50}$ ; 218.28 µg/mL (Acetone extract)	179.59 μg/mL (Ascorbic acid)	Scavenged the DPPH radical	[83]
		ABTS	IC <sub>30</sub> : 14.33 μg/mL (Methanolic extract);	IC $_{50}$ ; 11.25 μg/mL (Ascorbic acid); IC $_{50}$ ; 0.52 μg/mL (Rutin)	Scavenged ABTS radical	[224]
			IC <sub>50</sub> : 323.09 (Acetone extract)	and IC <sub>50</sub> :225 $\mu$ g/mL (BHT)		[83]
		Lipid peroxidation	$\mathrm{IC}_{\mathrm{so}}$ ; 700.00 µg/mL (Methanolic extract)	IC <sub>50</sub> : 95.00 μg/mL (BHA)	Mitigated hydroxyl radicals	[224]
		Hydrogen peroxide	IC <sub>50</sub> : 380.00 μg/mL (Methanolic extract)	IC50: 187.33 μg/mL, (Ascorbic Converted hydrogen peroxide acid); IC50: 36.16 μg/mL to water (Rutin)	Converted hydrogen peroxide to water	[224]
			IC <sub>50</sub> : 333.55 μg/mL (Acetone extract)	195 μg/mL (Ascorbic acid)		[83]
		Superoxide radical	$\rm IC_{50}$ ; 308.61 µg/mL (Acetone extract)	IC <sub>50</sub> : 148.53 μg/mL (Ascorbic acid);		[83]
		Ferric reducing	$\rm IC_{50}$ ; 320.19 µg/mL (Acetone extract)	IC <sub>50</sub> : 244.52 μg/mL (Ascorbic acid);	Converted Fe <sup>3+</sup> to Fe <sup>2+</sup>	[83]
		Hydroxyl radical	Hydroxyl radical IC <sub>50</sub> : 873.33 μg/mL (Methanolic extract)	$\begin{split} IC_{so}:>1000 \ \mu g/mL(Ascorbic acid); \ IC_{so}:>205.83 \ \mu g/mL (Rutin) \ and \ IC_{so}:>1000 \ \mu g/mL (BHA) \\ mL (BHA) \end{split}$	Scavenged hydroxyl radical	[224]
			$\rm IC_{50}$ : 317.33 µg/mL (Acetone extract)	IC <sub>50</sub> : 231.24 (Ascorbic acid)		[83]
		Nitric oxide	IC <sub>50</sub> : 490.00 μg/ml (Methanolic extract) and	IC <sub>50</sub> : 68.44 μg/ml (Rutin)	Scavenged nitric oxide	[224]
			IC <sub>50</sub> : 255.05 μg/mL (Acetone extract)	IC <sub>50</sub> : 231.24 µg/mL (Ascorbic acid)		[83]
		Total antioxidant	Total antioxidant IC <sub>50</sub> : 2.27 µg/mL (Methanolic extract) and	IC <sub>50</sub> : 196.80 µg/mL µg/mL	Reduced Mo (VI) to Mo( V)	[224]
			$IC_{s_0}$ : 244.24 µg/mL (Acetone extract)	(Ascorbic acid)		[83]

ED: Effective dose;  $\mathrm{IC}_{\mathrm{so}}$  Fifty percent inhibitory concentration

leaves and seeds for 96 hrs shown 77.4% and 49.5% mortality at higher concentration respectively by causing alterations in intestine, fat body and reproductive system. The red bug fed with cotton seeds and leaves having 2% extract revealed that seeds decreased the gut Na<sup>+</sup> whereas leaves decreased the K<sup>+</sup> and Ca<sup>+</sup>; both increased the glucose and protein levels in testes; seeds increased the aspertate in intestine and fat body.<sup>[279]</sup>

# DISCUSSION

## General information

*A.indica* has 20 synonyms which are available in "The plant database" and "Kew-Royal botanical garden". Due to the medicinal importance of this plant, many taxonomists have focused on it and given synonyms. At the same time, the local ethnics and accents of Asia and African countries have given many vernacular names. Especially, it has different vernacular names in Indian regional languages because it is a common weed growing in public places and the traditional healers used this plant for many health problems.

## Traditional clues

In modern era, apart from synthetic drugs, most of the drugs are identified for diseases from traditional information of extracts/decoctions/ paste etc., of plants/other natural sources or from skeleton moieties of plant compounds. Simply, building of modern medicine is constructed on pillars of traditional information of medicinal plants. The traditional information of A.indica existed with healers of African (Seychelles, Namibia, Mozambique, Réunion etc., ) and Asian countries (India, Srilanka, Nepal, Bangladesh etc.,) where they used this plant sources (whole plant, leaves, stem, root and seeds) as decoction, paste, juice, sap and others. These sources are used to treat asthma, bronchitis, burns, cough, diarrhea, dog bite, ear ache, epilepsy, skin infections, joint pains, rheumatoid arthritis, snake bite, ulcers, wounds, syphilis etc., (Table 1). Traditional information available on A.indica provides clue to the modern science for identification of potential medicine for respective ailments. Extensive research reports are also available on A.indica and strongly supported the traditional uses. All scientific evidences in connection with traditional data will be discussed in the following sections of this review.

## Phytochemistry

People show importance to the plants majorly due to the presence of medicinally useful secondary metabolites. In this connection, *A.indica* has been used by traditional healers against many ailments, this is followed by the researchers started extraction of phytochemicals into methanol, diethyl ether, ethyl acetate, ethanol, aqueous alcohol, petroleum ether, chloroform and water from total plant, leaves, roots and stem. Consequently they found flavanones, flavonoids, phenols and saponins quantitatively. Among these findings, our research group<sup>[78]</sup> reported rich content of flavonoids, phenols and saponins in methanolic extract of *A.indica* leaves. We also identified methanol, ethyl acetate and water extracts of *A. indica* leaves are hygroscopic in nature (unpublished data from our study). This query has been conquered by applying lyophilization method to get solid or powder forms and also taken further care in storage to get accurate weighing and results.

Isolation of bioactive compounds is very essential element in phytochemistry because it gives fruitful functional property of the plants. Accordingly, an attempt on *A.indica* total plant and leaves for isolation of bioactive compounds is more when compared with stem, root and inflorescences. This plant is very precise in having alkaloids and their glycosides because few of them are cynogenic glycosides, a class of toxic compounds. Flavonoids, tannins, coumarins, phenols, fatty acids, steroids and terpenes/terpenoids are othe constituents.

## Cynogenic compound

In a list of 15 alkaloids, seven of them are cynogenic glycosides (1-4, and 9-11 in Table 4) which are identified higher in quantity from methanolic extract of leaves and inflorescence (0.35% on fresh weight basis) than roots and stem whereas no such toxic compounds are found in seeds.<sup>[87]</sup> These are considered as hydrogen cyanide (HCN) releasing phytotoxins, produced against pathogens, herbivores and are very harmful to humans if consumed.

After having information on phytochemistry and pharmacology of A.indica, everyone thought that why cyanide toxicity reports are not available on animal studies for phytoextracts of A.indica?. Hence, in this review we are providing some possible reasons i) As per WHO, cynogenic glycosides show toxicity when its range present in between 0.5 to 3.5 mg HCN per kg/b.w.<sup>[281]</sup> The A.indica extracts used for animal studies might have lower than 0.5 mg of HCN ii) Sulfur containing amino acids of rhodanase enzyme present in liver of animals detoxifies the HCN to thiocynide and thiocyanic acid which are excreted through urine. iii) The hydroxocobalamin (B12a vitamin) of liver converted into cynocobalamine (B12 vitamin) by the process of detoxification of HCN.<sup>[281]</sup> iv) Soaking, boiling, fermentation and drying of plant material also remove the cyanide.<sup>[282]</sup> Generally, sequential procedure of drying, phytoextraction with solvents by soaking, concentration using rotary evaporator at suitable boiling temperature of solvents and biological evaluation of A.indica plant/parts might reduce the cynogenic glycosides.

# Polyphenols

Due to the presence of hydrogen bond donor and acceptors, polyphenols of plant playing crucial role in protection of biomolecules by stabilizing free radicals. Apart from they also act as inhibitors against therapeutic targets of deadly diseases. *A.indica* has 27 polyphenols including flavonoids (10 numbers), tannins (11 numbers), coumarins (3 numbers), hydroxy benzoate (2 numbers) and hydroxy cinnamic (1 number). Diseases management of these compounds is summarized in Table 3. *A.indica* extracts having rich content of polyphenols showed potential antioxidant activity against DPPH, Hydroxyl radical, Hydrogen peroxide, lipid peroxidation etc., (Table 12).

## Volatile compounds

The volatility of plants is a sign of language, useful for the communication and interaction with surrounding environment. Flowers, leaves, and fruits being released the volatile compounds into air for attraction of pollinators and defense against herbivores, parasites, bacteria, and fungus whereas roots release into soil for the protection of pathogens. These volatiles are also giving major assistance in curing of diseases. GC-MS and HR-LC-MS analysis of *A.indica* provided volatile compounds which come under aldehydes, alkanes, esters, ethers and fatty acid derivatives (fatty acyls and fatty alcohols). The volatilization (smell) of *A.indica* also found when the plant leaves are crushed with hands as well extraction with low polar organic solvents.

## Pharmacological properties

*A.indica* extracts from whole plant, leaves, stem and root have used for animals studies (Table 12). The abundant usage order of this plant extracts are leaves> total plant> roots> stem. The reason behind the choice of plant leaves is due to having plenty of secondary metabolites resulted from the stress of biotic and abiotic factors. Based on various biological activities of *A.indica*, we categorized them into stabilization, killing/inhibition, protection, reduction and neutralization.

#### Stabilization

Free radicals are the molecules having one or more unpaired electrons in their outer shell, so they pair/interact with adjacent moieties (biomolecules in biological system) to get stable form. It results the adverse effects like DNA damage, protein degradation, lipid peroxidation etc. In this scenario, A.indica stabilizeed various radicals (DPPH, hydroxyl, superoxide, etc., as summarized in Table 12) due to its antioxidant polyphenolic compounds. Antioxidants donate either electron or electron followed by hydrogen to the radicals. Among extracts and fractions of A.indica, the methanolic extract exhibited potential radical stabilization property, which is equal to the standard antioxidant (ascorbic acid). As per Do et al. (2014),<sup>[283]</sup> highly polar phenols extracted more into methanol than water and also mixing of high polar water to other solvents like methanol, ethanol and acetone reduce the extraction of phenols. This extraction procedure and stabilization properties are strongly support the quantified phenolic content (Table 2), isolated and identified polyphenolic (tannins, flavonoids, coumarins and phenols etc.,) compounds (see in phytochemistry description) of A.indica. On the whole A.indica could be used as alternative antioxidant medicine for the radicals associated deadly diseases.

#### Killing/ inhibition

*A.indica* unveiled growth inhibition or killing ability on disease causing cells/organisms like bacteria, fungus, cancer, mosquitoes larvae, round worm parasite resembled *Pheretima posthuma*, *Plasmodium falciparum* and *Dysdercus cingulatus* (Red cotton bug).

Various solvent extracts/ fractions from leaf, root, stem and total plant of *A.indica* exhibited antimicrobial and antifungal properties. Among, the dilution method is a very reliable and accurate for assessing effective concentrations than disc diffusion and well diffusion. Extracts potentially exhibited antimicrobial activity on bacteria (*E.coli* and *S. auereus* summarized in Table 10) and fungus (*C.albicans*). Hence, to identify potentiality of extracts, researchers could choose dilution method/ other advanced methods and instruments.

The information available as on this date is insufficient to decide whether this plant possess cytotoxic/anticancer potentiality because of any *in vitro* report is not available on toxic effects of extracts on normal cells. Though, extracts at higher doses are not shown toxicity in invivo, its direct exposure to cells (cell lines, where no detoxification occurs) in *in vitro* may be possible. Moreover, all experiments on cancer have been conducted using cell lines and obtained positive results but many times the *in vitro* activities are failed in animal models. Hence, further extensive investigation on animal models with different dose ranges would clarify these remarkable queries.

Extracts of *A.indica* shown potential larvicidal and ovicidal activities at different concentrations (Table 12); hence, the synergistic study of *A.indica* extracts with other pesticides and mosquito predators would require to establish the toxic properties of this plant. Moreover, water extract of *A.indica* exhibited killing capacity on Red cotton bug (insect) with good mortality rate; ethanolic extract killed the *Plasmodium falciparum* cultured in RBC; and *Pheretima posthuma* has been killed by hydro alcoholic (70%) extract effectively than standard drug abendazole but in all cases, the results are not compared with standard drugs except in red cotton bug and *Pheretima posthuma*. Researchers who done work on this plant also not isolated any active compounds (except L-Quebrachitol isolated from leaves by Sanseera *et al.* (2012) from bioactive extracts. Hence, the comparative study on other solvent extracts of leaves, roots and stem of *A.indica* with reference drugs would help in isolation of future toxic constituents.

#### Protective property

A.indica shown protective property on cardiac, hepatic, liver and kidneys damage induced in experimental animals as follows: i) Methanolic and hydro methanolic (70%) extracts of leaves protected the isoproterenol, furosemide and potassium chloride induced cardiac damage in rats. ii) The leaves broth protected the intra vascular haemolysis in glucose-6phosphate dehydrogenase enzyme deficient humans. iii) In STZ induced diabetic condition, ethanolic extract protected the liver and kidney tissues. iv) Alongside, thioacetamide, paracetamol and CCL, induced liver damage is protected by methanolic extract from methanolic fraction and hydro alcoholic extracts of leaves and whole plant. Among the above results, 70% alcoholic extract (300 mg/kg) is a very potential cardio protective comparatively with standard silymerin (100 mg/kg) but other extracts are not reached upto the activities of standard drugs. Hence, the hydro alcoholic extracts of A.indica to be used in the preparation of herbal drug/ identifiaction of active moieties for the treatment of cardiac damage in future.

### Reduction

Biological experiments on A.indica proved that it has reduction potentiality on wounds, pains and inflammation. It is a potential wound healer than standard ointment povidone, good analgesic agent and anti-inflammatory source. Generally, there is an interconnection between wounds, pain and inflammation. These properties also supported by traditional information on this plant as wound healing agent on animals, treatment for joint pains and arthritis, antiseptic etc (Table 12). Through phytochemistry, the reduction of inflammation is strongly supported by having anti-inflammatory compounds such as n-(2hydroxyethyl)palmitamide (Propylene glycol), catechin, quercetin 3-0-β-D-glucoside, rutin, kaempferol, n-tris[hydroxymethyl]methyl2-aminoethanesulfonic acid, gallic acid, caffeic acid, sulindac sulfide, dimethylglycine, ibuprofen, beta-sitosterol and stigmasterol (Table 3). Usually, majority of antiinflammatory compounds can reduce the pain. Gathering, this reduction property of A.indica is due to active phytochemical compounds, isolation would be taken up, evaluated in experimental animals and further studies may be extended to human beings.

### Neutralization

Snake bite is aone of the public health problem in India, 2 Lakh people are being preyed, among 35 to 50 thousand death cases have been recorded annually. Traditional healers have used plant medicine against snake bites, like wise *A.indica* used traditionally as venom neutralizer, also been proved in biological experiments (Table 1). The same has been executed experimentally in animals. Shirwaikar *et al.*  $(2004)^{[223]}$  and Rajendran *et al.*  $(2010)^{[235]}$  have reported that *A.indica* works equal to the anti-snake venom. To date, the potential neutralizing extracts (aqueous ethanol an acetone) reported on this plant but no one identified active constituents of these extracts. So, future researchers should take up and cover this scientific gap by isolation and evaluation of venom neutralizing compounds from bioactive extracts of *A.indica*.

Presence of endophytic fungal species in *A.indica* is helpful to the plant but it consumption by humans is not safe. Hence, people may be advised to use this plant source with the help of organic solvents or heat resulted decoction.

The bioremediation is emerging, ecofriendly, lesser cost technology for the removal of environmental pollutants such as dyes and heavy metals to improve the quality of environment. Biosorption of dyes by *A.indica* helps the environmentalists to get effective solution for removal of hazardious dyes (carcinogens, mutagenic and effects on aquatic biota) and heavy metals. Further step should be taken to cultivate *A.indica* at highly polluted areas to reduce toxic effects.

## CONCLUSION

The current review article reports detailed information about A.indica traditional knowledge, phytochemistry and pharmacological properties. A.indica has been used by local/ traditional healers of Asia, Africa and American countries for wound healing, snake bites, asthma, cough, bacterial infections, dog bite etc. (Table 1). In India all parts of the plant/ total plant are being used for treating of various diseases. Traditionally, the effectiveness of this plant also considered to increase when it combined with other plant/ plant products. It is rich in phenolic content, as well as flavonoids. The phytochemistry of this plant revealed that it has mostly polyphenols (phenols, flavonoids, tannins and coumarins), alkaloids and their glycosides (few of them are toxic), volatile compounds and fatty acids. So far there is no evedance for toxic property on this plant even at higher concentration in animal studies, which might be due to elimination of such toxic agents during processing (at extraction, drying of extracts and detoxification in animal body) to use and detoxicfication in animal body. Biological or pharmaceutical studies showed that A.indica is a potential anti-microbial, anti-diabetes, anti-inflammation, larvicidal, anti-oxidant, wound healing and venom neutralizing agent.

## Scientific gaps

Several scientific gaps need to be highlighted based on this review about A.indica i) Geographical distribution of A.indica and published traditional knowledge is not correlated. The published reports available are only from few Asian (India, Srilanka, Nepal, Bangladesh, Indonesia and Thailand) and African (Seychelles, Namibia, Djibouti, Mozambique, Reunion, Madagascar and Mauritius,) countries but is occupied many areas as described elsewhere in this review hence, research on other places where this plant is distributed to be carried out and to be published to support the pharmaceutical evaluations against diseases. ii) The biological evaluation of the isolated compounds is in many cases lacking. The potentiality of the plant can only proved when biological evaluation of isolated compounds from bioactive extracts are done focusing on the relevant diseases. Clinical studies, mechanism of action and effective doses for the bioactive extracts, pharmacokinetic and pharmacodynamics evaluation to the bioactive compounds are lacking. iii) Many traditional uses (Table 1) have not been evaluated experimentally yet, including asthma, burns, diarrhoea, dog bite, epilepsy, haemorrhoids, constipation, aches of stomach/ear/head, syphilis, wheezing etc., of traditional information based experiments to be conducted in future at preclinical then clinical stage to explore the strength of A.indica in pharmacy/medicine. iv) The extracts which showed potential activity will be properly utilised for development of drug candidates. v) All pharmacological properties except few (see in pharmacological properties) are reported with preliminary evidences, so these activities must be extended extensively with different dosage studies, various modes of experimentation, molecular mechanism, and phytochemicals responsible. vi) A.indica has been utilized by traditional healers as antifertility agent, also was proven in experimental animals pharmaceutically, hence it is suggested that, pregnant women should avoid this plant as remedy to treat any health issues to them.

In future, this plant would be utilized and highly beneficial for pharmacological evaluations of the overlooked traditional applications particularly on snake bites, oragan specific aches, asthma and microbial related issues. The effective crude extracts could be utilized for bioactives isolation to assist in therapy of diseases and drug development. Isolation and extract detoxification process of cynogenic compounds of this plant need to be explored to prove this plant is non-toxic for further clinical applications in the interest of human health.

#### Authors' contribution

The first and second authors played substantial role in data acquisition, analysis, interpretation and manuscript preparation. The other co-authors contributed their efforts equally towards acquiring additional data making script in good way by their expertise in their respective research fields. The corresponding author critically revised and finalized the manuscript for publication.

## ACKNOWLEDGEMENT

All authors express sincere gratitude to the University Grants Commission New Delhi, India for the financial assistance, especially to the SR and PJ in the form of RGNF (No: F117.1/2012-13/RGNF-2012-13-SC-AND 34335 and F1-17.1/2015-16/RGNF-2015-17-SC-AND-17449 respectively) and to the corresponding author (KSR) through Major Research Project (No: 43-586/2014 (SR), Dated 30-10-2015).

## **CONFLICT OF INTEREST**

The authors declare no conflict of interest.

## ABBREVIATIONS

HR-LC/Q-TOF/MS: High resolution liquid chromatography/ Quadruple time-of-flight/ mass spectrometer; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; ALP: Alkaline Phosphatase; CRE: Creatinine; UA: Uric Acid; TC: Total Cholesterol; TG: Triglyceride; T Bil: Total Bilirubin; D Bil: Direct Bilirubin; ALB: Albumin; GLB: Globulin; TP: Total protein; GLU: Glucose; RP- HPLC: Reverse phase high-performance liquid chromatography; MIC: Minimum inhibitory concentration.

## REFERENCES

- Helton LR. Folk medicine and health beliefs: an Appalachian perspective. J Cult Divers. 1996; 3: 123–8.
- 2. http://www.theplantlist.org
- 3. https://www.kew.org
- 4. http://shodhganga.inflibnet.ac.in
- Senanayake N, Sanmuganathan, PS. Acute intravascular haemolysis in glucose-6-phosphate dehydrogenase deficient patients following ingestion of herbal broth containing Acalypha indica. *Trop Doct* 1996; 26: 32.
- 6. https://www.prota4u.org
- 7. Chopra RN et al. Glossary of Indian medicinal plants. CSIR, New Delhi. 1956;
- Nandhakumar M et al. In Vitro Assay of Alpha Amylase Inhibitory Activity of Indian Medicinal Herb Acalypha Indica. J Clin Diagn Res 2009; 3: 1475-8.
- Stone BC. The flora of Guam. A Manual for the Identification of the Vascular Plants of the Island. *Micronesica* 1970; 6: 1–659.
- Krishnan N et al. Effect of seasonal and altitudinal variations on growth performance of Acalypha indica Linn. in Alagar Hill (Eastern Ghats), South India. Trop Ecol 2000; 41: 41-5.
- 11. http://powo.science.kew.org
- Mohan VR et al. Ethno-Medico-Botany of the Palliyars of Saduragiri Hills, Western Ghats, Tamilnadu. J Econ Taxon Bot 2010; 34: 639-57.
- Bagul RM. Status report on wild medicinal plants of Satpuda Forest East, Maharashtra India, Lulu publication. ISBN:1365414248, 9781365414244. 2016.
- Mallik B et al. Traditional Herbal Practices by the Ethnic People of Kalahandi District of Odisha, India. Asian Pacific Journal of Tropical Biomedicine. 2012; doi:10.1016/S2221-1691(12)60349-9.
- Jayaprakasam R, Ravi TK. Evaluation of Anti-Arthritic Activity of the Root Extract of Acalypha Indica Linn. Using In Vitro Techniques. Int J Phytopharm. 2012; 2: 169-73.
- Mohanty N et al. Herbal folk remedies of Dhenkanal district, Odisha, India. International Journal of Herbal Medicine 2015; 3: 24-33.
- Seebaluck R *et al.* Medicinal plants from the genus Acalypha (Euphorbiaceae)–a review of their ethnopharmacology and phytochemistry. *J Ethnopharmacol* 2015; 159: 137-57.
- Singh AG et al. An ethnobotanical survey of medicinal plants used in Terai forest of western Nepal. J Ethnobiol Ethnomed 2012; 8: 19.
- Sinhababu A, Banerjee A. Ethno-medicinal Plants of the Family Euphorbiaceae in Southern Bankura of West Bengal. *RRJoB* 2016; 5: 36-9

- Sivakumar R et al. Larvicidal and repellent activity of tetradecanoic acid against Aedesaegypti (Linn.) and Culexquinquefasciatus (Say.) (Diptera: Culicidae). Asian Pac J Trop Med 2011; 4: 706–10.
- Schmelzer GH et al. Plant Resources of Tropical Africa 11(1) Medicinal Plants 1, Backhuys Publishers, Wageningen, Netherlands. 2008. ISBN 978-90-5782-204-9.
- Senthilkumar M et al. Some medicinal plants used by Irular, the tribal people of Marudhamalai hills, Coimbatore, Tamil Nadu. Nat. prod. Radiance 2006a; 5: 382-8.
- Dharmadasa RM *et al.* Ethnopharmacological survey on medicinal plants used in snakebite treatments in Western and Sabaragamuwa provinces in Sri Lanka. *J Ethnopharmacol* 2016: 179:110-27.
- Hada BS, Katewa SS. Ethnomedicinal plants used against various diseases in Jhalawar district of Rajasthan, India. *Journal of Global Biosciences* 2015; 4: 2077-86.
- Neamsuvan O, Ruangrit T. Corrigendum: A survey of herbal weeds that are used to treat gastrointestinal disorders from southern Thailand: Krabi and Songkhla provinces. J Ethnopharmacol 2017; 209: 318-27.
- Silambarasan R, Ayyanar M. An ethnobotanical study of medicinal plants in Palamalai region of Eastern Ghats, India. J Ethnopharmacol 2015; 172: 162-78.
- Mahishi P et al. Medicinal plant wealth of local communities in some villages in Shimoga District of Karnataka, India. J Ethnopharmacol 2005; 98: 307-12.
- Ayyanar M, Ignacimuthu S. Ethnobotanical survey of medicinal plants commonly used by Kani tribals in Tirunelveli hills of Western Ghats, India. *J Ethnopharmacol* 2011; 134:851-64.
- Divya K et al. Ethno-Medicinal Plants used in East Godavari District, Andhra Pradesh, India. International Journal of Pharmacological Research 2015; 5: 293-300.
- Sivasankari B et al. An ethnobotanical study of indigenous knowledge on medicinal plants used by the village peoples of Thoppampatti, Dindigul district, Tamilnadu, India. J Ethnopharmacol 2014; 153: 408-23.
- Mutheeswaran S et al. Documentation and quantitative analysis of the local knowledge on medicinal plants among traditional Siddha healers in Virudhunagar district of Tamil Nadu, India. J Ethnopharmacol 2011; 137: 523–33.
- Muthu C *et al.* Medicinal plants used by traditional healers in Kancheepuram District of Tamil Nadu, India. *Journal of Ethnobiology and Ethnomedicine* 2006; 2: 43.
- Panda T et al. Folk Knowledge on Medicinal Plants Used for the Treatment of Skin Diseases in Bhadrak District of Odisha, India. Med Aromat Plants 2016; doi: 10.4172/2167-0412.1000262.
- Halim MA *et al.* The Use of Plants in Traditional Health Care Practice of the Shaiji Community in Southwestern Bangladesh. *Journal of Tropical Forest Science* 2007; 19: 168-75.
- Yabesh JEM et al. An ethnobotanical study of medicinal plants used by traditional healers in silent valley of Kerala, India. J Ethnopharmacology 2014; 154: 774-89.
- Shanmugam S et al. Traditional uses of medicinal plants among the rural people in Sivagangai district of Tamil Nadu, Southern India. Asian Pacific Journal of Tropical Biomedicine 2012; S429-34.
- Savithramma N et al. Ethnobotanical survey of plants used to treat asthma in Andhra Pradesh, India. J Ethnopharmacol 2007; 113: 54–61.
- Ribeiro A *et al.* Ethnobotanical survey in Canhane village, district of Massingir, Mozambique: medicinal plants and traditional knowledge. *J Ethnobiol Ethnomed* 2010; 6: 33.
- 39. http://www.stuartxchnge.org
- Divya N et al. Antibacterial Activity of Medicinal Plant against Wound Infected Pathogens. IJPSR 2014; 5: 4942-47.
- Lingaraju DP et al. Ethnopharmacological survey of traditional medicinal plants in tribal areas of Kodagu district, Karnataka, India. J Pharm Res 2013; 6: 284-97.
- 42. Shukla AN *et al.* An ethnobotanical study of medicinal plants of Rewa district, Madhya Pradesh. *Indian Journal of Traditional Knowledge* 2010; 9: 191-202.
- Mohan SC et al. Phytochemical, GC-MS analysis and Antibacterial activity of a Medicinal Plant Acalypha indica. Int J PharmTech Res 2012; 4: 1050-54.
- Sharma J *et al.* Ethnomedicinal plants used for treating epilepsy by indigenous communities of sub-Himalayan region of Uttarakhand, India. *J Ethnopharmacol* 2013; 150: 353–70.
- Ayyanar M, Ignacimuthu S. Herbal medicines for wound healing among tribal people in Southern India: ethnobotanical and scientific evidences. Int J Appl Res Nat Prod 2009; 2: 29–42.
- Pushpangadan P, Atal CK. Ethno-medico-botanical investigations in Kerala I. Some primitive tribals of western ghats and their herbal medicine. *J Ethno-pharmacol* 1984; 11: 59-77.
- Das PR *et al.* A Selection of Medicinal Plants Used for Treatment of Diarrhea by Folk Medicinal Practitioners of Bangladesh. *Am.-Eurasian J. Sustain. Agric* 2012; 6: 153-61.
- Silalahi M *et al.* The local knowledge of medicinal plants trader and diversity of medicinal plants in the Kabanjahe traditional market, North Sumatra, Indonesia. *J Ethnopharmacol* 2015; 175: 432-43.
- 49. Kumar SV et al. Mosquito Larvicidal, Oviposition deterrent and Repellent

properties of Acalypha indica L extracts against Aedes aegypti, Anopheles stephensi, and Culex quinquefasciatus. Int J Med Biosci 2012; 1: 33 – 41.

- Nadkarni AK. KM Nadkarni's Indian Materia Medica, Popular prakashana, Bombay, 3rd edition, 2002; 1: 811-16.
- Steyn DG. The Presence of Hydrocyanic Acid in Stock Feeds and Other Plants. Student American Veterinary Medical Association. 1938. 9: 60-4.
- Colley FC. Traditional Indian Medicine in Malaysia. Journal of the Malaysian Branch of the Royal Asiatic Society. 1978; 51: 77-109.
- Basha SK, Sudarshanam G. Multiple herbal therapy Antimicrobial activity of wound healing paste(Pasuru) used by Sugali tribes of Yerramalais of Kurnool district., Andhra Pradesh, India. Int J PharmTech Res 2011; 3: 1238-41.
- Panda SP et al. Potential Medicinal Plants of Odisha Used in Rheumatism and Conservation. American Journal of Ethnomedicine 2014; 1: 260-5.
- Adsul YD et al. Ethnobotanical Euphorbian plants of North Maharashtra Region. IOSR Journal of Pharmacy and Biological Sciences 2013; 7: 29-35.
- Boissya CL *et al.* Some Medicinal Plants from Darrang District of Assam, India. *Anthropos* 1981; 76: 220-2.
- 57. Basumatary SK *et al.* Some medicinal plant leaves used by Boro (tribal) people of Goalpara district, Assam. *Natural product radiance* 2004; 3: 88-90.
- Ghatapanadi SR *et al.* Documentation of folk knowledge on medicinal plants of Gulbarga district, Karnataka. Indian. *Journal of traditional knowledge* 2011; 10: 349-53.
- Kumar GP, Chaturvedi A. Ethnobotanical Observations of Euphorbiaceae Species from Vidarbha region, Maharashtra, India. *Ethnobotanical Leaflets* 2010; 14: 674-80.
- Patel RM et al. Some Noteworthy Ethno Medicinal Plants of Western Kachchh, Gujarat. Life sciences Leaflets 2010; 9: 244 – 50.
- Prakash R. Medicinal Plants Used By Tribal Communities: A Study of Uttarakhand Himalayan Region. *International Journal of Humanities and Social Science Invention* 2015; 4: 55-61.
- Hassan-Abdallah A et al. Medicinal plants and their uses by the people in the Region of Randa, Djibouti. J Ethnopharmacol 2013; 148: 701-13.
- Rudrapal M et al. Ethnomedicinal plants used by traditional healers in East Godavari district of Andhra Pradesh, India. Indian journal of natural products and resources 2012; 3: 426-31.
- Dagar HS. Plant Folk Medicines among Nicobarese Tribals of Car Nicobar Island, India. *Econ Bot* 1989; 43: 215-24.
- Kumar A et al. Traditional uses of medicinal plants for dermatological healthcare management practices by the Tharu tribal community of Uttar Pradesh, India. Genet Resour Crop Evol 2013c; 60:203–24.
- Subramanian A et al. Ethno-Medico Botany of The Valaiyans of adurai District, Western Ghats, Tamil Nadu. J Econ Taxon Bot 2010; 34: 363-79.
- Kumar RS *et al.* Ethno-botanical Uses of Some Plant Roots Used By "Gondu" Tribes of Seethagondi Grampanchayath, Adilabad District, Andhra Pradesh, India. Research and Reviews: *Journal of Botanical Sciences* 2013a; 2: 18-26.
- Kumari BS *et al.* Traditional knowledge of medicinal plants used to cure respiratory diseases in Krishna District of Andhra Pradesh, India. *Journal of Medicinal Plants Studies* 2014; 2: 34-7
- Ijjirouthu BR et al. An ethno-medicinal survey of medicinal plants used by traditional healers of Araku valley, Andhra Pradesh, India. International Journal of Current Research 2017; 9: 48633-45.
- Singh RS, Shahi SK. Diversity of medicinal plants of Ratanpur region of Bilaspur district (Chhattisgarh). Journal of Medicinal Plants Studies 2017; 5: 276-81.
- Singh A et al. Traditional Medicinal Flora of the District Buxar (Bihar, India). Journal of Pharmacognosy and Phytochemistry 2013; 2: 41-9.
- Kusum EK et al. Traditional Use of Medicinal Plants Practiced by Theoraon Tribe of Jashpur District (C.G.). India. Journal of Environmental Science, Toxicology and Food Technology 2012; 1: 60-4.
- Wadankar GD *et al.* Traditionally Used Medicinal Plants for Wound Healing in the Washim District, Maharashtra (India). *International Journal of PharmTech Research* 2011; 3: 2080-4.
- Poornima R, Prabakaran G. Preliminary phytochemical screening and antibacterial activity of Acalypha indica and euphorbia hirta of family euphorbiaceae against some pathogenic organisms. *Int J Agr Sci* 2012; 2: 34-8.
- Hussain AZ, Kumaresan S. GC-MS analysis and antibacterial evaluation of Acalypha indica. Asian J Plant Sci Res 2013; 3: 46-9.
- Pragada RR *et al.* Phytochemical Investigation and In Vitro Anti- Oxidant, Anti- Microbial Activity of Different Fractions of Acalypha Indica Linn. *Int J Pharm Pharm Sci* 2011; 3: 314-17.
- Gopalakrishnan V et al. Antimicrobial activity of extracts of Acalypha indica linn. Indian J Pharm Sci 2000; 62: 347-50.
- Ravi S et al. Identification of food preservative, stress relief compounds by GC–MS and HR-LC/Q-TOF/MS; evaluation of antioxidant activity of Acalypha indica leaves methanolic extract (in vitro) and polyphenolic fraction (*in vivo*). *J Food SciTechnol* 2017; 54: 1585-96.
- Balakrishnan N et al. The Evaluation of Nitric Oxide Scavenging Activity of Acalypha Indica Linn Root. Asian J Research Chem 2009; 2: 148-50.

- Itankar PR et al. Antidiabetic Potential and its Relationship with Polyphenol and Degree of Polymerization in Acalypha indica Linn Leaves. J Homeop Ayurv Med 2011; 1: 102.
- Kumar SVS et al. Hepatoprotective Activity of Aclypha Indica Linn against Thioacetamide Induced Toxicity. Int J Pharm Pharm Sci 2013b; 5: 356-59.
- Godipurge SS et al. Phytochemical and Pharmacological Evaluation of Acalypha indica Linn in Experimental Animal Models. International Journal of Pharmacognosy and Phytochemical Research 2015; 6: 973 – 79.
- Selvamani S. Evaluation of Antimicrobial and Antioxidant Potential of Acalypha indica Linn. Thesis Submitted to the Annamalai University, Tamilnadu, India. 2015.
- 84. Murugan. Cardioprotective Effect of Acalypha Indica against Myocardial Infarction and Left Ventricular Proteomic Analysis of Furosemide and Potassium Chloride Treated Rats. Thesis submitted to Madurai Kamaraj University, 2013.
- Suri R *et al.* Preliminary studies on the analysis of fatty acids, essential oils and flavonoids in Acalypha indica L. *J Trop Agric Fd Sc* 2004; 32: 163-9.
- Inagaki H et al. Identification of a pheromone that increases anxiety in rats. Proc Natl Acad Sci U S A 2014; 11: 18751–6.
- Hungeling M et al. Cynogenic and non-cyanogenic pyridone glucosides from Acalypha indica (Euphorbiaceae). Phytochemistry 2009; 70: 270-7.
- Schmitt H. Influence of Reserpine and Rescinnamine on the Spontaneous and Evoked Sympathetic Activity in Rats. *Pharmacology* 1968; 1:25–32.
- Wong DT *et al.* Dopamine receptor affinities in vitro and neurochemical effects *in vivo* of pergolide and its metabolites. *Arzneimittelforschung* 1993; 43: 409-12.
- Clemens JA *et al.* Dopamine agonist activities of pergolide, its metabolites, and bromocriptine as measured by prolactin inhibition, compulsive turning, and stereotypic behavior. *Arzneimittelforschung* 1993; 43(3):281-86.
- Usmanova DM. Experimental effect of a lupinine-based homopolymer on blood coagulation in animals. *Farmakol Toksikol* 1980; 43: 327-30.
- Schmeller T et al. Binding of quinolizidine alkaloids to nicotinic and muscarinic acetylcholine receptors. J Nat Prod 1994; 57: 1316-19.
- Nahrstedt A et al. Acalyphin, a cyanogenic glucoside from acalypha indica. *Phyrochemistry* 1982; 21: 101-5.
- Chaichoowong S *et al.* Chemical Profiling of Acalypha indica Obtained from Supercritical Carbon Dioxide Extraction and Soxhlet Extraction Methods. *Orient J Chem* 2017; 33: 66-73.
- Teklani PWNN, Perera BGK. Mosquito Repellent and Larvicidal Activities of Acalypha Indica Leaf Extracts. Int J Pharm Pharmacol 2017; 1: 107.
- Kokot Z. Studies of neutralizing properties of antacid preparations. Part 5: Dissolution kinetics of dihydroxyaluminum aminoacetate. *Pharmazie* 1989; 44: 274-5.
- Houghton DC *et al.* Amikacin nephrotoxicity in the rat. J. Environ. Pathol Toxicol 1980; 4: 277-91.
- Rahman MA *et al.* Analgesic and Antiinflammatory Activity of Methanolic Extract of Acalypha Indica Linn. *Pak J Pharm Sci* 2010; 23; 256-8.
- Sanseera D et al. Antioxidant and Anticancer Activities from Aerial Parts of Acalypha indica Linn. CMU J Nat Sci 2012; 11: 157-68.
- Raj J, Singh KP. Acalypha indica. In: Central Council for Research in Homeopathy 2000; 22: 1-6.
- Murota H, Katayama I. Emedastine difumarate: a review of its potential ameliorating effect for tissue remodeling in allergic diseases. Expert. Opin Pharmacother 2009; 10: 1859-67.
- 102. Subasri S et al. Phytochemical analysis, molecular docking and molecular dynamics simulations of selected phytoconstituents from four herbs as antidotes for snake bites. *Clinical Proteomics and Bioinformatics* 2016; 1: 1-13.
- Rajashekar Y et al. Acetylcholinesterase Inhibition by Biofumigant (Coumaran) from Leaves of Lantana camara in Stored Grain and Household Insect Pests. *BioMed Research International* 2014; doi.org/10.1155/2014/187019.
- Bhatia ML et al. HaemodynamicStudies with Peruvoside in Human Congestive Heart Failure. British medical journal 1970; 3: 740-3.
- Liu JR et al. Effects of beta-ionone on mammary carcinogenesis and antioxidant status in rats treated with DMBA. Nutr Cancer 2010; 62: 58-65.
- Sgarbossa A *et al.* Ferulic Acid: A Hope for Alzheimer's Disease Therapy from Plants. *Nutrients* 2015; 7: 5764-82.
- Hoult JR, Payá M. Pharmacological and biochemical actions of simple coumarins: Natural products with therapeutic potential. *Gen Pharmacol* 1996; 27: 713-22.
- Riveiro ME et al. Biochemical mechanisms underlying the pro-apoptotic activity of 7,8-dihydroxy-4-methylcoumarin in human leukemic cells. Biochem Pharmacol, 2008; 75: 725-36.
- Goel A et al. 7,8-Dihydroxy-4-methylcoumarin induces apoptosis of human lung adenocarcinoma cells by ROS-independent mitochondrial pathway through partial inhibition of ERK/MAPK signaling. FEBS Lett 2007; 581: 2447-54.
- Zhang W et al. Dicumarol inhibits PDK1 and targets multiple malignant behaviors of ovarian cancer cells. PLoS ONE 2017; 12: e0179672. doi: 10.1371/ journal.pone.0179672.
- 111. Talapatra B et al. Acalyphamide, a new amide and other chemical constituents

of Acalypha indica Linn. Ind J Chem Sect B 1981; 20: 974-7.

- Maehashi K et al. Bitter peptides activate hTAS2Rs, the human bitter receptors. Biochem Biophys Res Commun 2008; 365: 851–5.
- Jablonska-Trypuc A *et al.* Traumatic Acid Reduces Oxidative Stress and Enhances Collagen Biosynthesis in Cultured Human Skin Fibroblasts. *Lipids* 2016; 51: 1021-35.
- Van Overbeek J. Traumatic Acid and Thiamin as Growth Factors for Algae. Proc Natl Acad Sci U S A 1940; 26: 441-3.
- Sato T et al. Regulatory effects of the Llysine metabolites, L2-aminoadipic acid and Lpipecolic acid, on protein turnover in C2C12 myotubes. *Biosci Biotechnol Biochem* 2016; 18:1-8.
- Mickymaray S et al. Screening and antibacterial efficacy of selected Indian medicinal plants. Asian Pac J Trop Biomed 2016; 6: 185-91.
- Dellschaft NS et al. The dietary form of choline during lactation affects maternal immune function in rats. Eur J Nutr 2017; doi: 10.1007/s00394-017-1493-0.
- Ceci F et al. Therapy assessment in prostate cancer using choline and PSMA PET/CT. Eur J Nucl Med Mol Imaging 2017; doi: 10.1007/s00259-017-3723-3.
- Hussain AZ, Ignatius A. GC-MS Analysis and Antimicrobial Activity of Acalypha indica Linn. Asian J Chem 2010; 22: 3591-5.
- Or-Rashid MM *et al.* Fatty acid composition of yak (Bos grunniens) cheese including conjugated linoleic acid and trans-18:1 fatty acids. *J Agric Food Chem* 2008; 56: 1654–60.
- Meng Q et al. Myristic acid-conjugated polyethylenimine for brain-targeting delivery: In vivo and ex vivo imaging evaluation. J Drug Target 2010; 18: 438–46.
- 122. Kim YI *et al.* Potent PPARα activator derived from tomato juice, 13-oxo-9, 11-octadecadienoic acid, decreases plasma and hepatic triglyceride in obese diabetic mice. *PLoS One* 2012; 7: e31317. doi: 10.1371/journal.pone.0031317.
- Armstrong MM *et al.* Inhibitory and Mechanistic Investigations of Oxo-Lipids with Human Lipoxygenase Isozymes. *Bioorg Med Chem* 2014; 22: 4293–7.
- Kuehl F A et al. The Identification of N-(2-Hydroxyethyl)-Palmitamide as a Naturally Occurring Anti-Inflammatory Agent. J Am Chem Soc 1957; 79: 5577–8.
- Svec P et al. The effect of long-term administration of N-(2-hydroxyethyl)palmitamide on the chemotherapy of RBA rat leukemia. Neoplasma 1975; 22: 625-30.
- 126. Rani CK. Evaluation of antihyperglycemic, antihyperlipidemic and antioxidant potential of ethanolic extract of aerial parts of Acalypha indica Linn in Streptozotocin induced diabetic rats. Thesis submitted to the Mother Teresa Women's University, Kodaikanal, India 2014.
- 127. Xiao YH et al. Male-specific (Z)-9-tricosene stimulates female mating behaviour in the spider Pholcus beijingensis. *Proc Biol Sci* 2010; 277: 3009-18.
- Deng S *et al.* Effect of triacontanol on the pharmacokinetics of docetaxel in rats associated with induction of cytochrome P450 3A1/2. Xenobiotica 2014; 44: 583-90.
- Jiang Z et al. Catechin attenuates traumatic brain injury-induced impairments of blood-brain barrier damage and improve longer-term neurological outcomes in rats. Exp Physiol 2017; doi: 10.1113/EP086520.
- Bimonte S *et al.* An overview of pre-clinical studies on the effects of (-)-epigallocatechin-3-gallate, a catechin found in green tea, in treatment of pancreatic cancer. *Recenti Prog Med* 2017; 108: 282-7. doi: 10.1701/2715.27715.
- Santos LFS *et al.* Catechin and epicatechin reduce mitochondrial dysfunction and oxidative stress induced by amiodarone in human lung fibroblasts. J Arrhythm 2017; 33: 220-5.
- Meena KP et al. Catechin-loaded Eudragit microparticles for the management of diabetes: formulation, characterization and *in vivo* evaluation of antidiabetic efficacy. J Microencapsul 2017; 34: 342-50.
- Sae-Leaw T et al. Effect of catechin and its derivatives on inhibition of polyphenoloxidase and melanosis of Pacific white shrimp. J Food Sci Technol 2017; 54: 1098-07.
- Lee H et al. Therapeutic Efficacy of Nanocomplex of Poly(Ethylene Glycol) and Catechin for Dry Eye Disease in a Mouse Model. Invest Ophthalmol Vis Sci 2017; 58: 1682-91.
- Yoneshiro T *et al.* Tea catechin and caffeine activate brown adipose tissue and increase cold-induced thermogenic capacity in humans. *Am J Clin Nutr* 2017; 105: 873-81.
- Poncet-Montange G et al. A survey of antiprion compounds reveals the prevalence of non-PrP molecular targets. J Biol Chem 2011; 286: 27718-28.
- 137. MaY et al. Phenolics from Acalypha indica. J Chin Chem Soc 1997; 44: 499-02.
- 138. Yang J, Liu RH. Synergistic Effect of Apple Extracts and Quercetin 3-β-D-Glucoside Combination on Antiproliferative Activity in MCF-7 Human Breast Cancer Cells in Vitro. J Agric Food Chem 2009; 57: 8581–6.
- Rajamanickam M et al. Antibacterial and Wound Healing Activities of Quercetin-3-O-A-L-Rhamnopyranosyl- (1-6)-β-D-Glucopyranoside Isolated from Salvia leucantha. Int J Pharm Sci Rev Res 2013; 22: 264-8.
- Liu M, Chen Z. Protective effect of quercetin- 3'-O-β-glucoside on myocardial ischemic injury in mice. Anhui Yike Daxue Xuebao 2008; 43: 683 – 5.
- Alluis B, Dangles O. Quercetin (= 2-(3,4- dihydroxyphenyl)-3,5,7-trihydroxy-4H-1- benzopyran-4- one) glycosides and sulfates: chemical synthesis, complexation, and antioxidant properties. *Helv Chim Acta* 2001; 84: 1133 – 56.

- Arung ET *et al.* Melanin Biosynthesis Inhibitory and Antioxidant Activities of Quercetin-3'-O-β-D-glucoside Isolated from Allium cepa. *Z Naturforsch* 2011; 66: 209 – 14.
- 143. Yamazaki S *et al.* Quercetin-3-O-glucuronide inhibits noradrenaline-promoted invasion of MDA-MB-231 human breast cancer cells by blocking β2-adrenergic signaling. *Arch Biochem Biophys* 2014; 557: 18–27.
- 144. Yang HH *et al.* Quercetin-3-O-β-D-glucuronide isolated from Polygonum aviculare inhibits cellular senescence in human primary cells. *Arch Pharm Res* 2014; 37:1219-33.
- 145. Park JY et al. Quercetin-3-O-β-D-GlucuronideSuppresses Lipopolysaccharide-Induced JNK and ERK Phosphorylation in LPS-Challenged RAW264.7. Cells Biomol Ther (Seoul) 2016; 24: 610–15.
- Ganeshpurkar A, Saluja AK. The Pharmacological Potential of Rutin (Review). Saudi Pharm J 2017; 25: 149–64.
- 147. Nahrstedt A et al. Flavonoids from Acalypha indica. Fitoterapia 2006; 77: 484-486.
- Vellosa JCR et al. Antioxidant and cytotoxic studies for kaempferol, quercetin and isoquercitrin. Eclet Quím 2011; 36: 7-20.
- Chen AY, Chen YC. A review of the dietary flavonoid, kaempferol on human health and cancer chemoprevention. *Food Chem* 2013; 138: 2099–07.
- Calderón-Montaño JM et al. Review on the Dietary Flavonoid Kaempferol. Mini Rev Med Chem 2011; 11: 298-34.
- 151. Yu L *et al.* Neuroprotective Effect of Kaempferol Glycosides against Brain Injury and Neuroinflammation by Inhibiting the Activation of NFκB and STAT3 in Transient Focal Stroke. *PLoS One* 2013; 8: -e55839. doi: 10.1371/journal. pone.0055839.
- Patel K et al. A Review on Pharmacological and Analytical Aspects of Naringenin. Chin J Integr Med 2014; Doi: 10.1007/s11655-014-1960-x.
- 153. Sarkar A et al. β-D-Glucoside Suppresses Tumor Necrosis Factor-induced Activation of Nuclear Transcription Factor κB but Potentiates Apoptosis. J Biol Chem 2004; 279: 33768-33781.
- Wick MM et al. Reduction of streptozotocin toxicity by 3-O-methyl-D-glucose with enhancement of antitumor activity in murine L1210 leukemia. *Cancer Res* 1977; 37: 3901-03.
- Rivlin M *et al.* Functional molecular imaging of tumors by chemical exchange saturation transfer MRI of 3-O-Methyl-D-glucose. *Magn Reson Med* 2014; 72: 1375-80.
- Shamni O et al. Regulation of GLUT4 activity in myotubes by 3-O-methyl-dglucose. Biochim Biophys Acta 2017; doi: 10.1016/j.bbamem.2017.06.013.
- 157. FAO/WHO Report on Glyceryl Monopalmitate: Food Additives: Emulsifier. FAO/WHO Food Additive Evaluations – JECFA, 1973.
- Perlík F et al. The effect of N-(2-hydroxyethyl)-palmitamide on delayed hypersensitivity in guinea-pig. Experientia 1973; 29: 67-8.
- Badhani B et al. REVIEW :Gallic acid: a versatile antioxidant with promising therapeutic and industrial applications. RSC Adv 2015; 5: 27540- 57.
- Adefegha SA *et al.* Antioxidant and antidiabetic effects of gallic and protocatechuic acids: a structure–function perspective. *Comp Clin Pathol* 2015; 24: 1579. doi.org/10.1007/s00580-015-2119-7.
- Patel SS, Goyal RK. Cardioprotective effects of gallic acid in diabetes-induced myocardial dysfunction in rats. *Pharmacognosy Res* 2011; 3: 239–45.
- Hajipour S et al. Effect of Gallic Acid on Dementia Type of Alzheimer Disease in Rats: Electrophysiological and Histological Studies. *Basic Clin Neurosci* 2016; 7: 97–06.
- Wei X et al. Syringic Acid Extracted from Herba dendrobii Prevents Diabetic Cataract Pathogenesis by Inhibiting Aldose Reductase Activity. Evid Based Complement Alternat Med 2012; doi:10.1155/2012/426537.
- Cikman O *et al.* Antioxidant Activity of Syringic Acid Prevents Oxidative Stress in L-arginine-Induced Acute Pancreatitis: An Experimental Study on Rats. *Int Surg* 2015; 100: 891-6.
- Touaibia M et al. Caffeic Acid, A Versatile Pharmacophore: An Overview. Mini Rev Med Chem 2011; 11: 695-13.
- Magnani C et al. Caffeic acid: a review of its potential use in medications and cosmetics. Anal Methods 2014; 6: 3203-10.
- Cooke CE, Mehra IV. Oral ondansetron for preventing nausea and vomiting. Am J Hosp Pharm 1994; 51: 762-71.
- Schnadower D et al. Ondansetron and probiotics in the management of pediatric acute gastroenteritis in developed countries. *Curr Opin Gastroenterol* 2015; 31: 1-6.
- 169. Solomon RDJ et al. Isolation, identification and study of antimicrobial property of a bioactive compound in an Indian medicinal plant Acalypha indica (Indiannettle). World J Microbiol Biotechnol 2005; 21: 1231–6.
- Borhade V et al. Clotrimazole nanoemulsion for malaria chemotherapy. Part II: Stability assessment, *in vivo* pharmacodynamic evaluations and toxicological studies. *Int J Pharm* 2012; 431: 149–60.
- Crowley PD, Gallagher HC. Clotrimazole as a pharmaceutical: past, present and future. J Appl Microbiol 2013; 117: 611-17.
- Liggett JL et al. Anti-tumor activity of non-steroidal anti-inflammatory drugs: Cyclooxygenase-independent targets. Cancer Lett 2014; 346: 217–24.

- 173. Steiner JP *et al.* Role of limonoid compounds as neuroprotective agents. US 20100056617 A1, 1996.
- Denis D et al. Synthesis and biological activities of leukotriene F4 and leukotriene F4 sulfone. Prostaglandins 1982; 24: 801-14.
- 175. Kawaoka Y et al. Screen for inhibitors of filovirus and uses therefor. Patent No: US 20170097334 A1, 2017.
- Lin JC et al. N,N-dimethylglycine differentially modulates psychotomimetic and antidepressant-like effects of ketamine in mice. Prog Neuropsychopharmacol Biol Psychiatry 2016; 71: 7-13.
- 177. Kern JK et al. Effectiveness of N,N-dimethylglycine in autism and pervasive developmental disorder. J Child Neurol 2001; 16: 169-73.
- Pfeffer G et al. Treatment for mitochondrial disorders. Cochrane Database Syst Rev 2012; doi: 10.1002/14651858.CD004426.pub3.
- 179. Carlezon WA *et al.* Antidepressant-like effects of cytidine in the forced swim test in rats. Biol Psychiatry 2002; 51: 882-89.
- Nakatsugawa S et al. PLDR inhibitors: their biological and clinical implications. Br J Cancer 1984; 6: 43–7.
- Selvamani S, Balamurugan S. Phytochemical Screening and GC-MS Analysis of Acetone Leaf Extract of Acalypha indica (Linn.). *International Journal of Research Studies in Biosciences (IJRSB)* 2015; 3: 229-32.
- Bushra, R, Aslam N. An Overview of Clinical Pharmacology of Ibuprofen. Oman Med J 2010; 25: 155–61.
- Kaplowitz N, DeLeve LD. Book: Drug-Induced Liver Disease. Academic Press 2013; 59-60.
- Ball GL et al. Toxicological review and oral risk assessment of terephthalic acid (TPA) and its esters: A category approach. Crit Rev Toxicol 2012; 42: 28–67.
- Gorchein A et al. Harderoporphyrin: a misnomer. Biomed Chromatogr 2005; 19: 565-69.
- 186. Armstrong JM *et al.* A comparison of the vasodepressor effects of the cyclic endoperoxides PGG2 and PGH2 with those of PGD2 and PGE2 in hypertensive and normotensive rats. *Eur J Pharmacol* 1976; 39: 251-8
- Malmsten C *et al.* Physiological role of an endoperoxide in human platelets: hemostatic defect due to platelet cyclo-oxygenase deficiency. *Proc Natl Acad Sci U S A* 1975; 72: 1446-50
- Bersani FS *et al.* Injecting eye-drops: a mini-review on the non-clinical use of tropicamide. *Hum Psychopharmacol Clin Exp* 2015; 30: 262 – 264.
- 189. Terzi HA *et al.* The antibacterial effects of bilirubin on gram-negative bacterial agents of sepsis. *Biomed Res* 2016; 27: 207-09.
- Loizou S et al. Beta-sitosterol exhibits anti-inflammatory activity in human aortic endothelial cells. Mol Nutr Food Res 2010; 54: 551-8.
- 191. Yinusa I *et al.* Evaluation of the Pharmacological Activities of Betasitosterol Isolated From The Bark of Sarcocephalus Latifolius. Global Journal of Pure and Applied Chemistry Research 2015; 3: 7-14.
- 192. Kaur N et al. Stigmasterol: A Comprehensive Review. Ijpsr 2011; 2: 2259-65.
- Jang SE *et al.* Penta-O-galloyl-β-D-glucose ameliorates inflammation by inhibiting MyD88/NF-κB and MyD88/MAPK signalling pathways. Br J Pharmacol 2013; 170: 1078-91.
- 194. Puppala M *et al.* The Isolation and Characterization of β-Glucogallin as a Novel Aldose Reductase Inhibitor from Emblica officinalis. *Plos one* 2012; 7: e31399.
- Lee, JH et al. 1, 2, 3, 4, 6-penta-O-galloyl-beta-D-glucose attenuates renal cell migration, hyaluronan expression, and crystal adhesion. *Eur J Pharmacol* 2009; 606: 32–7.
- Ono K et al. Pentagalloylglucose, an antisecretory component of Paeoniae radix, inhibits gastric H+, K(+)-ATPase. Clin Chim Acta 2000; 290: 159–67.
- Ahn MJ et al. Inhibition of HIV-1 integrase by galloyl glucoses from Terminalia chebula and flavonol glycoside gallates from Euphorbia pekinensis. *Planta Med* 2002; 68: 457–9.
- Jin F et al. Anti-inflammatory and anti-oxidative effects of corilagin in a rat model of acute cholestasis. BMC Gastroenterol 2013; 13:79.
- Moreira J et al. Anti-hyperalgesic activity of corilagin, tannin isolated from Phyllanthus niruri L. (Euphorbiaceae). J Ethanopharmacol 2013; 146: 318–23.
- Xiao HT et al. Inhibitory Effect of the Gallotannin Corilagin on Dextran Sulfate Sodium-Induced Murine Ulcerative Colitis. J Nat Prod 2013; 76: 2120–5.
- Kinoshita S *et al.* Antioxidant and hepatoprotective actions of medicinal herb, Terminalia catappa L. from Okinawa Island and its tannin corilagin. *Phytomedicine* 2007; 14(11):755–62.
- Ming Y et al. Corilagin inhibits hepatocellular carcinoma cell proliferation by inducing G2/M phase arrest. Cell Biol Int 2013; 37: 1046–54.
- Ha, DK et al. In vivo anti-tumour activity of corilagin on HEP3B hepatocellular carcinoma. Phytomedicine 2010; 18:11–5.
- 204. Ho RB. Antioxidative Role of Geraniin in Lipid Peroxidation of Human LDL. Journal of life sciences 2004; 4: 180-7.
- Ren Z et al. Geraniin suppresses tumor cell growth and triggers apoptosis in human glioma via inhibition of STAT3 signaling. Cytotechnology 2017; 69(5):765-73
- 206. Boakye YD et al. Anti-infective Properties and Time-Kill Kinetics of Phyllanthus

muellerianus and its Major Constituent, Geraniin. *Med chem (Los Angeles)* 2016; 6: 095-04.

- 207. Elendran S et al. The physicochemical properties of geraniin, a potential antihyperglycemic agent. Pharm Biol 2015; 53: 1719–26.
- Ko H. Geraniin inhibits TGF-β1-induced epithelial-mesenchymal transition and suppresses A549 lung cancer migration, invasion and anoikis resistance. *Bioorg Med Chem Lett* 2015; 25: 3529-34.
- Malini P et al. Antibiabetic Efficacy of Ellagic Acid in Streptozotocin
   induced Diabetes Mellitus in Albino Wistar Rats. Asian J Pharm Clin Res 2011; 4: 124-8.
- Ndukwe GI, Zhao Y. Pharmacological activity of 2,3,8-tri-O-methyl ellagic acid isolated from the stem bark of Irvingia gabonensis. *Afr J Biotechnol* 2007; 6: 1910-2.
- Urban N et al. Identification and validation of larixyl acetate as a potent TRPC6 inhibitor. Mol Pharmacol 2016; 89: 197-213.
- de Moraes J *et al.* Phytol, a Diterpene Alcohol from Chlorophyll, as a Drug against Neglected Tropical Disease Schistosomiasis Mansoni. *PLoS Negl Trop Dis* 2014; 8: e2617.
- Costa JP et al. Anxiolytic-like effects of phytol: possible involvement of GABAergic transmission. Brain Res 2014; 1547: 34-42.
- Senthilkumar S et al. Effect of squalene on cyclophosphamide-induced toxicity. Clin Chim Acta 2006b; 364: 335-42.
- Bachmann GE et al. Male sexual enhancement after methoprene treatment in Anastrepha fraterculus (Diptera: Tephritidae): A sustained response that does not fade away after sexual maturation. J Insect Physiol 2017; 101: 7-14.
- Kupchan SM *et al.* The Isolation and Structural Elucidation of Two Novel Sesquiterpenoid Tumor Inhibitors from Elephantopus elatus1,2. *J Am Chem Soc* 1966; 188 (15): 3674–6.
- 217. Shmuel Y. Book: Dictionary of food compounds with CD-ROM: Additives, flavors, and ingredients. Boca Raton: Chapman & Hall/CRC. 2004.
- 218. Roldi LP *et al.* Vitamin E (alpha-tocopherol) supplementation in diabetic rats: effects on the proximal colon. *BMC Gastroenterol* 2009; 9: 88.
- 219. Smith RE *et al.* The evolution of proteinase substrates with special reference to dipeptidylpeptidase IV. *Histochem J* 1992; 24: 637-47.
- Berkov S et al. Antiproliferative Alkaloids from Crinum zeylanicum. Phytother Res 2011; 25: 1686 – 92.
- 221. Begum DT *et al.* Evaluating phytochemical and antidiabetic activity of ethanolic extract of whole Plant of Acalypha indica Linn on alloxan induced diabetic Rats. *J Pharm Res* 2011; 4: 1704-6.
- Sathya M et al. Acute and Sub-acute Toxicity Studies of Ethanolic Extract of Acalypha Indica Linn in Male Wistar Albino Rats. Asian J Pharm Clin Res 2012; 5, 97 -100.
- Shirwaikar A *et al.* Neutralization potential of Viper russelli russelli (Russell's viper) venom by ethanol leaf extract of Acalypha indica. J Ethnopharmacol 2004; 94: 267–73.
- Badami S, Channabasavaraj KP. In Vitro. Antioxidant Activity of Thirteen Medicinal Plants of India's Western Ghats. *Pharm Biol* 2007; 45: 392–6.
- 225. Shanmugapriya R et al. Evaluation of Antioxidant Potential and Antibacterial Activity of Acalypha indica Linn. using in vitro model. Asian Journal of Biomedical and Pharmaceutical Sciences 2011; 1: 18-22.
- 226. Ravi S et al. Assessment of Potential Antioxidant Activity of Polyphenolic Fraction Separated from Acalypha Indica Leaves: An In vitro Approach. International Journal of Pharma Research & Review 2015; 4: 77-82.
- 227. Reddy TRP *et al.* Exploring the Anti-inflammatory and Anti-cancer compounds from the leaves of Acalypha indica. *IOSR J Pharm Biol Sci* 2012; 4: 01-07.
- Amarnath K et al. Cytotoxicity induction by ethanolic extract of Acalypha indica loaded casein-chitosan microparticles in human prostate cancer cell line in vitro. Biomedicine & Preventive Nutrition 2013; 4: 445-50
- Krishnaraj C *et al.* Acalypha indica Linn: Biogenic synthesis of silver and gold nanoparticles and their cytotoxic effects against MDA-MB-231, human breast cancer cells. *Biotechnology Reports* 2014; 4: 42–9.
- Sivaraj R et al. Biosynthesis and characterization of Acalypha indica mediated copper oxide nanoparticles and evaluation of its antimicrobial and anticancer activity. Spectrochim. Acta A Mol Biomol Spectrosc 2014; 129: 255-8.
- 231. Reddy JS et al. Wound healing effects of Heliotropium indicum, Plumbago zeylanicum and Acalypha indica in rats. J Ethnopharmacol 2002; 79: 249-51.
- Kumarasamyraja D, Swamielamanickam M. Evaluation of invivo and invitro wound healing activity of aqueous extract of Acalypha indica. *Int Res J Pharm* 2015; 6: 57-61.
- Ganeshkumar M et al. Topical application of Acalypha indica accelerates rat cutaneous wound healing by up-regulating the expression of Type I and III collagen. J Ethnopharmacol 2012; 142: 14–22.
- Hiremath SP et al. Post-coital antifertility activity of Acalypha indica L. J Ethnopharmacol 1999; 67: 253–8.
- Rajendran K et al. In vitro and in vivo anti-snake venom (Daboia russelli) studies on various leaf extracts of Acalypha indica Linn. International Journal of Phytomedicine 2010; 2: 217-20.
- 236. Mandal N, Khora SS. Ameliorative action of aqueous extract of Acalypha indica

against puffer fish Lagocephalus lunaris induced toxicity. International Journal of Drug Development & Research 2013; 5: 257-71.

- 237. Fatima N et al. Cloning, expression, purification of active recombinant human 12R-LOX enzyme and its inhibition by Acalypha indica extracts. *PeerJ Preprints* 2017; 5:e2906v1
- Sathya M et al. Biopotency of Acalypha indica Linn on Membrane Bound ATPases and Marker Enzymes urolithic Rats. Anc Sci Life 2011; 31: 3–9.
- Mouli KC et al. Effectiveness of flavonoid-rich leaf extract of Acalypha indica in reversing experimental myocardial ischemia: biochemical and histopathological evidence. Zhong Xi Yi Jie He Xue Bao 2012a; 10: 784-92.
- Mouli KC. Protective effects of flavonoids from Acalypha indica on isoproterenol induced myocardial ischemia in rats. Theses submitted to Sri Venkateswara University, Tirupati, India. 2012b.
- 241. Masih M *et al.* Antidiabetic Activity of Acalypha Indica Linn. on Normal and Alloxan Induced Diabetic Rats. *Int J Pharm Pharm Sci* 2011; 3: 514.
- 242. Saha R, Ahmed A. Hypoglycemic Effect of Acalypha Indica Linn. Plant Extracts on Streptozotocin Induced Diabetes in Rat. *IJPSR* 2011; 2: 2934-7.
- Priya CL, Rao KVB. Postprandial Antihyperglycemic And Antioxidant Activities of Acalypha indica Linn Stem Extract: An In-vivo Study. *Pharmacogn. Mag* 2016; 12 (Suppl 4): S475–81.
- 244. Chengaiah B et al. In-Vitro Anthelmintic Activity of Roots of Acalypha Indica Linn. International Journal of PharmTech Research 2009; 1: 1499-02.
- George M et al. Anti-Arthritic Activity of Plant Acalypha Indica Extract. AJPP 2016; 3: 007-15.
- Mathew M et al. Preventive and curative effects of Acalypha indica on acetaminophen-induced hepatotoxicity. International Journal of Green Pharmacy 2011; 5: 49-54.
- 247. Armansyah TRT *et al.* Hepatoprotective activity of ethanol extract of cat-and-seek Leaf Acalypha indica L.) in the White Rat (Rattus Novergicus) which induced Paracetamol. *Jurnal Ilmiah Ilmu-Ilmu Peternakan Mei* 2010; XIII: 292-8.
- 248. Vijayabhaskar K *et al.* Enhance the Effect of Piperine on Heptoprotective Activity of Acalypha indica to Combat Oxidative Stress. *IJAPBC* 2013; 2: 32-6.
- Dwijayanti A *et al.* Hepatoprotective Effects of Acalypha indica and Centella asiatica in Rat's Liver Against Hypoxia. *Procedia Chemistry* 2015; 14: 11–4.
- Manjulatha K. Comparitive Study of Leaves and Roots Ethanolic Extracts of Acalypha Indica on Peptic Ulcers Induced by Physical and Chemical Agents in Rodents. VRI Phytomedicine 2013; 1: 19-25.
- 251. Olowu RA et al. Concentration of Heavy Metals in Root, Stem and Leaves of Acalypha indica and Panicum maximum jacq from Three Major Dumpsites in Ibadan Metropolis, South West Nigeria. American Journal of Chemistry 2015; 5(1): 40-8.
- Venkatachalam P *et al.* Accumulation efficiency, genotoxicity and antioxidant defense mechanisms in medicinal plant Acalypha indica L. under lead stress. *Chemosphere* 2017; 171: 544–53.
- Scaffidi A *et al.* Identification of the Cat Attractants Isodihydronepetalactone and Isoiridomyrmecin from Acalypha indica. *Australian Journal of Chemistry* 2016; 69: 169-73.
- Krishnaraj C et al. Synthesis of silver nanoparticles using Acalypha indica leaf extracts and its antibacterial activity against water borne pathogens. *Colloids Surf B Biointerfaces* 2010; 76: 50–6.
- Krishnaraj C et al. Optimization for rapid synthesis of silver nanoparticles and its effect on phytopathogenic fungi. Spectrochim Acta A Mol Biomol Spectrosc 2012; 93: 95-9.
- Malathi R, Ganesan V. Environmental Benign Route in the Synthesis of Palladium Nanoparticles Using Leaves of Acalypha Indica L. Int J Pharm Bio Sci 2015; 6: 603–10.
- 257. Kannan SK, Sundrarajan M. Biosynthesis of Yttrium oxide nanoparticles using Acalypha indica leaf extract. *Bull Mater Sci* 2015; 38: 945–50
- Gnanasangeetha D, Thambavani DS. Biogenic Production of Zinc Oxide Nanoparticles Using Acalypha Indica. *Journal of Chemical, Biological and Physical Sciences* 2014; 4: 238-46.
- Shanthi S, Tharani SSN. Green Synthesis of Zirconium Dioxide (ZrO2) Nano Particles Using Acalypha Indica Leaf Extract. *IJEAS* 2016; 3: 23-5.
- Kumar BS. Study on Antimicrobial Effectiveness of Sliver Nano Coating Over Cotton Fabric through Green Approach. Int J Pharm Sci Res 2016; 7: 363-8.
- 261. Sakthivel P, Anitha P. Synthesis and Characterization of Silver Nanoparticles Using Acalypha Indica Leaf Extract and Its Anti-Inflammatory Activity against Human Blood Cells. *International Journal of Research in Pharmaceutical and Nano Sciences* 2016; 5: 26-34.
- Govindarajan M *et al.* Studies on effect of Acalypha indica L. (Euphorbiaceae) leaf extracts on the malarial vector, Anopheles stephensi Liston (Diptera:Culicidae). *Parasitol Res* 2008; 103: 691–95.
- Murugan T, Saranraj P. Antibacterial Activity of Various Solvent Extracts of the Indian Herbal Plant Acalypha indica against Human Pathogens Causing Nosocomial Infection. *Int j pharm biol sci arch* 2011; 2: 1473-78.
- 264. Jayakumari M *et al.* Antibacterial potential of Acalypha Indica against human pathogens. *Int J Curr Res* 2010; 1: 001-004.
- 265. Saranraj P et al. Antibacterial Potentiality of Ethanol and Ethyl Acetate Extract

of Acalypha indica against Human Pathogenic Bacteria. *J Ecobiotechnol* 2010; 2: 23-27.

- Gangadevi V et al. The Antibacterial Activity of Acalypha indica L. Indian J Sci Technol 2008; 1: 1-5.
- 267. Durga KR *et al.* Isolation of potential antibacterial and antioxidant compounds from Acalypha indica and Ocimum basilicum. *J Med Plants Res* 2009; 3: 703-6.
- Gupta R *et al.* Anti-tuberculosis activity of selected medicinal plants against multi- drug resistant Mycobacterium tuberculosis isolates. *Indian J Med Res* 2010; 131: 809-13.
- Kumarasamyraja D, Jeganathan NS. Green Synthesis of Silver Nanoparticles using Aqueous Extract of Acalypha Indica and Its Antimicrobial Activity. Int J Pharm Bio Sci 2013; 4: 469 – 76.
- Ponnusamy K et al. In vitro antifungal activity of indirubin isolated from a South Indian ethnomedicinal plant Wrightia tinctoria R. Br J Ethnopharmacol 2010; 132: 349-54.
- 271. Tariq AL *et al.* Antifungal *in vitro* Screening of Plant Acalypha indica Against Opportunist Clinical Pathogen Candida albicans. *World Journal of Zoology* 2016; 11: 141-7.
- 272. Sakthi SS *et al.* Pharmacological Screening of Datura Metel and Acalypha Indica for Its Antifungal Activity against Pathogenic Fungi. *International journal of pharmaceutical science and health care* 2011; 2: 15-29.
- 273. Somchit MN et al. In vitro antimicrobial activity of leaves of Acalypha indica Linn. (Euphorbiaceae). Afr J Microbiol Res 2010; 4: 2133-6.
- Kurandawad JM, Lakshman HC. Diversity of the Endophytic Fungi Isolated from Acalypha Indica Linn - A Promising Medicinal Plant. *IJSRP* 2014; 4: 1-7.

**GRAPHICAL ABSTRACT** 

- 275. Nanda A, Nayak BK. Endophytic fungal community study of varied aging leaves of Acalypha indica. *Der Pharmacia Lettre* 2015; 7: 250-4.
- Bagavan A *et al.* Larvicidal activity of saponin from Achyranthes aspera against Aedes aegypti and Culex quinquefasciatus (Diptera: Culicidae). *Parasitol Res* 2008; 103: 223–9.
- 277. Santhoshkumar T et al. Efficacy of adulticidal and larvicidal properties of botanical extracts against Haemaphysalis bispinosa, Hippobosca maculata, and Anopheles subpictus. Parasitol Res 2012; 111:1833–40.
- Inbaneson SJ. In vitro antiplasmodial effect of ethanolic extracts of coastal medicinal plants along Palk Strait against Plasmodium falciparum. Asian Pac J Trop Biomed 2012; 2:364-7.
- 279. Sahayaraj K, Shoba J. Toxic Effect of Tephrosia purpurea (Linn.) and Acalypha indica (Linn.) Aqueous Extracts Impact on the Mortality, Macromolecules, Intestinal Electrolytes and Detoxication Enzymes of Dysdercus cingulatus (Fab.). Asian Journal of Biochemistry 2012; 7: 112-22.
- Kumarasamyraja D *et al.* Phytochemical investigation and antimicrobial activity of Acalypha indica.Linn. *Novel Science International Journal of Pharmaceutical Science* 2012; 1: 313-6.
- Bolarinwa IF. Cyanogenic Glycosides in Plant Foods. Thesis submitted to The University of Leeds. 2013.
- 282. Rawat K *et al.* Processing techniques for reduction of cyanogenic glycosides from bamboo shoots. 10th world Bamboo Congress, Korea. 2015.
- Do QD et al. Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of Limnophila aromatic. J Food Drug Anal 2014; 22: 296-302.

#### **SUMMARY**

- And the second s
- Traditionally, A.indica formulations have worked against microbial infections, stomach ulcers, snake bites, pains, wounds, liver/kidney problems, and rheumatism etc.
- Scientific evidences proved that A.indica is a good source of anti-microbial, anti-diabetes, anti-inflammation, larvicidal, anti-oxidant, wound healing and venom neutralizing agent.
- Traditional and pharmaceutical properties of *A.indica* is due to the presence of phytochemical such as phenols, flavonoids, tannins, coumarins, alkaloids and their glycosides, and saponins.

**Cite this article:** Ravi S, Jyothi P, Shanmugam B, Subbaiah GV, Prasad SH, Reddy KS. Comprehensive Review on Traditional Knowledge, Phytochemistry and Pharmacological Properties of *Acalypha indica* L. Pharmacog Rev. 2021;15(30):134-85.